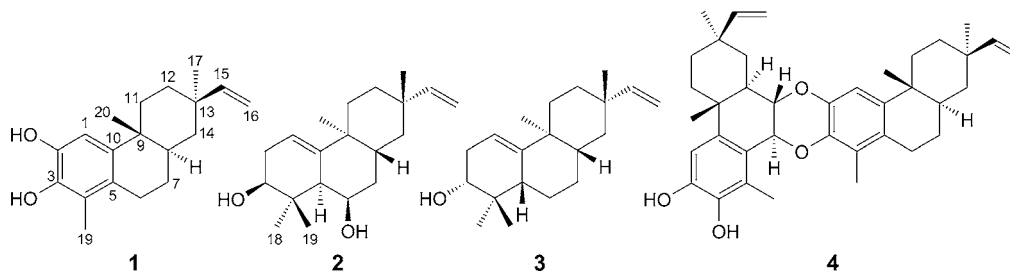


Two New Rosane-Type Diterpenoids from *Euphorbia ebracteolata* HAYATAby **Shu-Zhen Mu^{a)}**, **Chun-Rong Jiang^{a)}**, **Tao Huang^{a) b)}**, and **Xiao-Jiang Hao^{*a)}**^{a)} Key Laboratory of Chemistry for Natural Products of Guizhou Province and Chinese Academy of Sciences, Guiyang 550002, P. R. China (phone: +86-851-3804492; fax: +86-851-3805081; e-mail: haoxj@mail.kib.ac.cn)^{b)} China College of Life Science, Guizhou University, Guiyang 550025, P. R. China

Two new diterpenoids, euphebracteolatins A and B (**1** and **2**, resp.) were isolated from *Euphorbia ebracteolata* HAYATA, along with two known ones (3*R*)-*ent*-rosa-1(10),15-dien-3 α -ol (**3**) and yuexiandajisu D (**4**). The structures were established by extensive spectroscopic analyses and comparison with literature data.

Introduction. – The genus *Euphorbia* is the largest in the family Euphorbiaceae, comprising more than 2,000 species. About 80 species of the genus *Euphorbia* are distributed in China, with *Euphorbia ebracteolata* HAYATA widely occurring in China, Japan, Korea, and other countries. The root of *E. ebracteolata* is usually used to treat oedema, indigestion, bone tuberculosis, cough, and chronic bronchitis [1]. Previous chemical studies revealed the presence of terpenoids, acetophenone derivatives, flavonoids, tannins, steroids, and volatile oils in this genus [2], and some of them were found to exhibit several kinds of bioactivities [3]. Further investigation on the EtOH extract of this plant resulted in the isolation of two new diterpenoids, euphebracteolatins A and B (**1** and **2**, resp.), along with two known compounds, (3*R*)-*ent*-rosa-1(10),15-dien-3 α -ol (**3**), yuexiandajisu D (**4**; Fig. 1). In this article, we describe the isolation and structural elucidation of these compounds.

Fig. 1. The structures of compounds **1–4**

Results and Discussion. – Compound **1** was obtained as a light yellow oil. Its molecular formula was deduced as $C_{19}H_{26}O_2$ from its HR-EI-MS (m/z 286.1926 (M^+ , calc. 286.1933)), with seven degrees of unsaturation. The IR spectrum of **1** showed the

absorptions of OH (3424 cm^{-1}) and phenyl (1613 and 1485 cm^{-1}) groups. The ^{13}C -NMR (*Table*) and DEPT spectra indicated that **1** contains 19 C-atoms, including three tertiary Me, six CH_2 , three CH groups, and seven quaternary C-atoms. Among them, one monosubstituted olefinic C=C bond ($\delta(\text{C})$ 151.1 and 108.8) and one polysubstituted aromatic ring ($\delta(\text{C})$ 108.7, 140.9, 139.9, 122.7, 126.9, and 140.2) were indicated, which accounted for four degrees of unsaturation, the remaining three could be attributed to the presence of a three-ring system for **1**. The ^1H -NMR data (*Table*) exhibited characteristic signals for the aromatic H-atom ($\delta(\text{H})$ 6.70 (*s*, 1 H) and the olefinic H-atoms ($\delta(\text{H})$ 5.90 (*dd*, $J = 10.4, 17.2$, 1 H), 4.98 (*dd*, $J = 1.2, 17.2$, 1 H), and 4.90 (*dd*, $J = 1.2, 10.4$, 1 H).

