(-)-DEHYDRONORCHELIDONINE AND (-)-ISODIDEHYDROCHELIDONINE,
TWO PROBABLE BIOGENETIC PRECURSORS IN THE BENZOPHENANTHRIDINE SERIES OF ISOQUINOLINE ALKALOIDS

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Abstract- The preparation of the title compounds 2 and 5 is described. Their probable biogenetic role is suggested on the basis of chemical evidence obtained by oxidation studies with a one electron oxidant.

The biogenesis of N-methylated benzophenanthridines such as chelidonine and sanguinarine has recently been established in Chelidonium majus by Battersby et al. 1 Their well supported conclusions come from incorporation of adequate precursors. However, to our knowledge there is no parallel study concerning the origin of norbenzophenanthridines, e.g. norchelidonine 1 and luguine 3. We have found that Fremy's salt (FS) oxidation of some aporphine and cularine alkaloids is accompanied by N-demethylation affording the corresponding oxoaporphines<sup>2</sup> and oxocularines.<sup>3</sup> To account for the observed results aminium radicals were proposed. This type of intermediates have also been suggested for oxidation of amines by the flavin-dependent enzyme MAO. 4 To investigate the likely "in vitro" formation of N-demethylated benzophenanthridines such as luguine 3 and norsanguinarine 7, FS<sup>5</sup> oxidation of norchelidonine 1, chelidonine 6, didehydrochelidonine 4b6 and the new compound isodidehydrochelidonine 5 was carried out. Treatment of (-)-norchelidonine 1 with FS under phase transfer conditions  $(CHCl_3/4%$  aq.  $NaCo_3$ , methyltrialkyl $(C_8-C_{10})$  ammonium chloride) for 6 h gave an 86% yield of (-)-dehydronorchelidonine 2. Longer reaction time (7 days) yielded the known (+)-luguine 3 in a 45% yield, identical with an authentic sample in all respects. 10 (+)-Luguine 3 easily dehydrates in acidic media to give norsanguinarine 7. On this basis it looks very likely that this latter compound was derived biogenetically from (-)-norchelidonine  $\underline{1}$ , via the sequence  $\underline{1} \longrightarrow \underline{2} \longrightarrow \underline{3} \longrightarrow 7$ . This is also supported by the fact that 1 and 3 are major products (0.13% and 0.04% respectively) in Glaucium flavum Cr. var. vestitum, 10 7 also being present although in minor proportions (0.002%).

The next target was the N-methylated series. (-)-Dehydronorchelidonine  $\underline{2}$  was treated with MeI in CHCl $_3$ , to give the corresponding (-)-methiodide  $\underline{4a}$  in a quantitative yield. Treatment of  $\underline{4a}$  with 10% NaOH gave (-)-didehydrochelidonine  $\underline{4b}$  having identical properties to those reported for the dextrorotatory compound, obtained earlier by permanganate oxidation of (+)-chelidonine. A new simple and effective alternative preparation of  $\underline{4b}$  is thus available. FS oxidation (Py/2% aq. Na $_2$ CO $_3$ ) of  $\underline{4b}$  gave (+)-luguine  $\underline{3}$  in 5% isolated yield among a very complex mixture of unidentified products.

In an attempt to obtain (-)-chelidonine  $\underline{6}$ , treatment of (-)-norchelidonine  $\underline{1}$  with formaldehyde followed by sodium borohydride led to the isolation by column chromatography of only a 5% yield of (-)-chelidonine 6,  $^{12}$  the main product being the new compound  $(-)-\underline{5}$ . The formation of  $(-)-\underline{5}^9$  was envisaged as the result of an internal Mannich reaction with the hydroxyl group at  $C_6$  on the basis of the following results: a) reaction of  $(-)-\underline{1}$  with formaldehyde at room temperature without added borohydride gave a 90% yield of  $(-)-\underline{5}$  as the only product isolated, b)  $(-)-\underline{5}$  was not reduced by borohydride at room temperature, c) upon refluxing with 98% formic acid or by treatment with NaCNBH3 at a controlled pH 3-4,  $(-)-\underline{5}$  gave (-)-chelidonine  $\underline{6}$  in quantitative yield. When  $\underline{6}$  was submitted to FS oxidation at room temperature, (-)-didehydrochelidonine  $\underline{4b}$  was isolated in a 60% yield. When the reaction was kept going for a longer time, only (+)-luguine could be

isolated in a low yield (5%). However, the above FS oxidation of (-)-isodidehydrochelidonine  $\underline{5}$  at room temperature for 7 days led to the isolation of (+)-luguine  $\underline{3}$  in 27% yield, along with 22% of starting material.

