Isoquinoline Alkaloid Biosynthesis

Speculations on some unsolved problems

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Introduction

It is now generally accepted that the 1-benzyl-1,2,3,4-tetrahydroisoquinoline alkaloids arise in Nature from tyrosine, and thence **3,4-dihydroxyphenylalanine** (DOPA), approximately in accordance with **¹**the scheme outlined in 1910 by Winterstein and Trier . **Onoe** formed, these isoquinoline derivatives **are** the preoursors of a large array of polynuclear structures (Scheme **I).** An **enormous** amount of effort has been expended over the last 20 years or so to establish these inter- $2-6$
relationships $2-6$. The methods used have involved feeding the plant with a postulated precursor to the alkaloid(s) under examination, suitably labelled with 14 C, 15 _N, 2 _H or ³H atoms at specific sites, More recently some very elegant work has shown how the enzyme systems of plants are used to generate, stereospecifically, asymmetrio centres, **⁸**especially from prochiral methylene groups . The concept of phenol oxidation⁹⁻¹⁶ has proved to be of supreme importance for a clearer understanding of the individual steps in the biosynthetic pathways.

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Despite this vast body of knowledge there are **some** intriguing problems still outstanding. Among these are **(a)** the mechanism of formation of the **1-benzyltetrahydroisoquinolines,** especially the decarboxylation of tetrahydroisoquinoline-1-carboxylic acids, (b) the dehydrogenation of the nitrogen-containing ring to the fully aromatic state present in such structures **as** papaverine, berberine and sanguinarine. **(c)** the biosynthesis of the pavinane and isopavinane alkaloids, (d) the mechanism of phenol oxidation in vivo and (e) some Unusual Oxygenation patterns.

Formation of **1-benzyl-1,2,3,4-tetrahydroisoquinolineg**

1 In their original scheme , Winterstein and Trier postulated that DOPA (1) is converted into dopamine (2) by decarboxylation, and into **3.4-dihydroryphenylacetaldehyde** (3) by decarboxylation and deamination (Scheme II), followed by a Pictet-Spengler type of condensation to give rise to norlaudanosoline (4). It was shown¹⁷ that in vitro high vields of **1-benzyltetrahydroisaquinolines** could be obtained from (2) and (3) under so-called "physiological conditions". It has been postulated that DOPA is converted into the pyruvic acid (5), rather than the aldehyde **(3), and** that condensation with **(2)** provides **norlaudanosoline-I-carboxylic** aoid ¹⁸**(6),** which subsequently undergoes decarboxylation to **(4)** . This proposal has the great merit that both dopamine and (5) are normal products of \propto -amino acid metabolism. Hahn, et al.¹⁸ showed that the condensation reactions occur in vitro in high yield under "physiological" conditions, but the 1-benzyltetrahydroisoquinoline-1-carboxylic acids proved to be very resistant to decarboxylation under conditions even remotely similar to those existing in the plant cell. However, these proposals have been

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revived recently^{19,20}, especially since the isolation of peyoxylic acid **22 (7)** and peyoruvic acid **(8) from cacti²¹.** It was established²² that **these acids are efficiently incorporated into anhalanine (9) and anhalonidine (lo), when fed to the appropriate plants. Further, it was reported**²² that when peyoruvic acid (8) was incubated with fresh

 (8) , $R = Mc$

 (11)

cactus slices, the corresponding 3.4-dihydroisoquinoline (11) was produced. This observation suggests that the decarboxylation of a tetrahydroisoquinoline-1-carboxylic acid to the tetrahydroisoquinoline may be a two-step process involving oxidative decarboxylation to the 3,4-dihydroisoquinoline, by processes discussed below, followed by a (stereospecific) reduction 23,24 . Some recent results using <u>Papaver orientale</u> 20 and P. somniferum²⁵ have shown conclusively that (6) is formed from DOPA and that **(6a)** is probably an intermediate between (6) and (4). **Some** time ago the oxidative decarboxylation of **tetrahydroisoquinoline-3** carboxylic acid, using sodium hypochlorite, (equation 1) was studied as a biosynthetic model, but the results were inconclusive²⁶. More recently²⁷

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\left(\bigcup_{N\in\mathbb{N}}\right)_{N\in\mathbb{N}}
$$

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it has been found that decarboxylation of (12) can be achieved by treatment with dioycloherylcarbodiimide (equation 2). The method of generation of the imminium ion **was** used in a synthesis of ajmaline.

