

Rearrangements to the JP1, JP and JP2 segments in the human T-cell rearranging gamma gene (TRG γ) locus

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In the human T-cell rearranging gamma (TRG γ) locus, five joining (J) segments have been identified: J1, J2 and three additional segments JP, JP1 and JP2. We report the sequence of the germline JP1 segment and compare it with the other human and mouse J γ segments. We also demonstrate that rearrangements to the three additional J γ segments can be identified by hybridization of the *Kpn*I digests to the J γ 1 probe pH60. Since rearrangements to J1 or J2 can be assigned, using the same pH60 probe, to one of the nine variable (V) γ genes known to rearrange [(1987) EMBO J. 6, 1945–1950], our results show that a unique probe can detect all the TRG γ rearrangements and be particularly useful for assessing the preferential usage of V γ and J γ segments in the TRG γ -expressing cells.

T-cell; Lymphocyte; Rearrangement; γ -Chain; Leukaemia; Lymphoma

1. INTRODUCTION

The human T-cell rearranging γ gene (TRG γ) has been identified [1,2] by homology with a mouse gene that has been shown to undergo rearrangement specifically in T-cells [3,4]. Human TRG γ genes have been mapped to chromosome 7 [5] at band 7p15 [2]. The TRG γ locus like other studied rearranging genes has variable (V) region genes, joining (J) and constant (C) region segments which join during the early stage of T-cell differentiation [6]. Two human constant-region genes have been identified [1,7–9] which are linked to each other at 16 kilobases [1,9]. 14 variable γ genes belonging to four subgroups [7,10,11] are located upstream of the two C γ genes (fig.1A). Nine V γ genes belong to subgroup I [10,11] whereas subgroups II, III and IV each consist of a single

gene, respectively designated V9, V10 and V11 [7,10,11]. Two joining gene segments, J γ 1 and J γ 2 were first identified upstream of C γ 1 and C γ 2 [7], as well as a J γ segment designated JP [10] upstream of J γ 1. More recently, two additional J γ gene segments, JP1 and JP2, have been located in the C γ 1 and C γ 2 loci, respectively [12,13] (fig.1). It is of interest to know the germline sequence of these segments to evaluate the diversity of the V-J junction. So far, germline sequences of four J γ segments have been published [7,10,13]. We now report the sequence of the germline JP1 and compare it with the other human and mouse J γ segments. We previously showed that rearrangements to J1 or J2 could be assigned, using the J γ 1 probe pH60 [7] to one of the nine V γ genes known to rearrange [11]. We now demonstrate that rearrangements to the additional J γ segments, JP1, JP and JP2 can be identified by hybridization of the *Kpn*I digests to the J γ 1 probe pH60. A unique probe can therefore detect all the TRG γ rearrangements.

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2. MATERIALS AND METHODS

Southern filter hybridization [14] was carried out with 10 μ g genomic DNA using nick-translated probes [15]. Conditions for hybridization, washing and monitoring have been described [10]. Nucleotide sequence analysis was carried out by the dideoxy chain-termination procedure [16] in M13 vectors [17].

2.1. Probes

The J γ probe, pH60, containing the 700 base pair (bp) *Hind*III-*Eco*RI from M13H60 [1] subcloned in pUC9, includes the J γ 1 segment [7], this J γ 1 probe cross-hybridizing with J γ 2 but not with the additional J γ segments. The V γ I probe, 1.1 kilobase (kb) *Sac*I V γ 3 fragment from λ SH4 [10], detects the nine V γ genes belonging to subgroup I [11]. The V γ II probe, a 400 bp *Pst*I-*Acc*I isolated from K20PR [7], contains V9 and detects the single gene belonging to subgroup II [10]. The V γ III probe, a 700 bp *Pst*I-*Eco*RI V γ 10 fragment from λ R12, contains the 5'-region of V γ 10 and detects the single gene belonging to subgroup III [11].

