CONSTITUENTS OF WEST AFRICAN MEDICINAL PLANTS XVIII¹: THE ANNONIDINES - A NEW CLASS OF PRENYLATED BISINDOLE ALKALOIDS FROM <u>ANNONIDIUM</u> <u>MANNII</u>

Hans Achenbach * and Christian Renner

Institute of Pharmacy, Department of Pharmaceutical Chemistry, University of Erlangen, D-8520 Erlangen

<u>Abstract</u> — From the stem bark of <u>Annonidium mannii</u> annonidines A - E (3 - 7) have been isolated and their structures determined. **3** to **7** belong to a new structural type of prenylated bisindole alkaloids.

The stem bark of the West African <u>Annonidium mannii</u> Engl. & Diels (Annonaceae)² was extracted with petroleum ether; on concentration large quantities of polycarpol $(1)^3$ precipitated and were removed. The better soluble constituents were subjected to column chromatography, and this yielded 7-(3-methyl-2-butenyl)-indole $(2)^4$ and the hitherto unknown prenylated bisindole alkaloids 3 to 7. Since these compounds constitute a new group of bisindole alkaloids they were named annonidines.













7: R = 7-indolyl

4: R = 3-indolyl

-2075 -

Structure determination is based on 1 H- (at 400 MHz) and 13 C-NMR measurements and on UV- and MSinvestigations of the original alkaloids and partly their hydrogenation derivatives. The spectra show, that in all compounds two indole systems are present and these are linked together by an isoprenoid C_{s} -unit. In annonidines A and B (3 and 4) both indoles are connected to C-1 of a 3-methyl-2-butene, whereas in annonidines C to E (5 to 7) the indoles are bound at C-1 and C-3 of a 3-methyl-l-butene. Another 3-methyl-2-butenyl unit is observed in 3 to 6 as a substituent of one of the indole nuclei. This $C_{\sf S}$ -substituent must be localized at C-7 of the indole; besides other arguments evidence comes from the 13 C-NMR resonances of the methylene carbon atoms which appear at δ = 31 ppm; in case of substitution at C-3 δ = 24 ppm and at C-6 δ = 35 ppm would be typical values 5 . From analysis of the 13 C-signals of the indole carbon atoms the other positions of substitution can be determined and this is particularly easy, if C-3 or C-7 are involved, since the resonances of these carbon atoms if unsubstituted appear typically at highest field (δ $_{\Gamma-3}$ = 103 ppm; $\delta_{\Gamma_{-7}} = 111 \text{ ppm}$)^{5,6}. Only annonidine D (6) contains an indole substituted at C-6 and this was deduced from the 1 H-NMR and a singlet at δ = 132.5 ppm in the 13 C-NMR. The data mentioned above allow to establish structures 3 and 4 for annonidines A and B unambiguously; but for annomidines C and D (5 and 6) still exist the alternative formulae 5a and 6a. Therefore 5 and 6 were hydrogenated to give their tetrahydro derivatives 8 and 9 , whose MS - fragmentation exhibit key fragments at m/z 228 and m/z 130 and thus exclude structures 5a and 6a.



The 1 H- and 13 C-NMR of annohidine E (7) show close similarity with the spectra of 5 for all signals except for C-7 of the disubstituted indole nucleus and the C₅-unit attached to that carbon atom. However, the resonances of this substituent and all other data fit completely to an epoxidized prenyl group⁵ and therefore establish formula 7 for annohidine E.

From a biogenetic point of view 3, 5 and 7 can be regarded as dimeric 7-prenylindoles (dimerization products of 2), whereas for the formation of 4 and 6 various pathways might be possible. Polycarpol (1) has exclusively be isolated from some plants of the Annonaceae family⁷; 7-(3-methyl-2-butenyl)-indole (2) up-to-now has been only described from the liverworts <u>Riccardia</u> <u>sinuata</u> and <u>R. incurvata⁴</u>.