However, Bobbitt and Cheng²³ have suggested that the phenolic hydroxyl group of, for example (4), rather than the nitrogen atom, might be the point of oxidation thaf triggers the loss of carbon dioxide. Several mechanisms could be written for such a process; one showing the intermediate formation of an 0-quinone and another involving a phenoxy cation (see later) are represented in Scheme III. It has been found^{23,24} that anodic oxidation of the **tetrahydroisoquinoline-1-carboxylio** acids (13) give the **3,4-dihydroisoquinolines** in a reaction that involves a single, two-electrons **wave** at a potential at which phenol oxidations

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are known to occur. When the phenolic isoquinoline derivative (14) was

 α xidised electrolytically^{23,24}, the major product was (15), together with **some (16) and (17). The product (16) could be isomerised to (17) with acids.**

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When the amino acid (18) was oxidised with periodic acid, the product was (21) and since the amount of oxidant consumed was exceptionally high, it was proposed²⁸ that the reaction proceeded via (19) and the quinone **methide (20).**

 (21)

 (20)

Alkaloids Derivable from 4-Hydroxylaudanosoline

Noradrenaline²² and adrenaline²³ are normal constituents of mannnalian tissues; they are biosynthesised from tyrosine by way of DOPA and dopamine²⁹. Noradrenaline has also been detected in a number of plant species³⁰ and probably is formed by a route similar to that in mammalian tissues. It has been shown that noradrenaline is the precursor of berberastine²⁴ in Hydrastis canadensis^{31,32}, When dopamine-1-¹⁴C $(3,4-$ (OH) ${}_{2}$ C₆H₃CH₂^{CH}₂NH₂) was fed to the plant, radioactive berberastine (24),

berberine (25), canadine (tetrahydroberberine) and hydrastine (26) were isolated. Degradations revealed that the berberine was specifically labelled at C_5 . It is

possible that (24) arose by hydroxylation of berberine, but this is unlikely since berberastine had a higher specific activity than berberine or canadine. When $({}^{+})$ -noradrenaline-2- 14 C was fed to H.canadensis high levels of incorporation into berberastine were found; incorporations into berberine and canadins were only one-sixth of those achieved with dopamine- 1^{-14} C. It was concluded that norlaudanosoline (4) cannot be the precursor of berberastine, since it had been established^{33,34} that (4) is the specific precursor for berberine, and **4-hydrorynorlaudanosoline** (27) seems to be the logical precursor; the stereochemistry of (27) at C_1 and C_4 have not been discussed.

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(2 8)
$$

 \mathbf{I}

5 Me

ÔН

JMe

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 (31)

 (30)

MeO

Me O

The intriguing possibility arises that 4-hydroxynorlaudanosoline is the starting point for a range of alkaloids in parallel to the large number of structural types derivable from norlaudanosoline itself (scheme I). This concept is supported by the characterisation of compounds such as thalidastine (28)³⁵, tetrahydroberberastine³¹, stephorphine (29)³⁶, cataline (30)³⁷, imenine (31)³⁸, erythrinine (32)³⁹ **⁴⁰and erythistemine (33)** .

 (32)

 (33)

It has always been supposed $41,42$ that the pavinane alkaloids, such **as** argemonine (34), are biosynthesised from reticuline **(35).** However, when 14c-labelled rstioulins **was** fed to Argemone mexicana **L.,** the argemonine was not radioactive⁴², but this may be due to the fact that only very small amounts of (34) are present in this plant⁴³. It was

postulated by Stermitz and Seiber⁴³ that, in view of the known⁴⁴ dehydrogenation of reticuline to (36) (which probably occurs via the N-oxide and then the pseudobase), the 3.4-dihydroisoquinoline **(36)** could isomerise to (37), which might then undergo cyclisation to the pavinane system.