3. RESULTS AND DISCUSSION

3.1. Sequence of the germline JP1 segment

Restriction maps from two λ phage clones isolated from a MOLT4 library showed by comparison with published maps of the human TRG γ locus [9–11] that two rearrangements occurred, V2-JP1 on one chromosome and V2-JP2 on the other (Rabbitts, T.H. et al. and [18]). Restriction maps showed that JP1 and JP2 are in the proximity of a *Hind*III site which is absent from the MOLT4 clones, as a consequence of the rearrangement (fig.1B). In unrearranged clones these additional J segments are localized in a 2.4 kb and a 1.1 kb *Eco*RI fragment located respectively upstream of those containing JP and J2 [9]. A 2.4 kb *Eco*RI fragment containing the JP1 segment was obtained from λ R γ , a clone isolated from a phage library of the Burkitt's lymphoma cell line Raji [1]. *Hind*III-*Eco*RI subclones were sequenced from the *Hind*III site. The JP1 germline sequence which encodes 19 amino acid residues appears in fig.2. The conserved heptamer-nonamer sequences, proposed to be involved in the V-J join-

ing, are separated by a 12-base spacer at the 5'-end of the J segment, whereas a conserved splice site is found at its 3'-end. Therefore, JP1 appears to be a functional gene segment.

3.2. Comparison of the human and mouse J γ segments and duplication in the TRG γ locus

In fig.2, JP1 is compared to the other human and mouse J γ gene segments. Both JP1 and JP2 are 19 amino acids long, as are the mouse J γ gene segments. Their protein sequences share a homology of 57 and 52% respectively with those of the murine J1/J4 segments [4]. This homology between species is similar to and even higher than that existing between the mouse J1 and J2 [19,20] gene segments (which are 52% homologous).

As noted in the restriction maps and underlined by the repetition of characteristic *Kpn*I or *Xho*I sites (fig.1B), the human C γ locus underwent recent duplication. A homology of up to 98% has been found previously from nucleotide sequence comparison of the 2.1 kb *Hind*III fragments which contain J γ 1 and J γ 2 (fig.1B), respectively ([7] and Rabbitts, T.H. and Lefranc, M.-P., unpublished). Comparison of the restriction sites in regions encompassing the JP1 and JP2 segments in fig.1B reveals that these two regions also result from a recent duplication, although the homology is lower. Indeed, the JP1 and JP2 segments have a homology of 63% for the amino acid sequence and 80% for the nucleotide sequence compared to the 100% homology observed between the J1 and J2 segments (one allelic form of J1 has one silent nucleotide substitution [7] whereas the other [10] is perfectly homologous to J2). Strikingly, the region encompassing the JP segment has no equivalent in the C γ 2 locus [10] and this is of interest, since most of the T-cells expressing γ -chain seem to use to JP segment [22].

3.3. *Kpn*I digests and rearrangement assignments to JP1, JP, and JP2

When a V γ gene is rearranged to J1 or J2, it is possible to identify this gene by *Eco*RI, *Hind*III and *Bam*HI digestion and hybridization to the J γ 1 probe pH60 [11]. We now report that using the same probe, rearrangements to the additional J γ segments JP1, JP and JP2 can also be identified when DNAs are digested with the *Kpn*I restriction enzyme.