Table. ^1H - and ^{13}C -NMR (400 and 100 MHz, resp.) Data of Compounds **1** and **2** in CDCl_3 , δ in ppm, J in Hz. Atom numbering as indicated in Fig. 1.

Position	1		2	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
1	6.70 (<i>s</i>)	108.7	5.45–5.42 (<i>m</i>)	116.2
2	–	140.9	2.31–2.27 (<i>m</i> , H_a), 1.96–1.89 (<i>m</i> , H_b)	31.8
3	–	139.9	3.53 (<i>dd</i> , $J = 6.0, 10.4$)	74.8
4	–	122.7	–	36.7
5	–	126.9	2.05–1.97 (<i>m</i>)	42.0
6	2.68–2.61 (<i>m</i>)	26.8	4.05–3.99 (<i>m</i>)	70.6
7	1.65–1.54 (<i>m</i>)	25.6	2.35–2.30 (<i>m</i> , H_a), 1.30–1.37 (<i>m</i> , H_b)	30.6
8	1.79–1.73 (<i>m</i>)	36.3	1.73–1.66 (<i>m</i>)	37.7
9	–	36.4	–	36.6
10	–	140.2	–	147.9
11	1.95–1.92 (<i>m</i>)	33.9	1.66–1.59 (<i>m</i> , H_a), 1.49–1.41 (<i>m</i> , H_b)	36.5
12	1.41–1.34 (<i>m</i>)	32.8	1.25, 1.50 (<i>2m</i>)	32.6
13	–	36.3	–	36.3
14	1.49–1.42 (<i>m</i> , H_a), 1.23–1.15 (<i>m</i> , H_b)	39.5	1.73–1.66 (<i>m</i> , H_a), 1.10–1.20 (<i>m</i> , H_b)	33.9
15	5.90 (<i>dd</i> , $J = 10.4, 17.2$)	151.1	5.90 (<i>dd</i> , $J = 10.8, 17.6$)	151.0
16	4.90 (<i>dd</i> , $J = 1.2, 10.4, \text{H}_a$), 4.98 (<i>dd</i> , $J = 1.2, 17.2, \text{H}_b$)	108.8	4.91 (<i>dd</i> , $J = 1.2, 10.8, \text{H}_a$), 5.00 (<i>dd</i> , $J = 1.2, 17.6, \text{H}_b$)	109.1
17	1.02 (<i>s</i>)	22.7	1.00 (<i>s</i>)	22.0
18	–	–	1.04 (<i>s</i>)	23.7
19	2.11 (<i>s</i>)	11.4	0.70 (<i>s</i>)	12.8
20	1.00 (<i>s</i>)	21.3	1.10 (<i>s</i>)	23.8

Comparison of the ^1H - and ^{13}C -NMR data of **1** with those of the known compound 19-norrosa-1,3,5(10),15-tetraene-2,18-diol indicated that both compounds were very similar except for C(3) and C(19), implying they share the same basic skeleton of a rosane-type diterpenoid [4]. Detailed analysis of 1D-NMR spectra of **1** revealed the following differences of chemical shifts: one OH group and the Me group were indicated in place of the CH and CH_2OH groups, respectively, in the known compound 19-norrosa-1,3,5(10),15-tetraene-2,18-diol. On the basis of the HMBCs features of H–C(1) to C(3), and of Me(19) to C(4), C(3), and C(5), the OH group and the Me group were assigned to C(3) ($\delta(\text{C})$ 139.9) and at C(19) ($\delta(\text{C})$ 11.4, $\delta(\text{H})$ 2.11),

respectively. The gross structure of **1** was finally established from its 2D-NMR spectra as shown in Fig. 2. The $^1\text{H},^1\text{H}$ -COSY spectrum of **1** evidenced the presence of three partial structures, **a**–**c**. The connectivity of each partial structure was clarified by the HMBC spectrum as shown in Fig. 2.