EXPERIMENTAL

Melting points were determined on a Kofler hot stage apparatus and are uncorrected. Optical rotations were measured in CHCl_3 ; IR in KBr, UV in EtOH. NMR spectra were recorded in CDCl_3 using tetramethylsilane as the internal reference on a Jeol GX-400 spectrometer (¹H at 400 MHz, if not stated otherwise) and on a Jeol FX-90 Q instrument (¹H at 90 MHz and ¹³C at 22.5 MHz); compounds 3 to 9 were numbered in the following manner: C-2 to C-7a for the disubstituted indole, C-1' to C-5' for the substituent at C-7, C-2" to C-7a" for the monosubstituted indole and C-1" to C-5" for the connecting substituent. Mass spectra were obtained at 70 eV on a Finnigan 4000 instrument and on a Varian MAT 311 A spectrometer (high resolution MS data). Ready made Nano-plates SIL-20 UV₂₅₄ (Macherey-Nagel) were used for TLC; detection by spraying with anisaldehyde (reagent No. 15 according to Stahl⁸); standard solvent system was petroleum ether (bp 65-70°C)/ethyl acetate (9:1).

<u>Plant material</u> <u>Annonidium mannii</u> was collected at Bobiri Forest Reserve (near Kumasi, Ghana) in November 1984 by Mr. A. A. Enti (Forestry Enterprises, Accra, Ghana). A voucher specimen (No. 8407) is deposited in our herbarium.

<u>Extraction</u> and <u>separation</u> — Dried ground stem bark (2 kg) was percolated at 20°C with 35 1 of petroleum ether (bp 65-70°C). The solvent was evaporated under reduced pressure (220 hPa; water bath temperature 40-50°C). During concentration of the extract to 300 ml a precipitate was formed, which was filtered off to give 7.5 g of crude polycarpol (1).

The filtrate was evaporated further yielding an oily red residue (5 g), which was subjected to column chromatography on silica gel 60 (0.04-0.063 mm, Macherey-Nagel). Elution with petroleum ether (bp 65-70°C)/ethyl acetate (increasing the concentration of ethyl acetate from 10 to 90%) gave fractions P₁ to P₁₉. Fraction P₃ (350 mg), P₆ (220 mg), P₈ (240 mg), P₁₀ (135 mg), P₁₂ (410 mg) and P₁₄ (62 mg) were separated by column chromatography on Fractogel PVA 500 (Merck) using either methanol or ethanol as the eluent and this yielded the pure compounds 2 to 7.

 $\frac{7-(3-\text{Methyl-2-butenyl})-\text{indole}}{(15\text{ g} \text{ each, ethanol})} (2) - Chromatography of fraction P_3 (3 x 115 mg) on PVA 500 (15 g each, ethanol) yielded crystalline 2 (240 mg), mp 43-44°C (1it.⁴: mp <math>\approx 20°$ C). TLC: Rf = 0.37, anisaldehyde: orange; IR: $v_{\text{max}} = 3390$ (NH), 1435, 792, 728 cm⁻¹; UV: $\lambda_{\text{max}}(\log \epsilon) = 219$ (4.59), 270 (3.87), 288 nm (3.68); MS: m/z (rel. int.) = 185 (100%, M⁺), 170 (99), 155 (22), 130 (77), 117 (29); ¹H-NMR (90 MHz): δ (ppm) = 8.12 (br s; 1H, NH), 7.51 (m; 1H, 4-H), 7.18 (dd, J_1 = 3.2 Hz, J_2 = 2.5 Hz; 1H, 2-H), 7.02 (m; 2H, 5-H and 6-H), 6.56 (dd, J_1 = 3.2 Hz, J_2 = 2.2 Hz; 1H, 3-H), 5.42 (tqq, J_1 = 7.1 Hz, J_2 = J_3 = 1.5 Hz; 1H, =CH-CH₂), 3.57 (br d, J = 7 Hz; 2H, =CH-CH₂), 1.81 and 1.78 (2 x s; 6H, =C(CH₃)₂); ¹³C-NMR: δ = 135.0 (s; C-7a), 133.1 (s; C-3'), 127.8 (s; C-3a), 123.9 (s; C-7), 123.8 (d; C-2), 122.1 (d; C-2'), 121.3, 119.9 and 118.6 (3 x d; C-6, C-5, C-4), 102.8 (d; C-3), 30.5 (t; C-1'), 25.6 (q; C-4'), 17.8 (q; C-5').