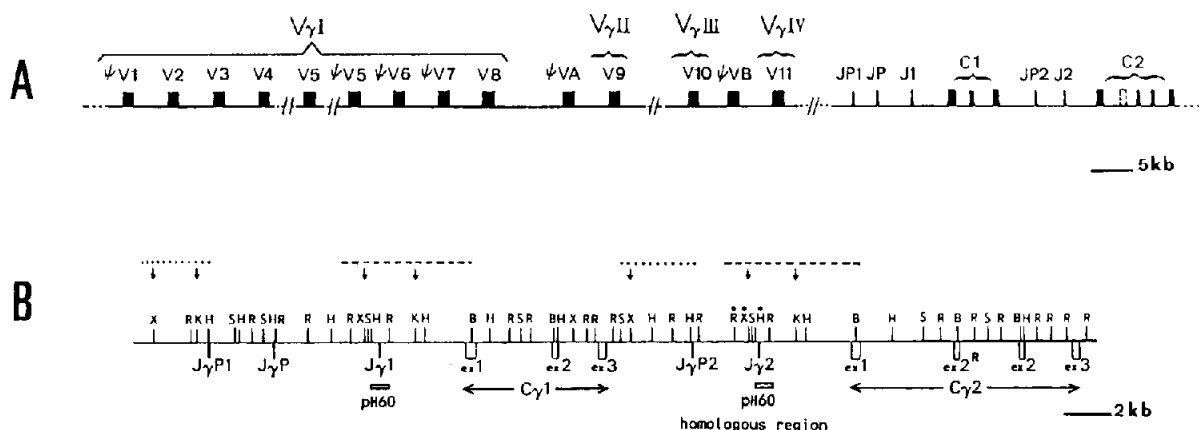


Fig.1. Organization of the human TRG γ locus. (A) Schematic representation of the human TRG γ locus [12]. For detailed maps, see [9–11] and panel B. (B) Restriction map of the human TRG γ constant region locus ([9] and this paper). Restriction sites: B, *Bam*HI; R, *Eco*RI; H, *Hind*III; K, *Kpn*I; S, *Sac*I; X, *Xho*I. Asterisks indicate polymorphic restriction sites [9]. Duplicated regions are shown by dashed lines.

The V γ I and V γ III subgroup genes have an internal *Kpn*I site [10,11]. According to the J segment to which the V γ I or V γ III gene is rearranged, *Kpn*I restriction fragments of different sizes can be detected: 1.8 kb (J1 or J2), 8.5 kb (JP1), 4.7 kb (JP2) (fig.3A). This is also shown in fig.4 where the 8.5 kb and 4.7 kb *Kpn*I bands detected in MOLT4 DNA (lane 1) and the 1.8 kb band observed in JM (lane 2) correspond respectively to the V2JP1, V2JP2 (MOLT4) and V8-J2 (JM) rear-

rangements [11,18]. Only one example of a V γ gene belonging to the V γ I or V γ III subgroup and involving JP has been described so far, a V2-JP out-of-frame rearrangement in the F8 cell line [10] which shows a 5.9 kb *Kpn*I band (fig.4, lane 3).

Two genes V9 and V11 (respectively, single members of the V γ II and V γ IV subgroups) have no internal *Kpn*I site [7,11] and display characteristic *Kpn*I bands when rearranged to J1 or J2: 7.5 kb for V9 and 6 kb for V11 (fig.3B). As

Ref.	HUMAN	N	Y	Y	K	K	L	F	G	S	G	T	T	L*	V	V	T					
