

The cysteines in position 1 and 86 of rat interferon- α_1 are indispensable for antiviral activity

Remco A. Spanjaard, John A.J. van Himbergen and Jan van Duin

Department of Biochemistry, University of Leiden, Leiden, The Netherlands

Received 28 March 1989

The human, bovine, murine and rat interferon (IFN)- α families contain 4 conserved cysteines located at positions 1, 29, 99 and 139 that are involved in disulfide bridges. Rat and murine IFN- α subspecies carry a fifth Cys (Cys-86) which is not conserved in bovine and human IFN- α subspecies except for human IFN- α_1 . Changing Cys-86 in rat IFN- α_1 into Ser or Tyr virtually abolished antiviral activity. As shown by others, the substitution of Cys-86 to Ser in human IFN- α_1 had no pronounced effect on activity. This suggests that in contrast to human and bovine IFN- α , Cys-86 in rodent IFN- α plays a crucial role in receptor binding. Changing Cys-1 to Gly in rat IFN- α_1 also destroyed activity, in agreement with results obtained in the human IFN- α_1 system.

Interferon- α_1 ; Antiviral activity; Disulfide bridge; Interferon receptor; (Rat)

1. INTRODUCTION

The interferon (IFN)- α genes constitute a true multigene family [1] that code for proteins that show on average 60% interspecies homology [2]. For instance, all human [1], bovine [2], murine [3–5] and rat IFN- α [6] proteins carry four cysteines at position 1, 29, 99 and 139. Cys-1 and 99 as well as Cys-29 and 139 are connected by disulfide bonds (Cys-1 is the first amino acid of the mature IFN- α peptide) [7].

Interestingly, all murine IFN- α genes and the only sequenced rat IFN- α gene (α_1) contain a fifth Cys at position 86. However, all bovine IFN- α proteins carry Arg or Gly and all human IFN- α subspecies (about 15) with one exception carry Ser or Tyr at this position. The exception is human IFN- α_1 which also contains a Cys at position 86 [8]. Here, we assess the importance of Cys-86 in rat IFN- α_1 for antiviral activity by substituting this amino acid by Tyr or Ser. In addition, we changed

Cys-1 to Gly. All mutants are severely affected in biological activity. Comparison with mutations obtained by Beilharz et al. [9] in the human IFN- α_1 protein suggests that Cys-86 is essential for recognition of the receptor protein in the rat but not in humans.

2. MATERIALS AND METHODS

2.1. Bacterial strains and growth conditions

Escherichia coli K-12 strain M5219 was used in all experiments [10]. This strain harbors a defective and nonexcisable λ prophage expressing the *clts857* gene and the gene for the antitermination factor *N*. Bacteria were grown in LC medium which contained (per l) 10 g tryptone, 5 g yeast extract (Difco), 8 g NaCl and 5 ml of 1 M Tris (pH 7.3). Cells were grown at 28°C to an A_{650} value of 0.2 and then induced at 42°C for 120 min.

2.2. Construction of mutants of rat IFN- α_1

The sequence encoding mature rat IFN- α_1 was cloned in *E. coli* expression vector pIF.D [11], which is derived from pPLc236 [10]. pIF.D contains a synthetic ribosomal binding site that adds a methionine to the N-terminus of mature IFN- α_1 . Clones pIF.Ser-86 and pIF.Tyr-86 were obtained by site-directed mutagenesis of clone pIF.D essentially as described by Kunkel [12] and a mixture of two mutagenic heptadecamers. Clone pIF.Gly-1 was obtained as a gene fusion where the first

Correspondence address: J. van Duin, Department of Biochemistry, Gorlaeus Laboratory, University of Leiden, Einsteinweg 5, 2333 CC Leiden, The Netherlands

four nucleotides of the bacteriophage MS2 coat gene [13] are connected to the second nucleotide of the mature IFN- α_1 sequence. This yields a product identical to our wild-type except for the Cys-1 \rightarrow Gly substitution.

2.3. Rat IFN- α_1 activity assay

Bacterial lysates were prepared by suspending the bacteria in 6 M guanidine HCl, 1% β -mercaptoethanol. After centrifugation, the supernatant was isolated and tested for antiviral activity on RATEC cells as in [14]. The protein yields from the different plasmids were compared in Western blots [15] but did not differ significantly from one another (not shown).

3. RESULTS AND DISCUSSION

The mature rat IFN- α_1 sequence containing the 5 Cys residues is schematically represented in fig.1a. Cys-86 was mutated into either Ser (pIF.Ser-86) or Tyr (pIF.Tyr-86) (fig.1b). These clones were assayed for antiviral activity on rat cells and the results are listed in table 1. IFN.Ser-86 has only 10% of the activity of the wild-type protein and IFN-Tyr-86 even less, if any, antiviral activity. The Ser-86 mutation in the human IFN- α_1 gene still yields 40% activity [9]. Since Cys-86 is

