

The BChlc/e-binding polypeptides from chlorosomes of green photosynthetic bacteria

Regula Wagner-Huber, René Brunisholz, Gerhard Frank and Herbert Zuber

Institut für Molekularbiologie und Biophysik, ETH-Hönggerberg, CH-8093 Zürich, Switzerland

Received 31 August 1988

A 6.3 kDa polypeptide has been isolated from chlorosomes of the green photosynthetic bacterium *Pelodictyon luteolum*, and its complete amino acid sequence has been determined. It exhibits an overall homology of 30% to the BChlc-binding protein of *Chloroflexus aurantiacus*. Preliminary results from the N-terminal sequence analyses of the analogous polypeptides isolated from *Chlorobium limicola*, *Prosthecochloris aestuarii* and *Chlorobium phaeovibrioides* revealed a highly conserved sequence. This protein is suggested to be the BChlc/e-binding polypeptide in the family of the Chlorobiaceae.

Chlorosome; BChlc/e-binding polypeptide;
(*Pelodictyon luteolum*, *Chlorobium limicola*, *Prosthecochloris aestuarii*, *Chlorobium phaeovibrioides*)

1. INTRODUCTION

Green photosynthetic bacteria, the sub-order of the Chlorobiineae, consist of two families: the Chloroflexaceae and the Chlorobiaceae. The best characterized species of the Chloroflexaceae is *Chloroflexus aurantiacus*. Its photosynthetic apparatus contains two different light harvesting complexes: an intramembranously located BChla-binding complex B 806–866 consisting of an α/β pair of polypeptides [1,2], and an additional antenna containing BChlc absorbing at 740 nm. The latter is organized in the so-called chlorosomes, surface-located oblong vesicles [3–6]. Freeze fracture electron microscopy of chlorosomes by Staehelin et al. [7] showed rod-like three dimensional structures. SDS-polyacrylamide gel electrophoresis of chlorosomes revealed three major polypeptides (18, 11 and 3.7 kDa) and a fourth polypeptide (5.8 kDa) in minor amounts [8].

Correspondence address: H. Zuber, Institut für Molekularbiologie und Biophysik, ETH-Hönggerberg, 8093 Zürich, Switzerland

Abbreviations: PTH, phenylthiohydantoin; TFA, trifluoroacetic acid

Crosslinking as well as proteolytic digestion experiments led to the conclusion that the BChlc is associated with the 3.7 kDa polypeptide. This polypeptide was isolated from purified chlorosomes and its amino acid sequence was determined [9], yielding a true molecular mass of 5.6 kDa. Seven Asn/Gln specifically arranged on one side of the presumptive α -helix were suggested to bind the BChlc [9].

In the family of the Chlorobiaceae far less is known about the photosynthetic apparatus. There are some distinct differences between the Chloroflexaceae and the Chlorobiaceae. So far, no intracytoplasmatic core antenna complex has been found in cells of Chlorobiaceae. However, an additional water-soluble BChla-binding protein, located between the chlorosome and the reaction center, has been isolated and crystallized [10]. The existence of a further BChla-containing complex absorbing at 794 nm located in the chlorosome membrane of *Chlorobium limicola* has been reported by Gerola et al. [11]. From chlorosomes of this bacterium a 74 amino acid residues polypeptide has been isolated and sequenced [12,13]. Although the homology between this polypeptide and the BChlc-binding protein of *C.*

eluted in front of the 6.3 kDa polypeptide according to its higher molecular weight. These polypeptides were dialyzed and lyophilized and then subjected to the automated Edman degradation procedure. The primary structure analysis of the polypeptide from *C. limicola* yielded the already established sequence published by Gerola et al. [12,13] (fig.3). An analogous polypeptide has been isolated from cells of *P. luteolum* (fig.3). However, it is remarkable that it was not possible to isolate this polypeptide from purified chlorosomes of *P. luteolum* under exactly the same conditions.

4. DISCUSSION

The 6.3 kDa polypeptide from *P. luteolum*, as well as the equivalent polypeptides isolated from *C. limicola*, *P. aestuarii* and *C. phaeovibrioides*, show a homology of 30% to the BChlc-binding polypeptide of *C. aurantiacus*. This is rather high considering that there are significant differences between the Chlorobiaceae and Chloroflexaceae with respect to the photosynthetic apparatus. The Gln and Asn residues which have been suggested to coordinate to the central Mg-atom of the bacteriochlorophyll *c* in *C. aurantiacus* [9] are replaced by other amino acids in the 6.3 kDa polypeptide of the Chlorobiaceae. However, in three cases (fig.2, positions 23, 27, 34) the amide group is substituted by a carboxyl group which possibly can bind BChlc similarly via the oxygen atom. Remarkable is the highly conserved region around the His (position 25), which also is a possible ligand to the Mg-atom. Another strictly homologous region is found from residue 43 to 48.

The homology of the 6.3 kDa polypeptide within the members of the Chlorobiaceae is extremely high (fig.2). Besides the close relationship, there must be an additional reason for this fact. Most probably this 6.3 kDa polypeptide has a very important functional role that allows practically no substitutions in the amino acid sequence. It is notable that only the polypeptide of *C. phaeovibrioides* exhibits some structural differences to the three other polypeptides, even in the homologous clusters (fig.2, positions 27, 45). This may be explained by the fact that this bacterium belongs to the brown-green Chloro-

biaceae species containing BChle in the chlorosome antenna.