(7)	J γ 1-----GAGT <u>TTTT</u> GATATGGACTGAATCAGCTGTGG	AAT	TAT	TAT	AAG	AAA	CTC	TTT	GGC	AGT	GGA	ACA	ACA	CTG	GTT	GTC	ACA	GGTAAGT				
		N	Y	Y	K	K	L	F	G	S	G	T	T	L	V	V	T					
(7)	J γ 2-----GAGT <u>TTTT</u> GATATGGACTGAATCAGCTGTGG	AAT	TAT	TAT	AAG	AAA	CTC	TTT	GGC	AGT	GGA	ACA	ACA	CTT	GTT	GTC	ACA	GGTAAGT				
		G	Q	E	L	G	K	K	I	K	V	F	G	P	G	T	K					
(10)	J γ P--GAGATTCTTATAAAGCGCTTCTCAGGTGGT	GGG	CAA	GAG	TTG	GGC	AAA	AAA	ATC	AAG	GTA	TTT	GGT	CCC	GGA	ACA	AAG	CTT	ATC	ATT	ACA	GGTAAGT
		T	T	G	W	F	K	I	F	A	E	G	T	K	L	I	V	T	S	P		
(this paper)	J γ P1-----GATTTTCTAGAAAGCTTAGACCGGTGTGAT	ACC	ACT	GGT	TGG	TTC	AAG	ATA	TTT	GCT	GAA	GGG	ACT	AAG	CTC	ATA	GTA	ACT	TCA	CCT	GGTAAGT	
		S	S	D	W	I	K	T	F	A	K	G	T	R	L	I	V	T	S	P		
(13)	J γ P2-----GATTTTGTAGAAAGCTTAGACCAAGTGTGAT	AGT	AGT	GAT	TGG	ATC	AAG	ACG	TTT	GCA	AAA	GGG	ACT	AGG	CTC	ATA	GTA	ACT	TCG	CCT	GGTAAGT	
	MOUSE	S	S	G	F	H	K	V	F	A	E	G	T	K	L	I	V	I	P	S		
(4)	J γ 1-----GATTTTGTAGAAAGCTGTGAGCACTGTGAT	AGC	TCG	GGC	TTT	CAC	AAG	GTA	TTT	GCA	GAA	GGA	ACA	AAG	CTC	ATA	GTA	ATT	CCC	TCC	GGTAAGT	
		S	G	F	H	K	V	F	A	E	G	T	K	L	I	V	I	P	S			
(4)	J γ 4-----GATTTTGTAGAAAGCTGTGAGCACTGTGAT	AGC	TCA	GGT	TTT	CAC	AAG	GTA	TTT	GCA	GAA	GGA	ACT	AAG	CTC	ATA	GTA	ATT	CCC	TCT	GGTAAGT	
		G	T	S	W	V	K	I	F	A	K	G	T	K	L	V	V	I	P	P		
(19,20)	J γ 2-----TGTTT <u>TTT</u> GTAGAAAGCGCTGAACAATGTGTCA	GGC	ACA	TCA	TGG	GTC	AAG	ATA	TTT	GCC	AAA	GGG	ACA	AAG	CTC	GTA	GTA	ATT	CCC	CCA	GGTAAGT	
		S	W	D	F	H	-	V	F	A	E	G	T	K	L	I	V	I	P	S		
(21)	J γ 3R-----AT	AGT	TGG	GAC	TTT	CAC	-AG	GTA	TTT	GCA	GAA	GGA	ACT	AAG	CTC	ATA	GTA	ATT	CCT	TCT	GGTAAGT	