Annonidine A = 3-[1-(7-Indoly1)-3-methy1-2-buteny1]-7-(3-methy1-2-buteny1)-indole (3)

Chromatography of fraction P_6 (2 x 110 mg) on PVA 500 (15 g each, methanol) afforded 3 (38 mg), which was crystallized from petroleum ether/ethyl acetate to give colorless needles, mp 106-108°C; $[\alpha]_D^{20} \pm 0^\circ$ (c = 1.0). TLC: Rf = 0.23, anisaldehyde: orange-red; UV: $\lambda_{max}(\log \varepsilon) = 221$ (4.85), 280 (4.20), 289 nm (sh 4.12); MS: m/z (rel. int.) = 368.2253 (98%, M⁺; $C_{26}H_{28}N_2$), 353 (21), 184 (29), 168 (100); ¹H-NMR: δ = 8.00 (br s; 1H, NH), 7.95 (br s; 1H, NH), 7.53 (d, J = 7.8 Hz; 1H, 4"-H), 7.29 (d, J = 7.3 Hz; 1H, 4-H), 7.13 (d, J = 7.2 Hz; 1H, 6"-H), 7.07 (t, J = 7.5 Hz; 1H, 5"-H), 7.02 (dd, J_1 = 3.2 Hz, J_2 = 2.4 Hz; 1H, 2"-H), 6.99 - 6.94 (m; 2H, 5-H and 6-H); 6.83 (dd, J_1 = 2.4 Hz, J_2 = 1 Hz; 1H, 2-H), 6.49 (dd, J_1 = 3.2 Hz, J_2 = 2.2 Hz; 1H, 3"-H), 5.76 (dqq, J_1 = 9.3 Hz, J_2 = J_3 = 1.4 Hz; 1H, =CH-CH), 5.41 (tqq, J_1 = 7.1 Hz, J_2 = J_3 = 1.4 Hz; 1H, =CH-CH_2), 5.38 (d, J = 9.3 Hz; 1H, =CH-CH_3); ¹³C-NMR: δ = 136.0 (s; C-7a), 134.7 (s; C-7a"), 133.2 (s; C-3'), 132.4 (s; C-3"), 128.2, 127.1 and 126.9 (3 x s; C-7", C-3a, C-3a"), 126.5 (d; C-2"'), 124.0 (s; C-7), 123.6 (d; C-2"), 122.1 (d; C-2'), 121.8, 121.7, 121.0, 119.9, 119.7 and 118.9 (6 x d), 118.9 (s; C-3), 117.7 (d), 102.5 (d; C-3"), 38.9 (d; C-1"), 30.6 (t; C-1'), 25.7 (q; C-4' and C-4"'), 18.1 and 17.9 (2 x q; C-5', C-5"').

Annonidine B = 3-[1-(3-Indoly1)-3-methyl-2-butenyl]-7-(3-methyl-2-butenyl)-indole (4)

Chromatography of fraction P_{10} (2 x 67 mg) on PVA 500 (15 g each, methanol) yielded 4 (11 mg). TLC: Rf = 0.13, anisaldehyde: yellow-orange; UV: $\lambda_{max}(\log \varepsilon) = 226$ (4.83), 283 (4.13), 291 nm (sh 4.08); MS: m/z (rel. int.) = 368 (100%, M⁺), 353 (47), 313 (18), 168 (15); ¹H-NMR: δ = 7.86 (br s; 1H, NH), 7.83 (br s; 1H, NH), 7.53 (d, J = 7.6 Hz; 1H, 4"-H), 7.40 (m; 1H, 4-H), 7.32 (d, J = 8.3 Hz; 1H, 7"-H), 7.15 (m; 1H, 6"-H or 5"-H), 7.03 (m; 1H, 5"-H or 6"-H), 6.97 (m; 2H, 5-H and 6-H), 6.88 and 6.86 (2 x dd, J₁= 2.2 Hz, J₂= 0.8 Hz; 2H, 2-H and 2"-H), 5.71 (dqq, J₁= 9.5 Hz, J₂= J₃= 1.5 Hz; 1H, =CH-CH), 5.41 (tqq, J₁= 7.1 Hz, J₂= J₃= 1.5 Hz; 1H, =CH-CH₂), 5.35 (d, J = 9.5 Hz; 1H, =CH-CH), 3.54 (d, J = 7.1 Hz, 2H, =CH-CH₂), 1.86 (d, J = 1.2 Hz; 3H, =C-CH₃), 1.80 (br s; 3H, =C-CH₃), 1.77 and 1.76 (2 x d, J = 1.2 Hz; 6H, =C-CH₃); ¹³C-NMR: δ = 136.8 and 136.0 (2 x s; C-7a, C-7a"), 133.0 (s; C-3"), 130.6 (s; C-3""), 128.0, 127.1 and 127.0 (2 x s; C-3a, C-3a"), 123.8 (s; C-7), 122.4 , 121.9, 121.7, 121.6, 121.3, 120.3 and 120.0 (2 x s; C-3, C-3"), 120.0, 119.2, 119.0, 118.0, 111.0 (C-7"), 33.4 (C-1"); 30.7 (C-1'), 25.7 (C-4' and C-4"), 18.0 and 17.9 (C-5', C-5").

