

INTERMEDIACY OF 3',4'-DEHYDROVINBLASTINE IN THE
BIOSYNTHESIS OF VINBLASTINE-TYPE ALKALOIDS.

Kenneth L. Stuart,

Department of Chemistry, University of the West Indies,

Kingston 7, Jamaica.

and

James P. Kutney, Toshio Honda and Brian R. Worth,

Department of Chemistry, University of British Columbia,

2075 Wesbrook Place, Vancouver, B.C.

Canada, V6T 1W5

Cell free extracts from Catharanthus roseus plants have been used to demonstrate the formation of 3',4'-dehydrovinblastine and leurosine from radiolabelled catharanthine and vindoline. 3',4'-Dehydrovinblastine has been established as a precursor to the vinblastine group of alkaloids and a biosynthetic pathway is proposed.

Investigations of the biosynthetic pathway(s) to vinblastine (I) and related compounds have provided a source of both curiosity and understanding of the behaviour and mode of formation of these complex molecules. Even in recent work, much of the biosynthetic investigation of alkaloid biogenesis has been plagued by the inconclusive nature of very low incorporations of radiolabelled precursors. The use of plant derived tissue cultures enabled Zenk and Stöckigt¹⁻⁵ to demonstrate alkaloid interrelationships as high as 71% while, earlier, Scott and co-workers⁶⁻⁸ achieved successful results with cell free extracts from calluses and young plants. Recent efforts in these laboratories, using cell free extracts from mature Catharanthus roseus plants, have enabled similar, efficient, transformations⁹⁻¹¹. Although the exact meaning of precursor incorporation must always be evaluated with caution, the present in vitro work provides good insight into the probable, gross biosynthetic derivation of the vinblastine-type alkaloids.

A recent report¹² described the use of apical cuttings of C. roseus to give an absolute incorporation of catharanthine (II) and vindoline (III) into vinblastine (I) to the extent of 0.006%. In view of these and similar low incorporations with intact plants^{13,14}, our approach has been to test the validity of various segments of a proposed biosynthetic pathway to I (Figures 1 and 2). Exploitation of cell free extracts from C. roseus plants demonstrated significant incorporation of tryptamine into vindoline (III) (1.36%)⁹, and of a proposed, later stage intermediate (IV) into the natural products leurosine (V) (8.22%), catharine (VI) (15.15%) and vinblastine (I) (1.84%)^{10,11}.

Figure 1.

