STRUCTURE OF CASEADINE

Tuticorin R. Govindachari, Bantwal R. Pai, Hosbett Suguna and Manakkal S. Premila

Department of Chemistry, Presidency College, Madras-600005, India

A reinvestigation of the spectral data of the synthetic 1-hydroxy-2,10,11-trimethoxyberbine and natural caseadine shows that their nmr spectra are almost identical with very minor differences; so are their ir spectra in solution except for an additional band at 720 cm⁻¹ in the natural product. These minor differences could not be due to structural or stereochemical differences but perhaps due to some impurity in the natural sample.

Caseadine, caseamine and caseanadine are naturally occurring tetrahydroprotoberberines, isolated from Corydalis caseana A. Gray and assigned structures (1), (2) and (3) respectively. Cava et al. synthesised caseadine-O-methyl ether (4) and found it to be identical with the methyl ether of natural caseadine (1). Thus the oxygenation pattern in caseadine (1) was settled. By an independent route Ishiwata and Itakura³ synthesised (4) and found it to be identical with the O-methyl ether of caseadine. Iida et al. synthesised dl-caseadine (m.p.115-118°) and found the O-methyl ether to be identical

[†]Studies in Protoberberine Alkaloids. Part XIII

[‡] Present Address: Research & Developement Wing, Amrutanjan Limited, Madras - 600 004, INDIA.

with Cava's synthetic sample. dl-Caseadine (m.p. 90-93°) was also synthesised by Kametani et al. who found that the O-methyl ether was identical with the methyl ether of natural caseadine;

Scheme 1

$$H_3$$
COOH H_3

- (2) R₁=H; R₂=OH; R₃=OCH₃
- (3) R₁= R₂=OCH₃; R₃= H

but the ir spectrum of synthetic caseadine differed from natural caseadine. They also reported that natural caseadine showed a negative result in Gibbs test, but the synthetic sample gave a positive response. Thus they concluded that caseadine should be represented by structure (5). But exceptions to Gibbs test are known; for eg., tetrahydroprotoberberine alkaloids having a C3-hydroxy group are known to give a positive reaction, although Gibbs test is usually negative for compounds having substituents at p-position to the phenolic group.

We synthesised dl-caseadine (1) according to scheme 2.

1-(3,4-Dimethoxybenzyl)-8-benzyloxy-7-methoxyisoquinoline (6)⁸
was catalytically reduced with Adams catalyst to the tetrahydroisoquinoline (7). Mannich cyclisation of this, according to

the procedure of Kametani et al.⁵, yielded dl-caseadine (1), $m \cdot p \cdot 90-92^{\circ}$.

Scheme 2

$$H_3$$
CO H_3 CO H_3 CO H_3 CO H_3 H_2 COC H_3 H_2 COC H_3 H_2 COC H_3 H_3

The 220 MHz nmr chemical shifts of natural and synthetic caseadine in CDCl $_3$ and $\rm C_6D_6$ are given in Table 1.

Table 1

	220 MHz N	MR Chemical	Shifts (8, ppm)
	Natural ca	seadine (1) C ₆ D ₆	Synthetic CDCl ₃	caseadine C6D6
OCH3	3.84 (s)	3.21 (s)	3.85 (s)	3.22 (s)
	3.85 (s)	3.39 (s)	3.86 (s)	3.39 (s)
	3.87 (s)	3.49 (s)	3.89 (s)	3.49 (s)
Cl3a-H	4.10 (q) (J = 12 an	4.29 (q) d 4 Hz)	4.15 (q) (J=12 an	4.36 (q) d 4 Hz)
C9 and C12-H	6.58 (s)	6.42 (s)	6.59 (s)	6.39 (s)
	6.61 (s)	6.53 (s)	6.61 (s)	6.48 (s)
С3-Н	6.66 (d) (J=8	6.44 (d) Hz)	6.67 (d) (j=8	6.43 (d) Hz)
C4-H	6.76 (d) (J=8	6.62 (d) Hz)	6.77 (d) (J=8	6.60 (d) H z)

