

Cloning and sequence analysis of pituitary cDNA encoding the β -subunit of *Xenopus* proteasome

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The proteasome is a multicatalytic proteinase composed of a number of non-identical subunits. A *Xenopus* pituitary cDNA was isolated and found to code for the β -subunit of proteasome. The amino acid sequence deduced from the open reading frame consisted of 215 amino acid residues with a calculated molecular weight of 23 979. A comparative structural analysis indicated that the proteasome subunits can be divided into two groups with the same evolutionary origin. One group consists of subunits with an N-terminally blocked residue and includes components C2, C3, C8 and C9, while the second group of non-blocked proteins includes component C5 and the β -subunit.

Proteasome; Multicatalytic proteinase; β -Subunit; cDNA sequence; *Xenopus* pituitary

1. INTRODUCTION

The proteasome is a symmetrical, cylinder-shaped particle which is present in both nucleus and cytoplasm of a variety of eukaryotic cells [1]. This particle with a sedimentation coefficient of approximately 20 S (molecular mass about 750 kDa) is composed of a characteristic set of at least 13 non-identical polypeptide subunits with molecular masses between 20 and 35 kDa [2–4]. The individual subunits may each have a unique catalytic property, i.e. cleavage of peptide bonds on the carboxyl side of basic, hydrophobic or acidic amino acid residues [5]. The cellular function of the proteasome is, however, still unknown although recent evidence suggests that this highly-conserved particle is a multifunctional proteinase involved in the degradation of proteins (and possibly also RNA) in a non-lysosomal pathway [6–8].

For the elucidation of the role of the proteasome it is important to determine the primary structures of all subunits of this proteinase complex. Recently, cloning and sequence analyses of cDNAs revealed the structures of a number of subunits of human, rat, *Drosophila*, yeast and archaebacterial proteasomes [9–17]. Here we report the cloning of a *Xenopus* pituitary cDNA encoding another component, the β -subunit, of the proteasome.

2. MATERIALS AND METHODS

A cDNA library in the vector λ ZAP-II (Stratagene) was constructed with RNA isolated from intermediate pituitary glands of black-background-adapted *Xenopus laevis*. From this library single plaques were picked at random and the recombinant pBluescript SK-phagemids were rescued from the bacteriophage (λ ZAP) clones by in vivo excision according to the instructions of the manufacturer. Sequencing on both strands and with pBluescript subclones was performed with single-stranded and double-stranded DNA using T7 DNA polymerase and the dideoxy chain termination method [18]. For Northern blot analysis, poly(A)⁺ RNA was isolated from *Xenopus* liver with guanidineisothiocyanate and oligo(dT) cellulose, and RNA samples (10 μ g) were fractionated on a 1.2% agarose gel in 2.2 M formaldehyde. After transfer [19], the filter was hybridized in standard hybridization buffer [20] in the presence of 50% formamide with insert DNA of cDNA clone X7355 ³²P-labelled by random priming to 5 \times 10⁸ cpm/ μ g DNA. Homology searches were performed with the CAOS/CAMMSA computer facility of the University of Nijmegen which includes the sequence alignment program Clustal [21].

3. RESULTS AND DISCUSSION

3.1. Cloning and sequence analysis of pituitary cDNA encoding the β -subunit of *Xenopus* proteasome

The prohormone proopiomelanocortin (POMC) is expressed to very high levels in the melanotrope cells of the intermediate pituitary of *Xenopus* adapted to a black background [22]. During our search for proteins co-expressed with POMC we isolated from a *Xenopus* intermediate pituitary cDNA library a cDNA clone (X7355) with an insert of about 900 bp (which includes a poly(A)-tail of 35 nucleotides) (Fig. 1). A putative polyadenylation signal (ATTAAA) is located 29 nucleotides upstream from the polyadenylation site. The longest open-reading frame of clone X7355 codes for a protein of 215 amino acids, giving a protein with a