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If, on the other hand, retiouline can41 be hydroxylated enzymatically to (38), conversion to (37) is easily visualised.

 (38)

A very attractive alternative possibility⁴⁵ is that the precursor **of the pavinanes is 4-hydroxynorlaudanosoline (27) which becomes partially methylated, probably to 4-hydroxyreticuline (39). which then undergoes dehydration to the ensmine (40) and protonation to yield (37). It has been**

known for some time⁴⁶ that enamines such as (40) are cyclised to pavinanes under acid conditions. Since such enamines are also known^{47,48} to undergo rearrangement into **3-beneyl-3.4-dihydroisoquinolines** under mild acid conditions, it may be anticipated that 3-benzylisoquinoline derivatives should occur **8s** natural products, although none have been reported **so** far.

This scheme for the biosynthesis of pavinanes has the great merit that the genesis of isopavinanes is also readily explained by the cyclisation of the 4-hydroxy-1-benzyltetrahydroisoquinoline derivative, for example (39) \rightarrow (41) . Acid-catalysed cyclisations of this type in vitro are well

known⁴⁹; more significantly it has been found⁴⁹ that when the acetal (42) is treated with 2N HC1 at room temperature the isopavinane (43) and the pavinane (44) are produced. It is also significant that the pavinane and

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isopavinane alkaloids possess the same absolute configuration 50 .

It is generally accepted that in the biosynthesis of the isoquinoline ring, DOPA is first decarboxylated to dopamine, and it is known that when 14 Clabelled dopamine is fed to plants that produce alkaloids of the isoquinoline group, the products are specifically labelled. Nevertheless it is still possible that DOPA, and not dopamine is condensed with the α -keto acid (5), to give rise to a **tetrahydroisoquinoline-1,3-dicarbouylic** acid (45). It is then possible for this to lose one oarboxyl group to yield **(6)** or (46) (Scheme IV).

SCHEME IV

 (46)

When (\pm) -m-tyrosine-2-¹⁴C (47) is fed to Euphorbia myrsinites, the alkaloid (48) can be isolated, specifically labelled⁵¹.

An alternative mode of biosynthesis of pavinanes that does not involve **4-hydroxynorlaudanosoline** (27) can then be considered in which a l-beneyltetra**hydroisoquinoline-3-ca~boxylic** acid, for example **(49),** undergoes oxidative decarboxylation via (50) and thence to the enamine (40). However, to explain the formation of isopavinanes it would be necessary to postulate hydration of

(40) to give (39). This reaction is known $\underline{\text{in}}$ $\underline{\text{vitro}}$ ⁵². Of course both pavinanes and isopavinanes could arise from reticuline if initial oxidation to the quinone methide (51) is assumed to occur (Scheme V) followed by cyclisation, or hydroxylation to (39), followed by further reactions as discussed above. Examples of C₄-hydroxylation of 7-hydroxytetrahydroisoquinolines in vitro are known⁵³ involving oxidation with lead tetraacetate.

SCHEME V

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One of the many developments of this reaction concerned the synthesis of an isopavinane from a 1-benzyl-7-hydroxytetrahydroisoquinoline derivative⁵⁴. Dehydrogenation to Fully Aromatic Isoquinolines

The actual pathway adopted in vivo for the dehydrogenation of a **1,2,3,4-tetrahydroisoquinoline** into the fully aromatic state, **for** example in papaverine, **is** largely unknown, but several possibilities can be considered. Thus, if it is assumed that 4-hydroxyreticuline can be dehydrogenated to the 1,2-dehydro $\stackrel{\text{44}}{\text{compound}}$, by analogy with the known 44 similar reaction with reticullne, aromatisation is easily understood (equation 3) and laboratory analogies for this process do exist⁵⁵. Of

course the quinone methide (51) is equivalent to (39) and tautomeric shift would give the enamine (40), which could then be dehydrogenated as outlined in equation **4.** It is possible to envisage the formation of the fully aromatic isoquinoline by successive oxidative decarboxylations of a **tetrahydroisoquinoline-l,3-dioarboxylic** acid. However, in the opium poppy at least, it has been that tetrahydropapaverine is the immediate

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Precursor of papaverine. The data are not inconsistent with a demethylation-remethylation sequence which would be necessary if a quinone methide (52) were an intermediate (equation 5).

t.