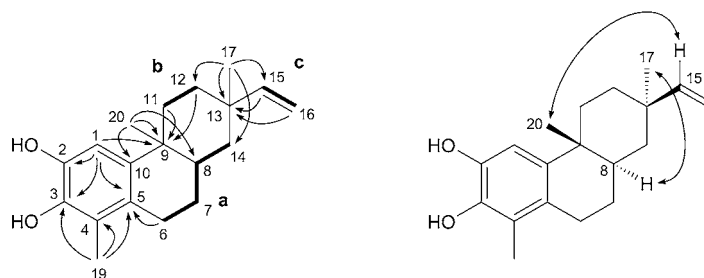


Fig. 2. Key $^1\text{H},^1\text{H}$ -COSY (—), HMB (H \rightarrow C), and ROESY (H \leftrightarrow H) correlations of **1**

The relative configuration of **1** was consistent with that of the known compound 19-norrosa-1,3,5(10),15-tetraene-2,18-diol as deduced from ROESY spectrum of **1** (Fig. 2). The correlation between Me(17) and H–C(8) indicated that Me(17) was α -oriented, while Me(20) had the β -orientation on the basis of the correlation between Me(20) and H–C(15). Thus, the structure of **1** was established as 2,3-dihydroxy-18-norrosa-1(10),2,4,15-tetraene, named euphebracteolatin A.

Compound **2** was obtained as optically active white amorphous powder. The molecular formula was determined as $\text{C}_{20}\text{H}_{32}\text{O}_2$ by HR-EI-MS at m/z 304.2401 (M^+ ; calc. 304.2402), with five degrees of unsaturation. The IR spectrum of **2** implied the presence of OH (3345 cm^{-1}) groups. The ^{13}C -NMR (Table) and DEPT data of **2** exhibited 20 signals due to one monosubstituted C=C bond ($\delta(\text{C})$ 151.0 and 109.1), one trisubstituted C=C bond ($\delta(\text{C})$ 116.2 and 147.9), three sp^3 quaternary C-atoms, and four sp^3 -CH, five sp^3 -CH₂, and four tertiary sp^3 -Me groups. Of these, two CH groups ($\delta(\text{C})$ 74.8 and 70.6) were ascribed as being attached to OH groups.

The ^1H -NMR (Table) of **2** confirmed the presence of three olefinic H-atoms ($\delta(\text{H})$ 5.90 (*dd*, $J = 10.8, 17.6$, 1 H), 5.00 (*dd*, $J = 1.2, 17.6$, 1 H), and 4.91 (*dd*, $J = 1.2, 17.6$, 1 H)), four tertiary sp^3 -Me groups ($\delta(\text{H})$ 1.10 (*s*), 1.04 (*s*), 1.00 (*s*), and 0.70 (*s*)), and two O-bearing CH groups ($\delta(\text{H})$ 4.05–3.99 (*m*) and 3.53 (*dd*, $J = 6.0, 10.4$). These data indicated that compound **2** was a rosane-type diterpenoid, similar to the known compound (3*R*)-*ent*-rosa-1(10),15-dien-3 α -ol (**3**) [4]. Further comparison of the 1D-NMR data of **2** with those of **3** revealed that the main difference between the two compounds was the presence of one OH group in **2**. The OH group was located at C(6) evidenced by the HMBCs of H–C(6) to C(4), of CH₂(7) to C(6), and of H–C(8) to C(6). Detailed analysis of the 2D-NMR, including HMQC, $^1\text{H},^1\text{H}$ -COSY, and HMBC as shown in Fig. 3, confirmed the above conclusion. The relative configuration of **1** was established from the ROESY spectrum. The correlations H–C(3)/Me(18), Me(18)/H–C(5), H–C(5)/H–C(6), and Me(20)/H–C(15) indicated that H–C(3), H–C(5), H–C(6), and Me(20) were α -oriented, and accordingly Me(17) and the OH at C(3) and C(6) were β -oriented. H–C(8) was β -oriented on the basis of the correlation Me(17)/

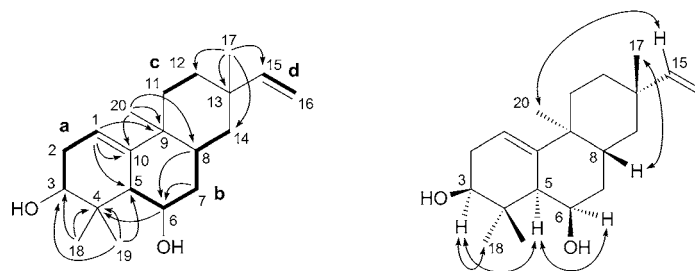


Fig. 3. Key $^1\text{H},^1\text{H}$ -COSY (\rightleftarrows), HMB ($\text{H} \rightarrow \text{C}$), and ROESY ($\text{H} \leftrightarrow \text{H}$) correlations of **2**

H–C(8). Thus, the structure of **2** was established as *ent*-rosa-1(10),15-dien-3 β ,6 β -diol, named euphebracteolatin B.