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5'---GAATTCGGTGAATCTGTGGCCC 22
GGGGGACCGCTCCTGGAGAGCTGCACTGTTTCCCGGGGCGGGTCCGGTCAGACACACACTGAACCCCT 90
1      10      20
Met Val Thr Gly Thr Ser Val Leu Gly Val Lys Phe Asp Gly Gly Val Ile Ile Ala Ala
ATG GTG ACG GGA ACC TCT GTG CTG GGA GTG AAA TTT GAT GGG GGA GTG ATT AAT GCG GCG 150
30      40
Asp Met Leu Gly Ser Tyr Gly Ser Leu Ala Arg Phe Arg Asn Ile Ser Arg Ile Met Lys
GAC ATG CTG GGC TCC TAT GGG TCG CTG GCG AGA TTC CGC AAC ATC TCC AGG ATA ATG AAA 210
50      60
Val Asn Glu Asn Thr Ile Leu Gly Ala Ser Gly Asp Tyr Ala Asp Tyr Gln Tyr Leu Lys
GTG AAT GAG AAC ACT ATT CTG GGG GCA TCT GGA GAC TAC GCG GAT TAT CAG TAT CTC AAG 270
70      80
Gln Val Ile Asp Gln Met Val Ile Asp Glu Glu Leu Val Gly Asp Gly His Asn Tyr Ser
CAG GTC ATT GAT CAG ATG GTC ATC GAT GAG GAG CTG GTG GGG GAC GGA CAC AAT TAC AAG 330
90      100
Pro Lys Ala Ile His Ser Trp Leu Thr Arg Val Met Tyr Asn Arg Arg Ser Lys Met Asn
CCA AAG GCC ATT CAC TCA TGG CTG ACC CGG GTC ATG TAC AAC CGG AGG AGC AAG ATG AAC 390
110     120
Pro Leu Trp Asn Thr Val Val Ile Gly Gly Phe Tyr Asn Gly Glu Ser Phe Leu Gly Tyr
CCC CTG TGG AAC ACC GTC GTT ATT GGG GGT TTC TAT AAT GGA GAG AGT TTC CTT GGA TAC 450
130     140
Val Asp Lys Leu Gly Val Ala Tyr Glu Ala Pro Thr Ile Ala Thr Gly Phe Gly Ala Tyr
GTG GAC AAA CTG GGC GTG GCC TAT GAA GCA CCA ACC ATT GCT ACG GCG TTT GCG GCA TAC 510
150     160
Leu Ala Gln Pro Leu Leu Arg Glu Val Thr Glu Asn Lys Ala Thr Leu Ser Lys Glu Glu
CTG GCA CAG CCG CTG CTG AGA GAA GTA ACC GAG AAT AAA GCG ACT CTG AGT AAG GAA GAG 570
170     180
Ala Arg Gln Leu Val Asp Arg Cys Met Lys Val Leu Tyr Tyr Arg Asp Ala Arg Ser Tyr
GCT CGG CAG CTT GTA GAT CGT TGC ATG AAA GTT CTG TAT TAC AGA GAC GCC CGC TCA TAC 630
190     200
Asn Arg Phe Glu Ile Thr Thr Val Thr Glu Ser Gly Val Glu Val Glu Gly Pro Leu Ser
AAT CGG TTT GAG ATC ACC ACG GTA ACA GAA AGC GGA GTG GAG GTT GAA GGG CCT CTG TCA 690
210
Ser Glu Thr Asn Trp Glu Ile Ala His Leu Ile Ser Gly Phe Glu ***
TCA GAA ACC AAC TGG GAA ATC GCT CAC CTG ATC AGC GGC TTC GAG TGA TCCCGCGGTCACGTT 753
TCTGCTCCTTCTTCTTCATCATCAGTTTTCTTATGTGTTTCTTGTGCTTATGTAATGGCACTTCTGTTTGTAAATGAAC 832
ATTAAACTGGGAGACAAATGGCTCGTTGGGACAACAAAAAAAAAAAAAAAAAAAAAAAAAAAAACGGAATTC--3' 910

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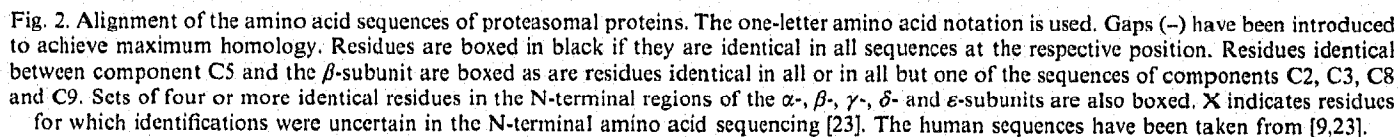
Fig. 1. Nucleotide sequence and deduced amino acid sequence of pituitary cDNA encoding the β -subunit of *Xenopus* proteasome. The putative signal for polyadenylation (ATTAAG) is overlined.

calculated molecular weight of 23 979. A computer homology analysis at the protein level (NBRF protein database; release 28) revealed that the N-terminal 24 amino acid residues of this protein are nearly identical (92% identity) to the N-terminal region that has been sequenced for the β -subunit of proteasome isolated from human erythrocytes [23] (Fig. 2); no significant identity with any other protein was found. A database search at the nucleotide sequence level (EMBL nucleotide database; release 27) showed no significant identity with any nucleotide sequence. We conclude that clone X7355 encodes the β -subunit of *Xenopus* proteasome. Northern blot analysis of *Xenopus* liver poly(A)⁺ RNA with clone X7355 as a probe revealed only one hybridizing band of about 1050 nucleotides (data not shown), indicating that X7355 is a nearly full-length cDNA clone.

3.2. Comparison between the amino acid sequences of proteasomal proteins

Purification of the human proteasome from erythrocytes revealed the existence of at least 13 subunits of different primary structures [23]. Most of these subunits have been found to be N-terminally blocked [23], a situation also reported for rat proteasome subunits [10].