While no conclusive biogenetic pathways can be proposed on the basis of the above results, they show that in the norbenzophenanthridine series the oxidative conversions 1 - 2 - 3 are easily carried out using a one electron oxidant such as Fremy's salt, perhaps, mimetizing the natural process. On the other hand, while the oxidation of benzophenanthridines 4b and 6 to luguine 3 is a low yield process, probably due to competitive reactions, that of isodidehydrochelidonine 5 gives a fair yield of luguine 3. The above results might therefore be considered of some relevance for future incorporation studies on the biogenetic origin of norbenzophenanthridines.

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  - $\begin{array}{c} \underline{\text{(c:0.1,EtoH)}} & \underline{\text{4a: Yellow needles, mp 242-244°C (isopropanol); }} & |\alpha|_D^{20} & -333° \\ \hline \\ \underline{\text{(c:0.1,EtoH); }} & \lambda_{\text{max}}^{\text{EtOH}} : 234, 300 \text{ and } 398 \text{ nm; }} & \lambda_{\text{max}}^{\text{EtOH+Na}} & 2^{\text{CO}} & 3^{4\%} : 228 \text{ and } 290 \text{ nm; }} \\ \\ \underline{\text{max}} & \underline{\text{Notation of the large states of the large$

 $v_{\text{max}}$  (KBr): 3300, 1650, 1490, 1460 and 1250 cm<sup>-1</sup>;  $\delta$  (CDCl<sub>3</sub>+TFA-d<sub>1</sub>):8.93 (broad s, 1H,H-8), 7.11 and 6.87 (AB<sub>q</sub>,J=8,1H each,H-11 and H-12), 6.81 (s,1H,H-1), 6.69 (s, 1H,H-4), 6.22 and 6.15 (AB<sub>q</sub>,J=1,2H,O-CH<sub>2</sub>-O), 6.00 (s,2H,O-CH<sub>2</sub>-O), 5.22 (m,1H,H-14), 4.45 (m,1H,H-6), 3.48 (s,3H,N<sup>+</sup>-Me), 3.43 (m,1H,H-13) and 3.18 (d,J=2.8,2H,H-5).

(-)-Isodidehydrochelidonine 5: White plates, mp 178-180°C (EtoH);  $|\alpha|_D^{20}$  -156° (c:0.025,EtoH);  $\lambda_{\text{max}}^{\text{EtoH}}(\log \varepsilon)$ : 211(4.33), 238(3.89) and 290(3.86) nm;  $\nu_{\text{max}}(\text{KBr})$ : 2900, 1510, 1490, 1460, 1390, 1360 and 1340 cm<sup>-1</sup>;  $\delta(\text{CDCl}_3)$ : 6.73(s,1H,H-1), 6.73 and 6.60(AB<sub>q</sub>,J=8,2H,H-11 and H-12), 6.68(s,1H,H-4), 5.98 and 5.94(AB<sub>q</sub>,J=1.5,2H,O-CH<sub>2</sub>-O), 5.92(s,2H,O-CH<sub>2</sub>-O), 4.79 and 4.19(AB<sub>q</sub>,J=11,2H,N-CH<sub>2</sub>-O), 4.49 and 4.20(AB<sub>q</sub>,J=18,2H,H-8),4.04(m,1H,H-6), 3.96(d,J=2.6,1H,H-14), 3.25 (d,J=3,2H,H-5) and 2.67(m,1H,H-13) ppm; CMR  $\delta(\text{CDCl}_3)$ : 77.76(t)(N-CH<sub>2</sub>-O) ppm; m/e (%): 351(M<sup>+</sup>,80), 323(13), 322(27), 308(8), 306(8), 293(17), 235(13), 176(26), 175(43), 174(32) and 148(100).

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