Fig.2. Sequence of the germline JP1 segment and comparison with the known human and mouse J γ gene segments. Heptamer and nonamer sequences are underlined. Dashed lines indicate *Hind*III sites. The splicing sites are denoted by arrows. An asterisk indicates the nucleotide which can be either G or T (silent substitution) in the J γ 1 sequence [7,10].

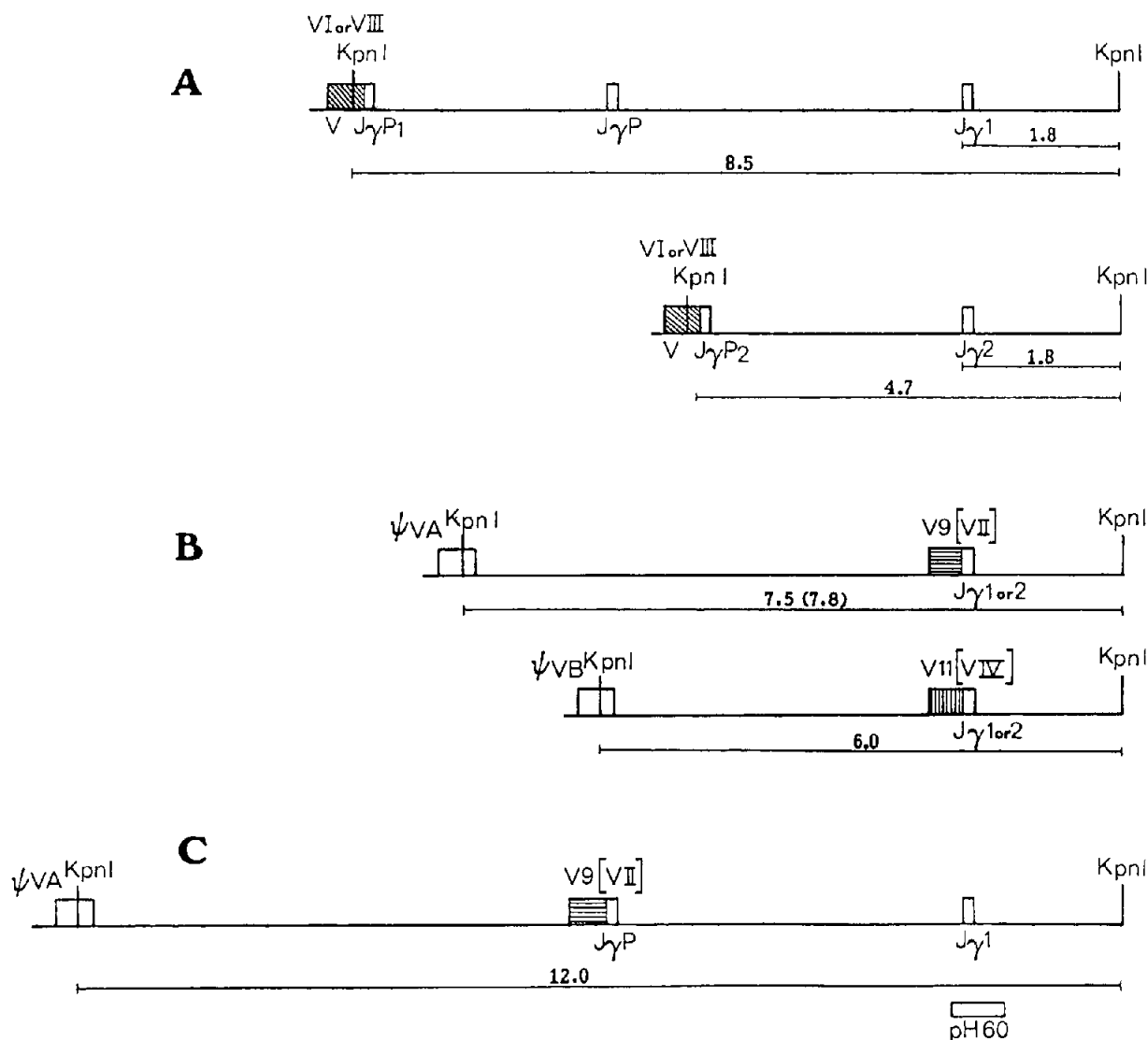


Fig.3. *KpnI* restriction fragment sizes when $V\gamma$ genes are rearranged to the different $J\gamma$ gene segments. (A) Rearrangements of the $V\gamma$ I or $V\gamma$ III subgroup genes to the $J\gamma$ 1, $J\gamma$ 2, JP1 or JP2 segments. The $V\gamma$ I subgroup genes which rearrange are V2, V3, V4, V5, ψ V7 and V8 [10,11]. V10 is the single member of the $V\gamma$ III subgroup [11]. (B) Rearrangement of the V9 and V11 genes (respectively, single members of the $V\gamma$ II and $V\gamma$ IV subgroups) to the $J\gamma$ 1 or $J\gamma$ 2 gene segments [7,10,11]. (C) V9-JP rearrangement observed in TRG γ^+ cells [22]. A *Hind*III restriction fragment length polymorphism (RFLP) due to a 300 bp insertion/deletion exists for one of the fragments located between ψ VA and V γ 9 [11], which explains the *KpnI* RFLP (7.5 or 7.8 kb *KpnI* bands) mentioned in panel B. The *KpnI* restriction fragment sizes have been deduced from cloned DNA fragments [9–11].

an example, the 6 kb *KpnI* band detected in JM (fig.4, lane 2) corresponds to a V11-J1 rearrangement ([11] and unpublished).

These *KpnI* bands are detected, in addition to the 16 kb (J2) and 9 kb (J1) germline bands when

thymus or peripheral T-lymphocytes are digested with *KpnI* and hybridized to pH60 (fig.4, lane 4) (our data and [23]). All these bands have been identified in T-cell clones displaying one or the other of these rearrangements. Interestingly, CD3 $^+$