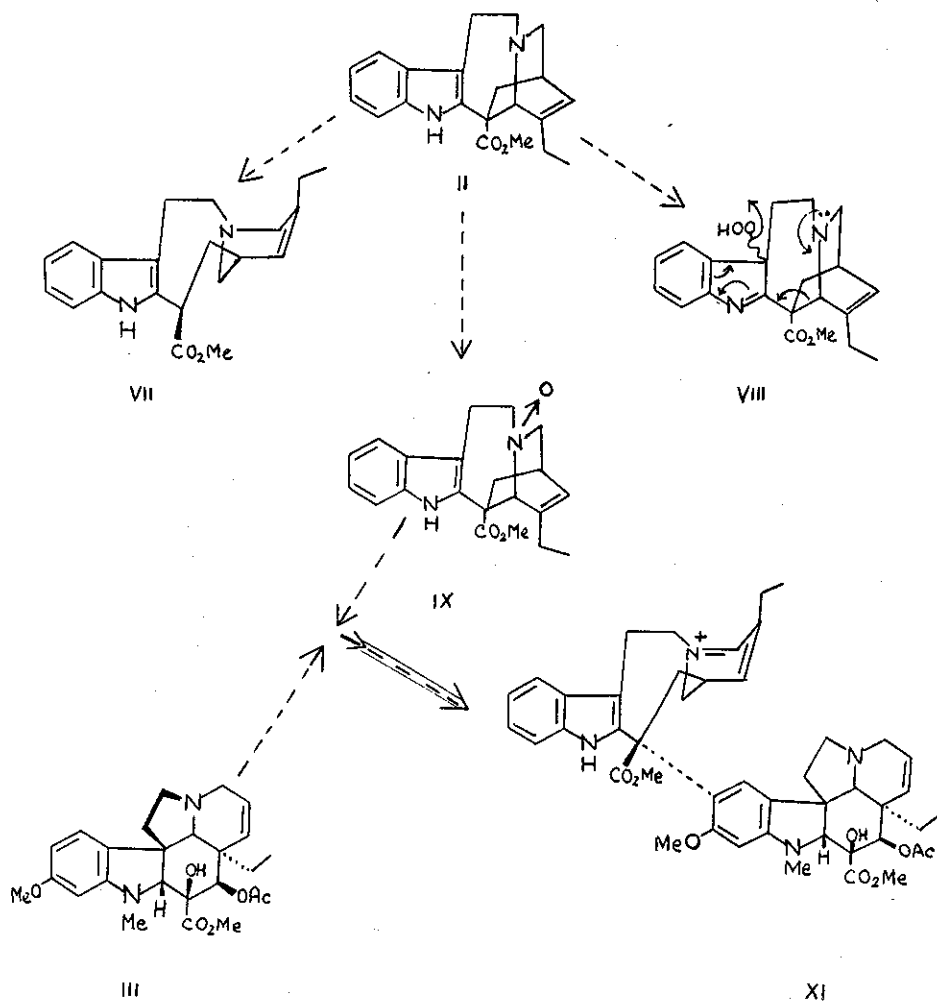
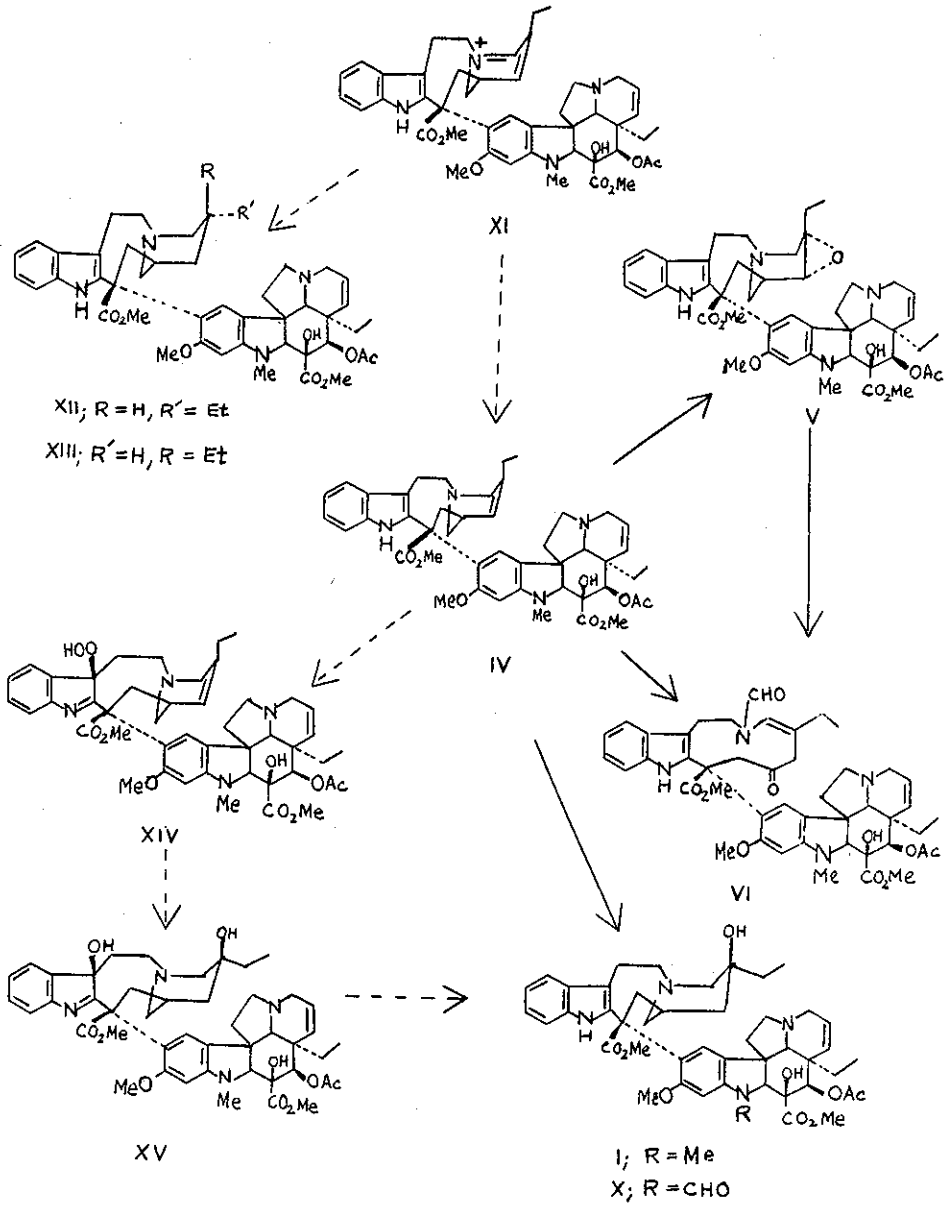
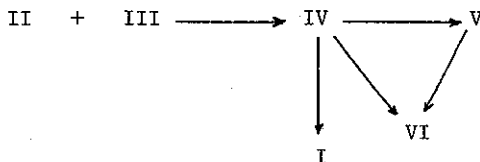