By comparison with the NMR and MS data with those reported in the literature, compounds **3** and **4** were identified as (3*R*)-*ent*-rosa-1(10),15-dien-3 α -ol [4] and yuexiandajisu D [5], respectively.

Compounds **1–4** were evaluated for their cytotoxic activities against human ovarian carcinoma cell line SK-OV-3. At the 1.00×10^{-5} M concentration, euphebracteolatin A (**1**) showed weak inhibition of tumor growth with a value of $14.90 \pm 13.46\%$, while compounds **2–4** were inactive.

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Experimental Part

General. All solvents used were distilled prior to use. Column chromatography (CC): silica gel (SiO_2 ; 200–300 and 300–400 mesh; *Qingdao Marine Chemical Company*, P. R. China), reversed-phase *RP-18* (40–63 μm ; *Merck*), and *Sephadex LH-20* (40–70 μm ; *Amersham Pharmacia Biotech AB*, Sweden). Optical rotations: *JASCO-P1020* digital polarimeter. UV: *Shimadzu UV-2401PC* spectrophotometer; λ_{max} ($\log \epsilon$) in nm. IR Spectra (KBr): *Bruker Tensor 27 FT-IR* spectrophotometer; $\tilde{\nu}$ in cm^{-1} . 1D- and 2D-NMR spectra: *INOVA-400* MHz NMR spectrometer in CDCl_3 ; δ in ppm rel. to Me_4Si as internal standard, J in Hz. HR-EI-MS: *Waters AutoSpec Premier P776* spectrometer; in m/z .

Plant Material. The roots of *E. ebracteolata* were collected from Changchun, Jilin Province of China, in February 2006. The sample was identified by Prof. *Li Gao* from the Natural Drug Resources Laboratory of the Yunnan Institute of Materia Medica. A voucher specimen (GZCNP 0807024) was deposited with the Key Laboratory of Chemistry for Natural Products of Guizhou Province and Chinese Academy of Sciences.

Extraction and Isolation. The air-dried roots (20 kg) of *E. ebracteolata* were powdered and extracted with 95% EtOH ($3 \times 30 \text{ l}$, each for 5 d) at r.t. and concentrated *in vacuo* to give a crude extract. The extract was suspended in H_2O and extracted successively with petroleum ether (PE) and CHCl_3 . The CHCl_3 fraction (300 g) was subjected to CC (SiO_2 200–300 mesh); PE/acetone 100:0 to 0:100) to afford five fractions, *Frs. 1–5*. *Frs. 1, 3, and 4* were purified by repeated CC (SiO_2 ; *RP-18*; and *Sephadex LH-20*; $\text{CHCl}_3/\text{MeOH}$ 0:1–1:1) to afford **1** (83 mg), **2** (74 mg), **3** (61 mg), and **4** (16 mg).

Euphebracteolatin A (=rel-(4*bR*,7*R*,8*aR*)-7-Ethenyl-4*b*,5,6,7,8,8*a*,9,10-octahydro-1,4*b*,7-trimethylphenanthrene-2,3-diol; **1**). Light yellow oil. $[\alpha]_{\text{D}}^{25} = +97.6$ ($c = 0.42$, CHCl_3). UV (CHCl_3): 286 (3.1), 241 (2.9). IR (KBr): 3424, 2925, 1635, 1613, 1485, 1175, 1026. ^1H - and ^{13}C -NMR: see the *Table*. HR-EI-MS: 286.1926 (M^+ , $\text{C}_{19}\text{H}_{26}\text{O}_2^+$, calc. 286.1933).

Euphebracteolatin B (= rel-(2R,4bR,7R,8aS,10S,10aS)-7-Ethenyl-1,2,3,4b,5,6,7,8,8a,9,10,10a-dodecahydro-1,1,4b,7-tetramethylphenanthrene-2,10-diol = rel-(3 β ,6 β ,8 β ,9 α ,13 α)-Rosa-1(10),15-diene-3,6-diol; **2**). White amorphous powder. $[\alpha]_{\text{D}}^{23.0} = +6.2$ ($c = 1.00$, CHCl₃). IR (KBr): 3578, 3345, 2966, 2930, 1636, 1450, 1363, 1048. ¹H- and ¹³C-NMR: see the Table. HR-EI-MS: 304.2401 (M^+ , C₂₀H₃₂O₂⁺; calc. 304.2402).

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