The angular proton at Clas of trans fused benzo [a] - and indolo[a]-quinolizidines resonates at a field higher than δ 3.80, whereas the cis fused compounds are characterised by a downfield signal below 8 3.80 for this proton. The nmr spectrum of both synthetic and natural compounds showed the angular proton at δ 4.15 and 4.10 ppm respectively, in CDCL, and at & 4.36 and 4.29 ppm in CoDo, as a quartet (J=12 and 4 Hz). This indicates that both the synthetic and natural compounds may be cis fused. This proton was not identified in the natural alkaloid, but the presence of transquinolizidine nucleus was inferred only on the basis of the presence of Bohlmann bands in its ir spectrum. Kametani et al. and Iida et al. reported the nmr spectral data for their synthetic compounds, but no mention has been made regarding configuration. The signal at & 4.05 (in CDCl3) was however assigned to the angular proton. The ir spectra of our synthetic sample and natural compound have been compared in solution (CHCl3; Figure 1) and the

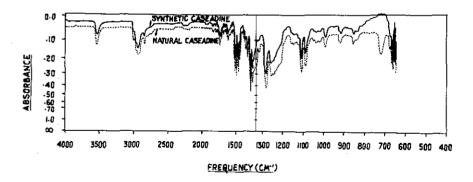


Figure 1

only difference is the presence of a band at 720 cm-1 in the ir spectrum of the natural compound. On the other hand, as is seen from Table 1, chemical shifts of various protons of natural caseadine and the synthetic compound are very close, especially those of the methoxy groups which are particularly identical. It appears somewhat doubtful whether structure (5) with a methoxyl at C1 can account for this observation. In our opinion the synthetic and natural compounds are identical. The minor differences between the two may be due to the contamination of some other compound in the natural caseadine. Thus the assigned structure to caseadine seems to be valid; it is however worthwhile to establish the structure and stereochemistry by X-ray diffraction studies; it would be most interesting if 2-hydroxy-1,10,11-trimethoxyberbine could be synthesised. All our attempts to synthesise this compound proved infructuous. Acknowledgement

The authors thank the Council of Scientific and Industrial Research, New Delhi, India for a post-doctoral fellowship to M.S. Premila and Junior and Senior Fellowships to H. Suguna. We are deeply indebted to Professor D.B. MacLean and Professor R.H.F. Manske who ran the 220 MHz nmr spectra and the ir spectra of both our synthetic sample and the natural caseadine and made valuable comments.

References

- 1 R.H.F. Manske and M.R. Miller, Canad. J. Research, 1938,
 16B, 153; C. -Y. Chen, D.B. MacLean and R.H.F. Manske,
 Tetrahedron Letters, 1968, 349; C.K. Yu, D.B. MacLean,
 R.G.A. Rodrigo and R.H.F. Manske, Canad. J. Chem.,
 1971, 49, 124.
- 2 M.P. Cava, M.V. Lakshmikantham and M.J. Mitchell, <u>J. Org.</u> Chem., 1969, 34, 2665.
- 3 S. Ishiwata and K. Itakura, Chem. and Pharm. Bull., 1970, 18, 1846.
- 4 H. Iida, H.C. Hsu, H. Miyano and T. Kikuchi, J. Pharm. Soc. Japan, 1971, 91, 795.
- 5 T. Kametani, T. Nakano, K. Shishido and K. Fukumoto, <u>J. Chem.</u> Soc.(C), 1971, 3350.
- 6 H.D. Gibbs, J. Biol. Chem., 1927, 72, 649.
- 7 H. Inouye, Y. Kanaya and Y. Naruto, Chem. and Pharm. Bull., 1959, 7, 573
- 8 F.R. Stermitz and D.K. Williams, J. Org. Chem., 1973, 38, 1761
- 9 M. Uskokovic, H. Bruderer, C. von Planta, T. Williams and
 A. Brossi, J. Amer. Chem. Soc., 1964, 26, 3364; T. Kametani,
 K. Fukumoto, M. Thara, A. Ujiie and H. Koizumi, J. Org.
 Chem., 1975, 40, 3280.
- 10 F. Bohlmann, Angew. Chem., 1957, 69, 641; Chem. Ber., 1958, 91, 2157.

Received, 18th July, 1977