Annoniding C = (E)-3-[3-(7-Indoly1)-1,1-dimethy1-2-propeny1]-7-(3-methy1-2-buteny1)-indole (5) Chromatography of fraction P₈ (2 x 120 mg) on PVA 500 (15 g each, methanol) gave 5 as a colorless oil (54 mg). TLC: Rf = 0.19, anisaldehyde: red; UV: $\lambda_{max}(\log \varepsilon) = 224$ (4.77), 238 (sh 4.51), 292 (4.22), 310 nm (sh 4.13); MS: m/z (rel. int.) = 368.2252 (100%, M⁺, C₂₆H₂₈N₂), 353 (47), 168 (68); ¹H-NMR: δ = 8.16 (br s; 1H, NH), 7.99 (br s; 1H, NH), 7.65 (m; 1H, 4-H), 7.50 (d, J = 7.8 Hz; 1H, 4-H"), 7.19 (d, J = 7.3 Hz; 1H, 6"-H), 7.08 (m; 3H), 6.99 (m; 2H), 6.67 and 6.55 (AB-system, J = 16.4 Hz; 2H, trans - CH=CH), 6.52 (dd, J₁= 3.2 Hz, J₂= 2.2 Hz; 1H, 3"-H), 5.44 (tqq, J₁= 7.1 Hz, J₂= J₃= 1.5 Hz; 1H, =CH-CH₂), 3.58 (d, J = 7.1 Hz; 2H, =CH-CH₂), 1.83 and 1.79 (2 x s; 6H, =C(CH₃)₂), 1.66 (s; 6H, >C(CH₃)₂); ¹³C-NMR: δ = 141.4 (d; C-2""), 136.4 (s; C-7a), 133.7 (s; C-7a"), 133.2 (s; C-3'), 128.3 and 126.1 (2 x s; C-3a, C-3a"), 124.4 and 124.2 (2 x s; C-7, C-3 or C-7"), 123.9 (d), 122.8 (d), 122.2 (d), 122.0 (s; C-7" or C-3), 121.3 (d), 120.0 (3 x d), 119.4 (2 x d), 119.3 (d), 102.9 (d; C-3"), 37.6 (s; C-1"), 30.7 (t; C-1'), 28.8 (q; C-4" and C-5"), 25.6 (q; C-4'), 17.9 (q; C-5').

<u>3-[3-(7-Indoly1)-1,1-dimethylpropyl]-7-(3-methylbutyl)-indole</u> (8)

5 (20 mg) was hydrogenated at 20°C on PtO_2 in ethanol. After purification by chromatography on PVA 500 (15 g, methanol) the product (15 mg) formed colorless crystals from petroleum ether, mp 118-119°C.

TLC: Rf = 0.26, anisaldehyde: orange-red; UV: $\lambda_{max}(\log \varepsilon) = 219$ (4.82), 280 (4.10), 288 nm (sh 4.02); M5: m/z (rel. int.) = 372 (21%, M⁺), 229 (32), 228 (100), 214 (5), 170 (8), 130 (29); ¹H-NMR (90 MHz): δ = 7.95 (br s; 1H, NH), 7.71 (m; 1H, 4-H), 7.40 (m; 1H, 4"-H), 7.33 - 6.83 (m; 6H), 6.71 (dd, J₁= 3.2 Hz, J₂= 2.4 Hz; 1H, 2"-H), 6.38 (dd, J₁= 3.2 Hz, J₂= 2 Hz; 1H, 3"-H), 2.86 (m, 2H), 2.63 - 2.06 (m; 4H), 1.67 (m; 3H), 1.51 (s; 6H, $\geq C(CH_3)_2$), 1.00 (d, J = 5.6 Hz; 6H, $CH(CH_3)_2$); ¹³C-NMR: δ = 136.6 (s; C-7a), 134.5 (s; C-7a"), 127.6 and 126.1 (2 x s; C-3a, C-3a"), 125.6 and 125.3 (2 x s; C-7 and C-7"), 124.6 (s; C-3), 123.5, 121.4 (two signals), 121.1, 119.7, 119.5, 118.7, 118.3, 102.4 (C-3"), 42.9 (C-2"), 38.8 (C-2'), 35.5 (s; C-1"), 29.0 (C-1'), 28.6 (C-4" and C-5"), 28.1 (C-3"), 27.6 (C-3"), 22.6 (C-4' and C-5').