Molecular cloning of cDNAs has revealed the structures of four of the N-terminally blocked proteasome subunits (components C2, C3, C8 and C9) [9–11,13,14]. In addition, for five of the non-blocked subunits the N-terminal sequences have been determined by Edman degradation and these subunits have been named α , β , γ , δ and ϵ in order of their decreasing apparent molecular weights (from 30 to 20 kDa) [23]. Cloning of one of the non-blocked subunits (C5) [9,12] has indicated that it corresponds to the γ -subunit of the proteasome. Alignment of the structures of components C2, C3, C5 (γ), C8 and C9 has shown that the amino acid sequences of C2, C3, C8 and C9 are similar while the sequence of C5 (γ) has diverged markedly from those of the other subunits. It has therefore been concluded that C5 (γ) is a new type of subunit of the proteasome complex [9,12]. The predicted amino acid sequence of the *Xenopus* β -subunit shows a low but significant identity with all subunits of human proteasome although the highest identity is with component C5 (γ) (Fig. 2). From the above it appears that the proteasome subunits form a multigene family which can be classified into at least two groups, one group consisting of the α -, β -, γ - (C5), δ - and ϵ -subunits with a free N-terminus and a second group including components C2, C3, C8 and C9 with



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- [5] Rivett, A.J. (1989) *J. Biol. Chem.* 264, 12215–12219.
- [6] McGuire, M.J. and DeMartino, G.N. (1989) *Biochem. Biophys. Res. Commun.* 160, 911–916.
- [7] Matthews, W., Tanaka, K., Driscoll, J., Ichihara, A. and Goldberg, A.L. (1989) *Proc. Natl. Acad. Sci. USA* 86, 2597–2601.
- [8] Driscoll, J. and Goldberg, A.L. (1990) *J. Biol. Chem.* 265, 4789–4792.
- [9] Tamura, T., Lee, D.H., Osaka, F., Fujiwara, T., Shin, S., Chung, C.H., Tanaka, K. and Ichihara, A. (1991) *Biochim. Biophys. Acta* 1089, 95–102.
- [10] Fujiwara, T., Tanaka, K., Kumatori, A., Shin, S., Yoshimura, T., Ichihara, A., Tokunaga, F., Aruga, R., Iwanaga, S., Kakizuka, A. and Nakanishi, S. (1989) *Biochemistry* 28, 7332–7340.
- [11] Tanaka, K., Fujiwara, T., Kumatori, A., Shin, S., Yoshimura, T., Ichihara, A., Tokunaga, F., Aruga, R., Iwanaga, S., Kakizuka, A. and Nakanishi, S. (1990) *Biochemistry* 29, 3777–3785.
- [12] Tamura, T., Tanaka, K., Kumatori, A., Yamada, F., Tsurumi, C., Fujiwara, T., Ichihara, A., Tokunaga, F., Aruga, R. and Iwanaga, S. (1990) *FEBS Lett.* 264, 91–94.
- [13] Tanaka, K., Kanayama, H., Tamura, T., Lee, D.H., Kumatori, A., Fujiwara, T., Ichihara, A., Tokunaga, F., Aruga, R. and Iwanaga, S. (1990) *Biochem. Biophys. Res. Commun.* 171, 676–683.
- [14] Kumatori, A., Tanaka, K., Tamura, T., Fujiwara, T., Ichihara, A., Tokunaga, F., Onikura, A. and Iwanaga, S. (1990) *FEBS Lett.* 264, 279–282.
- [15] Haass, C., Pesold-Hurt, B., Multhaup, G., Beyreuther, K. and Klotzel, P.-M. (1990) *Gene* 90, 235–241.
- [16] Fujiwara, T., Tanaka, K., Orino, E., Yoshimura, T., Kumatori, A., Tamura, T., Chung, C.H., Nakai, T., Yamaguchi, K., Shin, S., Kakizuka, A., Nakanishi, S. and Ichihara, A. (1990) *J. Biol. Chem.* 265, 16604–16613.
- [17] Zwickl, P., Lottspeich, F., Dahlmann, B. and Baumeister, W. (1991) *FEBS Lett.* 278, 217–221.
- [18] Sanger, F., Nicklen, S. and Coulson, A.P. (1977) *Proc. Natl. Acad. Sci. USA* 74, 5463–5467.
- [19] Thomas, P.S. (1980) *Proc. Natl. Acad. Sci. USA* 77, 5201–5205.
- [20] Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning, a Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- [21] Devereux, J., Haeberli, P. and Smithies, O. (1984) *Nucleic Acids Res.* 12, 387–395.
- [22] Martens, G.J.M., Weterings, K.A.P., Van Zoest, I.D. and Jenks, B.G. (1987) *Biochem. Biophys. Res. Commun.* 143, 678–684.
- [23] Lee, L.W., Moomaw, C.R., Orth, K., McGuire, M.J., DeMartino, G.N. and Slaughter, C.A. (1990) *Biochim. Biophys. Acta* 1037, 178–185.