 \cdots Eq. 5

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It has been established⁵⁷ that in Chelidonium majus chelidonine (57) is derived from (\pm) -stytopine (53); the intermediates between them probably **are (541, (55) and (56) (Scheme VI). It is possible that sanguinarine (59)**

 (59)

is the product from further transformation of chelidonine, but, alternatively, it may arise from an intermediate such **as** (56). via dihydrosanguinarine (58). Thus, aromatisation of the nitrogen ring in this case is a particularly favoured process.

The formation of protoberberines from the initially formed **tetrahydroprotoberberines** might involve the initial formation of the N-oxide (60) as depicted in Scheme VII. It is known⁵⁸ that tetrahydroprotoberberine N-oxides **can** be rearranged in vitro to the oarbinolamine structure (61).

SCHEME VII

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Phenol Oxidation

The role of phenolic oxidation in the biosynthesis of many groups of natural products in general, and of alkaloids of the isoquinoline group in particular, cannot be overemphasised. The subject has been reviewed on a number of occasions⁴, 5, 9, 10, 12-16, 59 Until recently it was assumed $9,10$ that the oxidative coupling of phenols involves the generation and pairing of radicals (equation 6). but in a preface to a book on oxidation in organic

 $A\text{rOH}$ \rightarrow $A\text{rO}$ \rightarrow $A\text{rO}$ \rightarrow $A\text{rO}$ \rightarrow eg 6 then $2 \times \text{ArO}^* \rightarrow C - O - C$ **C-C**)) dimers

Ohemistry, Barton6' has briefly reviewed several other mechanisms that have been proposed from time to time, and in a chapter in that book McDonald and $\frac{61}{100}$ have examined very carefully the mechanism of phenolic oxidation reactions. In vitro oxidative coupling reactions are usually carried out using one-electron transfer agents (potassium ferricyanide is very commonly used); the evidence in favour of a radical mechanism for many of these reactions is overwhelming⁶², but Thyagarajan⁶³ in his review of ferricyanide oxidations States "although the formation of products is well understood **8s** resulting from the radical substitution or radical pairing of the mesomeric aryloxy radicals, kinetic studies of the oxidation of phenols by ferricyanide show that the dimers could well be represented as arising from condensations between phenol molecules and mesomeric aryloxy cations⁶⁴". Aryloxonium ions have been investigated in the past $65-67$, and electrochemical studies have shown $68,69$ that two electrons can be removed from a phenol.

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Waters⁷⁰ points out that aryloxy cations should exist largely as the mesomeric carbonium ion and should be a powerful agent in carbon-carbon bond formation by electrophilic substitution, whereas the neutral radical species, **ArO,** should exist largely with the odd electron on oxygen and should give rise preferentially to carbon-oxygen coupling.

There are some oxidative coupling reactions that are best written as involving the aryoxy cation, or an equivalent species containing a metal to oxygen bond. Thus, oxidation of phloretic acid (62, $R=H$)⁷¹ and N-carbomethoxytyrosine (62, R=NHCOMe)⁷² to (64, R=H) and (64, R=NHCOMe), respectively can be regarded as proceeding through an intermediate (63a) rather than **(63b).**

 $(63b)$

 (64)

⊊q, 7

The oxidation6' of **(65)** to **(67)** is best written as proceeding via the cation (66). An aryloxy cation was postulated⁴² in the biosynthesis

of **tetrahydroprotoberberines from** reticuline (Scheme **VIII).** It has been argued, however, that since codamine is not incorporated into thebsine in P.somniferum a Cation cannot be involved (equation **7).** but it is possible that activation by methoxyl is not sufficient; phenolic hydroxyl or even a phenolate anion may **be** needed.