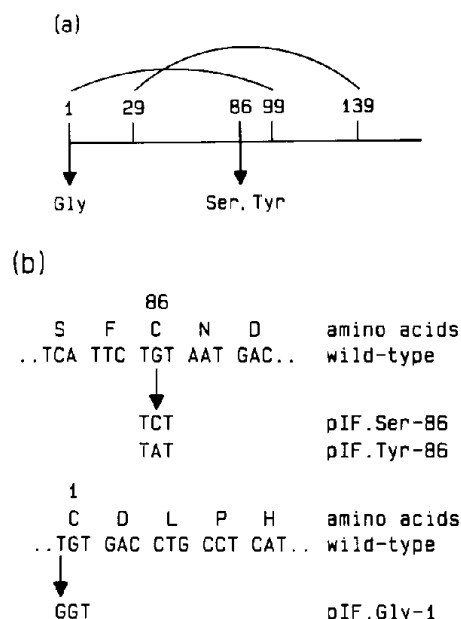


Fig.1. (a) Schematic representation of mature rat IFN- α_1 . The position of the cysteine residues in the protein is shown by their amino acid number. The proposed disulfide bonds are indicated by connecting lines. Mutations discussed in section 3 are shown. (b) Relevant nucleotide and amino acid sequence of clones pIF.Ser-86, pIF.Tyr-86 and pIF.Gly-1.

Table 1

Specific activities of rat and human mutant IFN- α_1

	Human IFN- α_1 ^a (%)	Rat IFN- α_1 ^b (%)	Rat IFN- α_1 activity (U/l)
Wild-type	100	100	8.4×10^5
Ser-86	39	10	8.1×10^4
Tyr-86	n.c.	≤ 1	$\leq 9.8 \times 10^3$
Ser-1	5	n.c.	—
Gly-1	n.c.	2	1.6×10^4

^a Tested on human cells (HEp-2) [9]. ^b Tested on rat cells (RATEC). See section 2 and [14]. n.c., not constructed

not known to be involved in a disulfide bond, this indicates that this residue might be crucial for direct recognition of the rat IFN- α_1 cell-surface receptor, whereas in humans the receptor-protein interaction appears less stringent at this particular position. These results agree with those obtained by Weber et al. [17] who used hybrids between human IFN- α_1 and human IFN- α_2 to locate regions that are important for antiviral activity on human, murine and bovine cells. Hybrids that contained Tyr at position 86 showed about 20-fold reduced specific activity on murine cells, whereas activity on human and bovine cells was unchanged. The fact that amino acid 86 is variable in human and bovine IFN- α also suggests this residue to be relatively unimportant.

We have also substituted Cys-1 for Gly-1 (fig.1b). In agreement with the findings for similar changes in human IFN- α_1 [9], this mutation almost completely inactivated rat IFN- α_1 (table 1).

Acknowledgements: We are indebted to Dr P.H. van der Meide for performing the bioassay and for critically reading the manuscript. This research was supported by Stichting Technische Wetenschappen Grant LBI37.0496 to R.A.S.

REFERENCES

- [1] Henco, K., Brosius, J., Fujisawa, J.-I., Haynes, J.R., Hochstadt, J., Kovacic, T., Pasek, M., Schamböck, A., Schmid, J., Todokoro, K., Wälchli, M., Nagata, S. and Weissman, C. (1985) *J. Mol. Biol.* 185, 227–260.
- [2] Weissmann, C. and Weber, H. (1986) *Prog. Nucleic Acid Res. Mol. Biol.* 33, 251–300.
- [3] Shaw, G.D., Boll, W., Taira, H., Mantel, N. and Weissmann, C. (1983) *Nucleic Acids Res.* 11, 555–573.
- [4] Zwarthoff, E.C., Mooren, A.T.A. and Trapman, J. (1985) *Nucleic Acids Res.* 13, 791–804.

- [5] Kelley, K.A. and Pitha, P.M. (1985) *Nucleic Acids Res.* 13, 805–823.
- [6] Dijkema, R., Pouwels, P., De Reus, A. and Schellekens, H. (1984) *Nucleic Acids Res.* 12, 1227–1242.
- [7] Wetzel, R. (1981) *Nature* 289, 606–607.
- [8] Mantel, N., Schwarzstein, M., Streuli, M., Panem, S., Nagata, S. and Weissmann, C. (1980) *Gene* 10, 1–10.
- [9] Beilharz, M.W., Nisbet, I.T., Tymms, M.J., Hertzog, P.J. and Linnane, A.W. (1986) *J. Interferon Res.* 6, 677–685.
- [10] Remaut, E., Stanssens, P. and Fiers, W. (1981) *Gene* 15, 81–93.
- [11] Spanjaard, R.A., Van Dijk, M.C.M., Turion, A.J. and Van Duin, J. (1989) *Gene*, in press.
- [12] Kunkel, T.A. (1985) *Proc. Natl. Acad. Sci. USA* 82, 488–492.
- [13] Fiers, W., Contreras, R., Deurinck, F., Haegeman, G., Iserentant, D., Merregaert, J., Min Jou, W., Molemans, F., Raeymakers, A., Van den Berghe, G., Volkaert, G. and Ysebaert, M. (1976) *Nature* 260, 500–507.
- [14] Van der Meide, P.H., Dijkema, R., Caspers, M., Vijverberg, K. and Schellekens, H. (1986) *Methods Enzymol.* 119, 230–236.
- [15] Towbin, H., Staehelin, T. and Gordon, J. (1979) *Proc. Natl. Acad. Sci. USA* 76, 4350–4354.
- [16] Mark, D.F., Lu, S.D., Creasy, A.A., Yamamoto, R. and Lin, L.S. (1984) *Proc. Natl. Acad. Sci. USA* 81, 5662–5666.
- [17] Weber, H., Valenzuela, D., Lujber, G., Gubler, M. and Weissmann, C. (1987) *EMBO J.* 6, 591–598.