Figure 2.



The present work describes the enzyme catalysed formation of 3',4'-dehydrovinblastine (IV) from naturally occurring alkaloids catharanthine (II) and vindoline (III), together with the formation of leurosine (V). Here the use of cell free extracts has simplified the methodology and, as quoted in Table 1, necessitated only short reaction times to give satisfying results. In fact, [Ar-³H]-catharanthine was transformed to labelled (IV) to the extent of ca 0.54%, and to the alkaloid leurosine (V) in ca 0.36% yield. Furthermore, simultaneous incubation of [Ar-³H]-II and [Ac-¹⁴C] vindoline (III) afforded doubly labelled IV and V. The ³H/¹⁴C ratio of an equimolar ratio of the substrates was 23.6 while that of the products was 20.8 for IV and 21.1 for V; in good agreement with the obvious stoichiometry of the reaction. These results, together with the transformations of IV to the alkaloids I, V and VI, strongly support the gross biogenesis of "bisindole" alkaloids as:



Although no conclusive, direct evidence is available for the exact route of each transformation, plausible suggestions for various intermediates are given in Figures 1 and 2. It is generally accepted that a precursor with the cleavamine skeleton, such as VII, is not involved in the biosynthesis since related monomeric compounds are not found in the plant. Moreover, initial cleavage of an indole-3-substituted catharanthine, such as VIII, would also be expected to

TABLE I

Exp	Substrate		Product		% Incorporation	
	catharanthine	vindoline	anhydro- vinblastine	leurosine	anhydro- vinblastine	leurosine
A	1.667 x 10 ¹⁰ ^a (1.791 x 10 ⁸) ^b		8.563 x 10 ⁵ ^b	1.857 x 10 ⁵ ^b	0.48 ^e	0.10 ^e
B	1.667 x 10 ¹⁰ ^a (1.671 x 10 ⁸) ^b		8.955 x 10 ⁵ ^b	5.925 x 10 ⁵ ^b	0.54 ^e	0.36 ^e
C	1.667 x 10 ¹⁰ ^a (1.748 x 10 ⁸) ^b	7.077 x 10 ⁸ ^c (9.451 x 10 ⁶) ^d	6.201 x 10 ⁵ ^b 2.979 x 10 ⁴ ^d	5.936 x 10 ⁵ ^b 2.812 x 10 ⁴ ^d	0.36 ^e 0.32 ^f	0.34 ^e 0.30 ^f