Annonidine D = (E)-3-[3-(6-Indoly1)-1,1-dimethy1-2-propeny1] - 7-(3-methy1-2-buteny1)-indole (6)Fraction P₁₂ mainly consisted of polycarpol (1). Chromatography of the methanol-soluble part(65 mg) of this fraction on PVA 500 (15 g, methanol) afforded 1 (40 mg) and 6 (7 mg) as a colorless oil. TLC: Rf = 0.09, anisaldehyde: red-brown; UV: $\lambda_{max}(\log \varepsilon) = 225$ (4.75), 238 (sh 4.54), 247 (sh 4.49), 292 nm (4.41); MS: m/z (rel. int.) = 368 (100%, M⁺), 353 (53), 168 (33); ¹H-NMR: $\delta = 7.95$ (br s; 1H, NH), 7.92 (br s; 1H, NH), 7.63 (m; 1H, 4-H), 7.52 (d, J = 8.3 Hz; 1H, 4"-H), 7.23 (br s; 1H, 7"-H), 7.20 (dd, J₁= 8.3 Hz, J₂= 1.5 Hz; 1H, 5"-H), 7.11 (dd, J₁= 3.2 Hz, J₂= 2.4 Hz; 1H, 2"-H), 6.99 (d, J = 2.4 Hz; 1H, 2-H), 6.97 (m; 2H, 5-H and 6-H), 6.53 (AB-system, J = 16.5 Hz; 2H, trans - CH=CH), 6.47 (ddd, J₁= 3.2 Hz, J₂= 2.2 Hz, J₃= 1 Hz; 1H, 3"-H), 5.42 (tqq, J₁= 7.1 Hz, J₂= J₃= 1.5 Hz; 1H, =CH-CH₂), 3.55 (br d, J = 7 Hz; 2H, =CH-CH₂), 1.81 and 1.77 (2 x s; 6H, =C(CH₃)₂), 1.62 (s; 6H, >C(CH₃)₂); ¹³C-NMR: $\delta = 138.3$ (C-2"), 136.4 and 136.3 (2 x s; C-7a, C-7a"), 133.1 (s; C-3'), 122.3 (C-2'), 121.2, 120.5, 119.9, 119.6, 119.2, 118.7, 108.8 (C-7"), 102.6 (C-3"), 37.1 (s; C-1"), 30.7 (C-1'), 28.8 (C-4"" and C-5""), 25.6 (C-4'), 17.9 (C-5').

3-[3-(6-Indolyl)-1,1-dimethylpropyl]-7-(3-methylbutyl)-indole (9)

6 (2 mg) was hydrogenated at 20°C on PtO₂ in ethanol yielding a homogenous product (2 mg). TLC: Rf = 0.14, anisaldehyde: orange-red; UV: $\lambda_{max}(\log \epsilon) = 221$ (4.82), 281 (4.10), 291 nm (4.05); MS: m/z (rel. int.) = 372 (32%, M⁺), 229 (54), 228 (100), 214 (6), 170 (5), 130 (22); ¹H-NMR (90 MHz): $\delta = 7.88$ (br s; 2H, 2 x NH), 7.70 (m; 1H, 4-H), 7.48 (d, J = 8 Hz; 1H, 4"-H), 7.15 - 6.94 (m; 6H), 6.85 (dd, J₁= 8 Hz, J₂= 1.5 Hz; 1H, 5"-H), 6.46 (m; 1H, 3"-H), 2.82 (m; 2H), 2.62 - 2.06 (m; 4H), 1.66 (m; 3H), 1.50 (s; 6H, >C(CH_3)₂), 1.00 (d, J = 5.6 Hz; 6H, CH(CH_3)₂).

