

Genomic Exploration of the Hemiascomycetous Yeasts: 15. *Pichia sorbitophila*

Jacky de Montigny^a, Catherine Spehner^a, Jean-Luc Souciet^a, Fredj Tekai^a, Bernard Dujon^b,
Patrick Wincker^c, François Artiguenave^c, Serge Potier^{a,*}

^aLaboratoire de Génétique et Microbiologie, UPRES-A 7010 ULP/CNRS, Institut de Botanique, 28 rue Goethe, F-67000 Strasbourg, France

^bUnité de Génétique Moléculaire des Levures, Institut Pasteur/CNRS, département des Biotechnologies, Institut Pasteur, 25 rue du Docteur Roux, F-75724 Paris Cedex 15, France

^cGenoscope, Centre National de Séquençage, 2 rue Gaston Crémieux, P.O. Box 191, F-Evry Cedex, France

Received 3 November 2000; accepted 9 November 2000

First published online 29 November 2000

Edited by Horst Feldmann

Abstract This paper reports the genomic analysis of the strain CBS7064 of *Pichia sorbitophila*, a homothallic diploid yeast. We sequenced 4829 random sequence tags of about 1 kb and compared them to the *Saccharomyces cerevisiae* gene products. Approximately 1300 nuclear genes, 22 tRNAs, the rDNA locus, and six mitochondrial genes have been identified. The analysis of the rDNA genes has permitted to classify this organism close to the *Candida* species. Accession numbers from AL414896 to AL419724 at EMBL databank. © 2000 Federation of European Biochemical Societies. Published by Elsevier Science B.V. All rights reserved.

Key words: Comparative genomics; Random sequence tag; Hemiascomycete; *Saccharomyces cerevisiae*

1. Introduction

The hemiascomycete *Pichia sorbitophila* was isolated as a contaminant of a 70% sorbitol solution [1]. This organism has been described as a homothallic diploid [2]. It has not yet been classified in any phylogenetic tree. Interest in *P. sorbitophila* arose from its physiological properties distinct from *Saccharomyces cerevisiae*. In fact, this yeast is highly resistant to osmotic stress in general and to salt stress (4 M NaCl) in particular. These properties seem to be due to its ability to actively co-transport with protons glycerol, the major solute this yeast accumulates, in order to maintain osmotic balance [3,4].

Little is known about its genomic content. Recent approaches by pulsed field gel electrophoresis led us to propose that this yeast contains seven chromosomes with sizes ranging from 1 to 2.9 Mbp corresponding to an estimated total of 13.9 Mbp [5]. Two genes have been sequenced thus far: the *TIM10* *S. cerevisiae* homolog, a putative component of the mitochondrial protein import system [6] and the equivalent to the *LYS2* gene [7]. Three other ORFs with unknown function are present in the EMBL/DDJB databank (accession number Y18559).

This paper reports the genomic analysis of strain CBS7064 by exploring the sequence of 4829 random sequence tags

(RSTs) of about 1 kb and comparing them to the yeast gene products as described in [8]. Approximately 1300 new nuclear gene products, 22 tRNAs, the rDNA locus and six mitochondrial proteins were identified. The sequence of the rDNA has permitted to propose a phylogenetic position of this organism close to the *Candida* species [9].

2. Materials and methods

2.1. Strain

The *P. sorbitophila* strain used is CBS7064.

2.2. DNA extraction, library construction, sequencing, RST analysis and annotations

The protocols and the modifications described in [8,10–12] were followed with a minor modification for genomic DNA extraction: the DNA was recovered with a sterile glass rod instead of a classical isopropanol precipitation.

3. Results and discussion

3.1. Characterization of the RST

4829 RSTs which represent a total of 4036247 nucleotides were sequenced by Genoscope with less than 1.5% ambiguity. The RSTs have an average length of 836 nucleotides ranging from 40 to 1209 nucleotides (standard deviation, 189 nt). A total of 2159 inserts representing 4318 RSTs (95% of all RSTs) were sequenced on both ends and 90 had overlapping sequences. The remaining 511 RSTs (5%) were sequenced only at one end.

The DNA library sent to Genoscope contained 3936 clones representing a potential of 7872 RSTs; 4829 sequences representing 61.3% of the total expected RSTs were obtained and 3044 RSTs (38.6%) could not be sequenced. The experimental determination by *Eco*R1 digestion of the insert size from 96 random clones revealed that 28 plasmids (30%) were empty or had lost part of the insert or plasmid sequences including the hybridization site of the sequencing primer. These plasmid rearrangements pointing out the instability of the *P. sorbitophila* DNA in *Escherichia coli* explain why around 40% of the sequences could not be obtained. This instability increased after three or four successive cultures even on selective media.

The *Eco*R1 analysis showed that the 68 remaining plasmids carry inserts between 1 and 9 kb. The average size is 4.4 kb with a standard deviation of 1.7 kb. These results are in agreement with the expected size of the library inserts.