The 74 amino acid polypeptide of *C. limicola* described by Gerola et al. [12,13] as BChlc-binding polypeptide (fig.3, line 2) exhibits only 17.6% homology to the BChlc-binding protein of *C. aurantiacus* (fig.3). Most of the homologous amino acids are alanines and other residues which seem to be of less functional importance. The point that we could only isolate the analogous protein from cells of *P. luteolum* but not from chlorosomes is possibly due to different preparation procedures for the chlorosomes. The miranol treatment probably induced a release of some proteins including the 74 amino acid polypeptide which are not released by the method of Gerola et al. [11]. Thus the 74 amino acid polypeptide is presumably not part of the internal B 740 antenna complex of the chlorosome. It could be bound to chlorosome surface or membrane surface. On the other hand it is reasonable to assume that the 6.3 kDa polypeptide from *P. luteolum*, as well as the equivalent proteins from *C. limicola*, *P. aestuarii* and *C. phaeovibrioides* are the BChlc/e-binding polypeptides in the Chlorobiaceae.

This is suggested by the following observations: (1) the 6.3 kDa polypeptide is found in isolated chlorosomes and seems to be located within the chlorosome; (2) the sequence homology of the 6.3 kDa polypeptides of several Chlorobiaceae is 30% to the BChlc-binding polypeptide of *C. aurantiacus*; (3) the 74 amino acid polypeptide is also missing in the chlorosome of *C. aurantiacus*.

Acknowledgements: We are indebted to Professor Norbert Pfennig, who kindly provided the cultures of *P. luteolum*, *C. limicola*, *P. aestuarii* and *C. phaeovibrioides* and taught us how to grow them. We wish to thank Mrs Monika Wirth (amino acid analysis) for her skilled and excellent technical assistance. This work was supported by the Eidgenössische Technische Hochschule, Kredit Unterricht und Forschung and by the Swiss National Science Foundation (project no. 3.207.0.85).

REFERENCES

- [1] Wechsler, T., Brunisholz, R., Frank, G. and Zuber, H. (1987) FEBS Lett. 210, 189–194.
- [2] Wechsler, T., Brunisholz, R., Suter, F., Fuller, R.C. and Zuber, H. (1985) FEBS Lett. 191, 34–38.
- [3] Pierson, B.K. and Castenholz, R.W. (1974) Arch. Microbiol. 100, 5–24.

- [4] Pierson, B.K. and Castenholz, R.W. (1978) in: *The Photosynthetic Bacteria* (Clayton, R.K. and Sistrom, W.R. eds) pp.179–197, Plenum, New York.
- [5] Schmidt, K., Maarzahl, M. and Mayer, F. (1980) *Arch. Microbiol.* 124, 21–31.
- [6] Sprague, S.G., Staehelin, A., Di Bartolomeis, M.J. and Fuller, R.C. (1981) *J. Bacteriol.* 147, 1032–1039.
- [7] Staehelin, L.A., Golecki, J.R., Fuller, R.C. and Drews, G. (1978) *Arch. Microbiol.* 119, 269–277.
- [8] Feick, R.G. and Fuller, R.C. (1984) *Biochemistry* 23, 3693–3700.
- [9] Wechsler, T., Suter, F., Fuller, R.C. and Zuber, H. (1985) *FEBS Lett.* 181, 173–178.
- [10] Matthews, B.W., Fenna, R.E., Bolognesi, M.C., Schmidt, M.F. and Olson, J.M. (1979) *J. Mol. Biol.* 131, 259–285.
- [11] Gerola, P. and Olson, J.M. (1986) *Biochim. Biophys. Acta* 848, 69–76.
- [12] Gerola, P.D., Hojrup, P., Knudsen, J., Roepstorff, P. and Olson, J.M. (1988) in: *Green Photosynthetic Bacteria* (Olson, J.M., Ormerod, J.G., Ames, J., Stackebrandt, E. and Trüper, H.G. eds) pp.43–52, Plenum, New York.
- [13] Gerola, P.D., Hojrup, P. and Olson, J.M. (1988) in: *Photosynthetic Light-Harvesting Systems Organization and Function* (Scheer, H. and Schneider, S. eds) pp.129–139, Walter de Gruyter, Berlin.
- [14] Zuber, H. (1988) in: *Green Photosynthetic Bacteria* (Olson, J.M., Ormerod, J.G., Ames, J., Stackebrandt, E. and Trüper, H.G. eds) pp.53–55, Plenum, New York.
- [15] Pfennig, N. and Trüper, H.G. (1981) in: *The Prokaryotes* (Starr, M.P., Stolp, H., Trüper, H.G., Balows, A. and Schlegel, H.G. eds) pp.279–289, Springer, Berlin.
- [16] Fowler, V.J., Pfennig, N., Schubert, W. and Stackebrandt, W. (1984) *Arch. Microbiol.* 139, 382–387.
- [17] Rümeli, R., Wirth, M., Suter, F. and Zuber, H. (1987) *Biol. Chem. Hoppe-Seyler* 368, 1–9.
- [18] Akabori, S., Ohno, K. and Narita, K. (1952) *Bull. Chem. Soc. Jpn.* 25, 905–915.
- [19] Ambler, R.P. (1967) *Methods Enzymol.* 11, 155–166.
- [20] Frank, G. (1988) in: *Methods in Protein Sequence Analysis. Proceedings of the 7th Int. Conf.*, Springer, Berlin, in press.