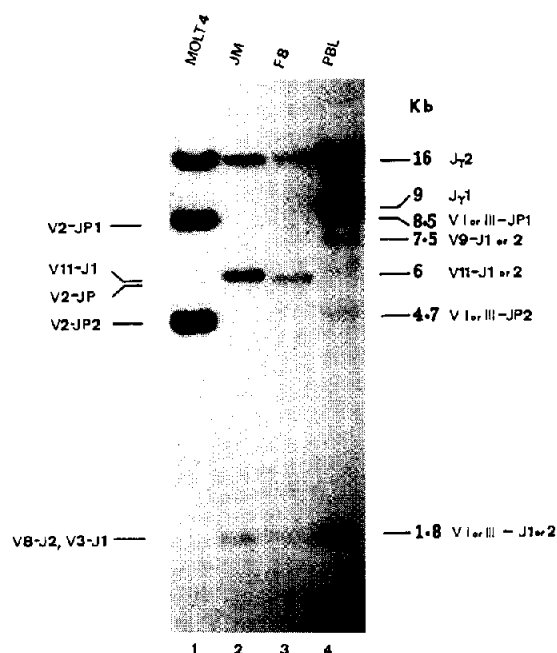


Fig.4. Southern hybridization of pH60 to *KpnI*-digested DNA from T-cells (lanes 1–3) and peripheral blood leukocytes (PBL) (lane 4). Lanes: 1, MOLT4; 2, JM; 3, F8; 4, PBL of a normal donor. On the left are indicated rearrangements observed in MOLT4: V2JP1 and V2JP2 (see text); JM: V11JP1, V8J2 ([11] and unpublished); F8: V2JP, V3J1 [10]. The sizes of the rearrangements observed in PBL and their assignment are given on the right.

TCR $\alpha\beta^+$ clones expressing γ and recognized by the monoclonal antibody anti-Ti γ A (TRG γ^+ cells) [24] display a productive V9-JP rearrangement identified by a characteristic 12 kb *KpnI* band (fig.3C) [22]. Altogether these results show that *KpnI* digests hybridized to pH60 allow the identification of rearrangements occurring to the additional J γ segments (JP1, JP or JP2). Moreover, the precise assignment of the V γ genes involved in these rearrangements can be made using specific V γ probes (see section 2). Table 1 shows the *EcoRI* restriction fragments allowing the identification of the V γ I genes rearranged to JP1 or JP2. A V9-JP rearrangement can be confirmed by hybridization of an *EcoRI* digest to the V γ II probe (a 2 kb *EcoRI* rearranged band instead of the 5.2 kb germline one) [11,22] and a V10-JP1 rearrangement can be confirmed by hybridization of a *HindIII* digest to the V γ III probe (a 4.2 kb *HindIII*

Table 1

Assignment of the V γ I genes joined to J γ P1 or J γ P2 (*EcoRI* digests, hybridized to the V γ I probe)

	JP1	JP2
V2	2.1 (18.5)	0.5 (14.5)
V3	6.6	5.0
V4	2.1 (28)	0.5 (24)
V5	3.4	1.8
V7	4.3	2.7
V8	5.4	3.8

Sizes of the *EcoRI* rearranged bands are in kilobases (kb). Sizes of the *BamHI* bands for the V2 and V4 rearrangements are given in parentheses

rearranged band instead of the 3.5 kb germline band) [11].

4. CONCLUSION

We reported the sequence of the human germline JP1 segment and compared it with the other human and mouse J γ gene segments. This segment, as well as the other additional J γ segments, JP and JP2, have been shown to be used in T-cell clones. The function of the TRG γ -expressing cells remains unclear. It is therefore of interest to characterize fully the V-J rearrangements undergone by these cells. We previously showed that rearrangements to J1 or J2 could be assigned to one of the nine V γ genes known to rearrange [11]. Here, we demonstrated that *KpnI* digests hybridized to the J γ I probe, pH60, allow a characterization of the rearrangements occurring to the JP1, JP or JP2 segments. Together these results show that a unique probe, pH60, can detect all rearrangements in the TRG γ locus whatever the J γ segment involved in the rearrangements. T-cells displaying only one (or none) TRG γ rearrangement with *EcoRI*, *HindIII* and *BamHI* [11] should be digested with *KpnI* in order to detect rearrangements to the additional J γ segments. In particular, the V9-JP rearrangement frequently observed in TRG γ^+ cells [22] is detected with the *KpnI* digest (the *BamHI* rearranged band has virtually the same size as the germline band and the *EcoRI* and *HindIII* bands are in germline configuration). Moreover, it is also possible to identify (or confirm) the V gene rearranged to an addi-

tional J γ segment by using specific V γ I, V γ II or V γ III probes. Such precise determination of the V γ -J γ rearrangements should help in elucidating the function of the γ protein by assessing the preferential usage of given V and J segments in the TRG γ -expressing cells.

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