*Corresponding author. Fax: (33)-3-88 35 84 84.
E-mail: potier@gem.u-strasbg.fr

Table 2

Non-*S. cerevisiae* homologs of *P. sorbitophila*

Organism	Accession number	Functional comments
Ascomycetes		
<i>Aspergillus niger</i>	Q92406	NADH ubiquinone oxido-reductase 51 kDa subunit
<i>C. albicans</i>	P12461	thymidylate synthase (EC 2.1.1.45)
<i>C. albicans</i>	P43062	G ₁ /S-specific cyclin CLN2
<i>C. albicans</i>	P46599	serine threonine protein kinase STE7 homolog
<i>C. albicans</i>	P56553	cell growth protein
<i>C. albicans</i>	Q04802	glucosamine 6 phosphate isomerase (EC 5.3.1.10)
<i>C. tropicalis</i>	Q12600	SIS2 protein (halotolerance protein HAL3)
<i>Emmericella nidulans</i>	P48777	purine permease
<i>N. crassa</i>	P24918	NADH ubiquinone oxidoreductase precursor 78 kDa
<i>N. crassa</i>	P40915	NADH-ubiquinone oxido-reductase 24 kDa subunit
<i>N. crassa</i>	P48467	kinesin heavy chain
<i>Pichia angusta</i>	P12807	peroxisomal copper amine oxidase (EC 1.4.3.6)
<i>P. canadensis</i>	P48917	NADH-ubiquinone oxido-reductase chain 4 (EC 1.6.5.3)
<i>Schizosaccharomyces pombe</i>	T41665	putative dipeptidase
<i>S. pombe</i>	Q10088	putative agmatinase precursor (EC 3.5.3.11)
<i>S. pombe</i>	Q10449	hypothetical 57.2 kDa sp protein
Other eukaryotes		
<i>Rattus norvegicus</i>	P14408	fumarate hydratase mitochondrial precursor

5.8S and 5S in the same organization as in *S. cerevisiae*, sharing 89, 92, 94 and 89% of identity, respectively. In contrast, the intergenic regions within the rDNA locus show no similarity with those of *S. cerevisiae* [9].

3.5. Transposons

Transposable elements are ubiquitous in eukaryotes. In *S. cerevisiae*, 52 full-length retrotransposable elements were found as well as 268 solo long terminal repeats [16], which had been classified into five types: Ty1, 2, 4 and 5, which belong to the copia-like family and Ty3 which is a member of the gypsy-like family [17].

All RSTs and contigs were compared with the *S. cerevisiae* Ty element sequences but no significant matches were obtained. Comparisons of all *P. sorbitophila* putative ORFs with the protein sequences TyA and TyB did not reveal significant similarities. Since in *S. cerevisiae* the majority of the Ty elements are located in the vicinity of tRNA genes [18], the RSTs containing tRNA genes were carefully checked for retrotransposons though without success.

These results could therefore mean that transposon-like elements are present at a very low copy number in *P. sorbitophila*. Indeed, as we have identified roughly one third of the genome, one would expect to find a significant number of RSTs corresponding to Ty-like elements if their number would be in the same range as in *S. cerevisiae*.

At least three other explanations should be considered. The first were that Ty-like elements are present in *P. sorbitophila* but have considerably diverged from those in *S. cerevisiae*. The second would have to imply that *P. sorbitophila* does not contain retrotransposons at all and therefore constitutes a notable exception among eukaryotes. The third assumption would imply a cloning bias, due for instance to instability of *P. sorbitophila* transposons in *E. coli*.

3.6. Mitochondrial DNA

Mitochondrial DNA of *P. sorbitophila* was not comprised in a unique large contig. The BLASTX comparison of the 4829 RSTs to the *S. cerevisiae* mitochondrial protein database revealed 10 RSTs matching significantly and corresponding to the mitochondrial mRNA maturases BI2, BI3 and BI4, the

cytochrome *c* oxidase subunit II and the membrane-associated ATPase subunit 9. No mitochondrial tRNA or rDNA genes could be identified by a BLASTN analysis.

If only one RST of an insert matches with a mitochondrial sequence, while the second RST from the same insert shows no match, one may presume that the second RST also monitors the *P. sorbitophila* mitochondrial genome. This argument was used when analyzing the contigs including the 10 RSTs identified as belonging to mitochondrial DNA. By this procedure we have identified 32 further putative mitochondrial RSTs. The total of 42 RSTs were included in eight contigs representing 7.8 kb of mitochondrial DNA. This by excess evaluation estimates the size of the entire mitochondrial genome to be around 24 kb since one third of the genome was covered by the library. This is clearly less than the 86 kb of the *S. cerevisiae* mitochondrial genome [19].