Annonidine E = (E)-3-[3-(7-Indoly1)-1,1-dimethyl-2-propenyl]-7-(3-methyl-2,3-epoxybutyl)-indole (7)

Fraction P_{14} was separated on PVA 500 (15 g, methanol). The fraction containing 7 was rechromatographed on the same column to give pure 7 (2 mg). TLC: Rf = 0.07, anisaldehyde: red; UV: $\lambda_{max}(\log \varepsilon) = 224$ (4.83), 238 (sh 4.53), 293 (4.27), 310 nm (sh 4.16); MS: m/z (rel. int.) = 384 (100%, M⁺), 369 (49), 313 (7), 297 (21), 182 (10), 168 (49), 167 (15), 130 (16); ¹H-NMR: δ = 9.07 (br s; 1H, NH), 8.19 (br s; 1H, NH), 7.69 (m; 1H, 4-H), 7.50 (d, J = 7.8 Hz; 1H, 4"-H), 7.20 (d, J = 7.3 Hz; 1H, 6"-H), 7.11 (m; 2H, 2-H and 2"-H), 7.07 (t, J = 7.6 Hz; 1H, 5"-H), 6.97 (m; 2H, 5-H and 6-H), 6.67 and 6.56 (AB-system, J = 16.2 Hz; 2H, trans - CH=CH), 6.52 (dd, J₁= 3.2 Hz, J₂= 2.0 Hz; 1H, 3"-H), 3.24 (dd, J₁= 14.8 Hz, J₂= 1.5 Hz; 1H, ⁰>CH=CH₂), 3.05 (dd, J₁= 9.3 Hz, J₂= 1.5 Hz; 1H) and 2.96 (dd, J₁= 14.8 Hz, J₂= 9.3 Hz; 1H, ⁰>CH=CH₂), 1.66 (s; 6H, >C(CH₃)₂), 1.52 and 1.39 (2 x s; 6H, ⁰>C(CH₃)₂); ¹³C-NMR: δ = 141.6 (C-2"), 136.8 (s; C-7a), 133.7 (s; C-7a"), 128.2 and 126.3 (2 x s; C-3a, C-3a"), 123.9 (s; C-3 or C-7"), 123.9, 122.8, 122.0, 122.0 and 121.8 (2 x s; C-7, C-7" or C-3), 120.7, 120.2, 120.1, 119.9, 119.4, 118.9, 102.9 (C-3"), 64.5 (C-2'), 59.8 (s; C-3'), 37.5 (s; C-1"), 33.5 (C-1'), 28.9 (C-4" and C-5"), 24.7 (C-4'), 18.9 (C-5').

ACKNOWLEDGEMENTS

This work was financially supported by the "Deutsche Forschungsgemeinschaft", the "Gesellschaft für Technische Zusammenarbeit (GIZ)" and the "Fonds der Chemischen Industrie".

REFERENCES

- 1. For part XVII, see H. Achenbach, R. Waibel and I. Addae-Mensah, Phytochemistry, in press.
- 2. F. R. Irvine, 'Woody Plants of Ghana', Oxford University Press, London, 1961, p. 5.
- M. Hammonière, A. Fournet, M. Leboeuf, A. Bouquet and A. Cavé, <u>C. R. Acad. Sci. Paris, Ser. C</u>, 1976, 282, 1045.
- 4. V. Benesová, Z. Samek, V. Herout and F. Sorm, Collect. Czech. Chem. Commun., 1969, 34, 1807.
- 5. H. Achenbach, C. Renner and I. Addae-Mensah, <u>Heterocycles</u>, 1984, 22, 2501.
- 6. R. G. Parker and J. D. Roberts, <u>J. Org. Chem.</u>, 1970, <u>35</u>, 996.
- M. Leboeuf, A. Cavé, P. K. Bhaumik, B. Mukherjee and R. Mukherjee, <u>Phytochemistry</u>, 1982, 21, 2783.
- Stahl, 'Dünnschichtchromatographie', 2nd. ed., Springer Verlag, Berlin-Heidelberg-New York, 1967, p. 817.
- C. M. Hasan, T. M. Healey, P. G. Waterman and C. H. Schwalbe, <u>J. Chem. Soc., Perkin Trans. I</u>, 1982, 2807.

Received, 14th May, 1985