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SCHEME VIII

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Oxidative coupling of monophenolic compounds using thallic trifluoracetate⁷³ and vanadium oxyhalides⁷⁴ has recently attracted considerable attention. The reactions (Scheme **IX)** can be written as involving an aryloxy cation or the equivalent metal complex $-$ a twoelectrons process is involved. Yields of coupled products have been very high - in marked contrast to those reactions involving one-electron transfer agents such as ferricyanide¹⁴.

SCHEME IX

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The argument in favour of radical mechanisms for oxidative coupling in vivo has always been that crude extracts of the "oxidase" enzymes, which can bring about phenolic coupling, seemed to be radical in character. The essential reaction has been thought to involve the $Fe³⁺ \rightarrow Fe²⁺ + \epsilon$ couple. However cytochrome P-450, which is probably involved in alkaloid biosynthesis⁷⁵ is more complex in character^{76,77}. Multi-electron oxidations are involved, where successive one-electron oxidations may be "stored", probably by an oxygen moleoule so that the effective oxidant may be 0_2 ⁻⁻, rather than Fe^{3+} . The mechanism of the coupling reaction can then be depicted⁷⁸ as in Scheme X.

SCHEME X

When a benzylic carbon-hydrogen bond is available an alternative site of attack can lead to the benzylic alcohol or the quinone methide (Scheme XI).

SCHEME XI

If the intermediacy of aryloxy cations is accepted it is possible to rationalise some otherwise puzzling biosynthetic sequences in the

isoquinoline alkaloids. Thus the 2-benzylisoquinolines sendaverine^{79,80} and corgoine⁸¹ might arise from the 1-benzyl isomers, for example coclaurine, either from a quinone methide (68) or the equivalent benzyl alcohol (69)(Scheme X11). When norarmepavine was oxidised with peroxidase and $H_2O_2^{82}$ one of the products was O-methylsendaverine. A dfradicsl intermediate **xas** postulated to account for this reaction (equation 8).

Corgoine $R = H$: Sendaverine $R = Me$

If a mono ether of a 4,5-dialkylcateohol is oxidised and hydroxylated in an ionic process, the product obtained can be rationalised as shown in equation \int .

The trioxyphenethylamines that are known $\frac{83-85}{10}$ to be on the pathway to the "simple" tetrahydroisoquinolines can be generated as shown in equation $\mathbb Q$.

The derivation of narcotine **(701,** and **some tetrahydroprotoberberines,** such as capaurine (71) and capaurimine (72) are easily rationalised in the same way, with hydroxylation occurring probably at the l-benayltetrahydroisoquinoline stage (equation 1). If the alternative quinone methide can be

formed (equation 12), the oxygenation patterns of thalifendlerine (73) and takatonine (74) are also explained. The occurrence of 3-oxyaporphines

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may also be rationalised in a similar way. It has recently been 8 6 realised that some of the **penta-oxybenzo[c]phenanthridine** alkaloids such as chelirubine are more properly represented with the fifth oxygen function at C_{10} , for example (75) rather than at C_{11} as hitherto. The biosynthetic origin of these compounds is not yet known.

 (75)

When 7-hydroxytetrahydroisoquinolines are treated with lead tetraacetate, the 4-acetoxy derivative is produced 53 and this reaction **can** be written as involving the cation (76), although the processes summarised in **(77)** may be preferred (Scheme XIII).

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The reaction has been applied to the synthesis of aporphines⁸⁷ (equation 13).

It mignt be worthwhile considering the biosynthesis of aporphines to involve aryloxy cations instead of the diradical processes ususlly aasumed. The latter concept requires two separate one-electron oxidations **st** different sites in the molecule. It is difficult to envisage such oxidations ocourring from two different haem units in one enzyme; the alternative, two separate oxidations from one haem unit by movement of the 1-beneylisoquinoline radical to bring the second site into position, is hard to visualise. Annelations could be more simply effected by a twoelectrons oxidation at the same site on the substrate molecule.

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