This notion together with the obvious absence of tRNA and rRNA genes suggests that the mitochondrial DNA from *P. sorbitophila* is largely different from the *S. cerevisiae* one. None the less, we cannot exclude that the underrepresentation of mitochondrial RSTs could be due to differential mitochondrial DNA extraction or to instability of some plasmids in the *E. coli* library as discussed above.

3.7. Proteins with no orthologues in *S. cerevisiae* and phylogenetic classification

The occurrence of yeast specific genes not found in *S. cerevisiae* has already been described for several yeast species [20,21]. We therefore performed a general comparison of the *P. sorbitophila* RSTs with several entirely sequenced genomes and the SwissProt database depleted of the *S. cerevisiae* genes and their homologues [8].

We retained 17 RSTs whereof 16 matched to ascomycete genes and to *Candida albicans* or *Neurospora crassa* in particular (Table 2). Three *P. sorbitophila* RSTs showed significant matches with NADH ubiquinone oxidoreductase subunit genes of *N. crassa* and *Pichia canadensis* but not with those of *S. cerevisiae*. For three RSTs, the matches were significantly higher with *CLN7* and *HST7* of *C. albicans* and *HAL3* of *Candida tropicalis* than with the equivalent genes of *S. cerevisiae*.

These observations are consistent with the phylogenetic position of *P. sorbitophila* closer to the *Candida* species as suggested by the cladogram established from 18S and 25S rRNA sequence comparisons [9].

Acknowledgements: We thank Marie-Laure Straub and Yves Tourrette for technical assistance. This work was supported by Grant No 11-0926-99 from BRG (Bureau des Ressources Génétiques). The yeast genetics department of the UPRES-A 7010 is a member of the 'Génopole Alsace-Lorraine'. B.D. is a member of Institut Universitaire de France.

References

- [1] Rodrigues de Miranda, L., Appel, K.R. and Seyfarth, H. (1980) *Antonie Van Leeuwenhoek* 46, 157–159.
- [2] Soares de Oliveira, R.P. (1995) Thesis Universidade Do Minho (Braga, Portugal).
- [3] Lages, F. and Lucas, C. (1995) *Yeast* 11, 111–119.
- [4] Oliveira, R.P., Lages, F. and Lucas, C. (1996) *FEMS Microbiol. Lett.* 142, 147–153.
- [5] Sychrova, H., Braun, V., Potier, S. and Souciet, J.L., *Yeast*, in press.
- [6] Kayingo, G., Potier, S., Hohmann, S. and Prior, B.A. (2000) *Yeast* 16, 589–596.
- [7] Bleykasten-Grosshans, C., Prior, C. and Potier, S. (2000) *Yeast*, in press.
- [8] Tekaia, F., Blandin, G., Malpertuy, A., Llorente, B., Durrens, P. et al., *FEBS Lett.* 487, 17–30 (this issue).
- [9] Souciet, J.L., Aigle, M., Artiguenave, F., Blandin, G., Bolotin-Fukuhara, M. et al., *FEBS Lett.* 487, 3–12 (this issue).
- [10] Blandin, G., Llorente, B., Malpertuy, A., Wincker, P., Artiguenave, F. and Dujon, B. (2000) *FEBS Lett.* 487, 31–36 (this issue).
- [11] Artiguenave, F., Wincker, P., Brottier, P., Duprat, S., Jovelín, F. et al., *FEBS Lett.* 487, 13–16 (this issue).
- [12] de Montigny, J., Straub, M.L., Potier, S., Tekaia, F., Dujon, B. et al., *FEBS Lett.* 487, 52–55 (this issue).
- [13] Kawaguchi, Y., Honda, H., Taniguchi-Morimura, J. and Iwasaki, S. (1989) *Nature* 341, 164–166.
- [14] Suzuki, T., Ueda, T., Yokogawa, T., Nishikawa, K. and Watanabe, K. (1994) *Nucleic Acids Res.* 22, 115–123.
- [15] Lépingle, A., Casaregola, S., Bon, E., Neuvéglise, C., Nguyen, V.H. et al., *FEBS Lett.* 487, 82–86 (this issue).
- [16] Hani, J. and Feldmann, H. (1998) *Nucleic Acids Res.* 26, 689–696.
- [17] Jordan, I.K. and McDonald, J.F. (1999) *Genetics* 151, 1341–1345.
- [18] Kim, J.M., Vanguri, S., Boeke, J.D., Gabriel, A. and Voytas, D.F. (1998) *Genome Res.* 8, 464–478.
- [19] Foury, F., Roganti, T., Lecrenier, N. and Purnelle, B. (1998) *FEBS Lett.* 440, 325–331.
- [20] Ozier-Kalogeropoulos, O., Malpertuy, A., Boyer, J., Tekaia, F. and Dujon, B. (1998) *Nucleic Acids Res.* 23, 5511–5524.
- [21] Barth, G. and Gaillardin, C. (1997) *FEMS Microbiol. Rev.* 19, 219–237.