Exp. A = 3 hrs; Exp. B = 8 hrs; Exp. C = 6 hrs.

a = dpm ³H/nmole; b = dpm ³H; c = dpm ¹⁴C/nmole; d = dpm ¹⁴C;

e = % incorporation for ³H; f = % incorporation for ¹⁴C

yield monomeric alkaloids, e.g. VII. Intermediacy of the N-oxide IX in a biological equivalent of the modified Polonovski reaction (used for the synthesis of IV) seems highly probable. Feeding of labelled IX to C. roseus plants has been carried out in these laboratories¹⁴, but the specific incorporation of only 0.008% into vincristine (X) was considered insignificant. However, the actual use of an intermediate such as IX on coupling with III could lead to the iminium species XI which on enzymic reduction, using NADPH as co-factor, would give IV. Similar, conjugate reduction could account for the alkaloids 4'-deoxyvinblastine (XII) and 4'-deoxyleurosidine (XIII).

The formation of I, V and VI from IV has already been demonstrated, but significant incorporation of IV into vincristine (X) was not shown¹¹. Several indirect routes to X can be proposed and one of these, involving initial N-demethylation and subsequent formylation gains some credence from the reported cleavage/oxidation of tertiary aromatic amines by peroxidase enzymes¹⁵. An interesting proposal for the elaboration of vinblastine (I) is shown in Figure 2. Molecular models show that the hydroperoxyindolenine (XIV) is ideally oriented for conversion to XV which would represent an immediate precursor to vinblastine. A discussion of the synthetic application of this route will be reported elsewhere.

Thus the use of cell free extracts of C. roseus plants has enabled a demonstration of the probable biogenesis of vinblastine-type alkaloids from catharanthine and vindoline, via 3',4'-dehydrovinblastine (or the corresponding iminium species XI) a late stage, pivotal inter-

mediate. The synthetic utility of enzyme mixtures, such as those used here, on solid support is currently under investigation (KLS).

Acknowledgements:

The International Development Research Centre is gratefully thanked for a Research Associateship to one of us (KLS). Financial assistance from the National Research Council of Canada is acknowledged, and we thank Professor Neil Towers, Botany Department, UBC, for use of research facilities. We are indebted to Dr. P.J. Salisbury, Chemistry Department, UBC for the plants used in these studies.

References

1. J. Stöckigt, J. Treimer and M.H. Zenk, FEBS Letters, **70**, 267 (1976).
2. J. Stöckigt, H.P. Husson, C. Kan-Fan and M.H. Zenk, J.C.S. Chem. Comm., 164 (1977).
3. J. Stöckigt and M.H. Zenk, FEBS Letters, **79**, 233 (1977).
4. J. Stöckigt and M.H. Zenk, J.C.S. Chem. Comm., 646 (1977).
5. J.F. Treimer and M.H. Zenk, Phytochemistry, **17**, 227 (1978).
6. A.I. Scott and S-L. Lee, J. Amer. Chem. Soc., **97**, 6906 (1975).
7. A.I. Scott, S-L. Lee and W. Wan, Heterocycles, **6**, 1552 (1977).
8. A.I. Scott, S-L. Lee and W. Wan, Biochem. Biophys. Res. Comm., **75**, 1004 (1977).
9. K.L. Stuart, J.P. Kutney, T. Honda, N.G. Lewis and B.R. Worth, Heterocycles, **9**, 647 (1978).
10. K.L. Stuart, J.P. Kutney and B.R. Worth, Heterocycles, **10**, in press.

11. K.L. Stuart, J.P. Kutney, T. Honda and B.R. Worth, Heterocycles, submitted.
12. S.B. Hassam and C.R. Hutchinson, Tetrahedron Letters, 1681 (1978).
13. A.I. Scott, Bioorg. Chem., 3, 398 (1974).
14. N.G. Lewis, Ph.D. Thesis, University of British Columbia, 1978.
15. G. Galliani, B. Rindore and A. Marchesini, J.C.S. Perkin I, 456 (1978).

Received, 24th July, 1978