ALKALOIDS OF HEDYCARYA ANGUSTIFOLIA

Y.A. Geewanda P. Gunawardana, Huck-Meng Leow, and I. Ralph C. Bick* Chemistry Department, University of Tasmania, Hobart, Tas. Australia, 7005

Abstract - Four new alkaloids, 6,6a-dehydronorlaureline (7), isosevanine (10), isouvariopsine (12), and O-methylcinnamolaurine (14) have been isolated from H. angustifolia, and their structures have been determined. The plant also contains the known aporphines corydine (1) , laurotetanine (2) , boldine (3) , glaucine (4) , and laureline (5) .

RESULTS AND DISCUSSION

Material from the monimiaceous plant Hedycarya angustifolia A. Cunn., which grows in rainforests of south-eastern Australia, was collected from King Island in Bass Strait, the southern limit of its distribution, and extracted by standard procedures. Small amounts of crude alkaloids were obtained, accompanied by considerable quantities of persistent non-basic impurities. These could not be completely removed by continuous extraction of the crude bases in dilute mineral acid over a prolonged period with ether; however, droplet countercurrent chromatography (dccc) 19 proved effective in removing the remaining neutral contaminants, and in partially separating the bases from one another. The individual alkaloids were finally isolated from the dood fractions and purified by preparative tlo.

The alkaloid content was found to be small and variable: the maximum amount obtained, from autumn collections in two separate years, was ca. 2 x 10^{-3} %, and material collected in winter from the same location furnished no alkaloids at all. Of the nine bases found, five were identified as known aporphines. The uv spectrum of one of the latter indicated that it was a 1,2,10,11-substituted aporphine¹, and this substitution pattern was supported by the $^1\rm H$ nmr spectrum which showed a doublet of doublets corresponding to the H-8 and H-9 protons. The latter spectrum also indicated the presence of an N-methyl group, and three methoxyls, all of which appeared to be too deshielded $2,3$ to be located at C-1. These data suggested that the alkaloid had structure 1, which was confirmed by comparison with an authentic sample of corydine⁴. Another alkaloid from the same dooc fraction turned out from $\mathrm{uv}^\mathbf{1}$ and 1 H nmr data to be a 1,2,9,10-substituted noraporphine; the $^{\mathrm{1}}$ H nmr spectrum in particular indicated the

presence of a highly deshielded aromatic proton corresponding to $H=11^3$, and three methoxyls of which one must be located at C-1 from its high chemical shift^{2,3}. The uv spectrum showed a prominent bathochromic shift above 300 nm on addition of alkali, typical of an aporphine with a phenolic group at $C-9^5$. Structure 2 suggested by these data was confirmed by comparison with an authentic sample of laurotetanine⁴.

Amongst the other aporphines obtained was an isomer of laurotetanine with the same 1,2,9,10 pattern of substitution including an hydroxyl group at C-9, as shown by the uv and 1 H

mur spectra; however, the latter spectrum also indicated the presence of an N-methyl group, and two methoxyls of which one must be attached at C-1 from the high chemical shift^{2,3} of its protons; this location was supported by the strong $(M-31)^{+}$ peak in the ms⁵. The spectroscopic evidence suggested that the compound was boldine $(3)^4$, and this was confirmed by comparison with an authentic sample.

An aporphine isolated from another docc fraction proved to have the same $1,2,9,10$ substitution pattern as the two above-mentioned bases, but it was found to be a higher homologue with four methoxyls and an N-methyl group. Structure 4 was confirmed by comparison with an authentic sample of q laucine⁴. The remaining aporphine had a uv spectrum that indicated 1,2,10-trisubstitution¹, and this was supported by the pattern of aromatic signals in the ¹H nmr spectrum. The latter also showed the presence of methylenedicxy, N-methyl, and methoxy groups; the evidence thus pointed to structure 5, which was confirmed by comparison with a sample of laureline⁴ prepared by 0-methylation of mecambroline $(6)^4$.

Another alkaloid with a methoxyl and a methylenedioxy group like laureline (5) did not, however, correspond with any previously reported base. The uv spectrum of the new alkaloid

 $-448-$

resembled that of 5, but it had an additional long-wavelength band around 330 nm, indicating the **1** presence of extra mnjugation. mihe H *m* **spednm** was also similar to that of **2,** with the **same** pattern of aromatic proton signals, but it gave no evidence of a methylimino group, and the mthylenedioxy **group** produced a sinqlet instead of a douhlet as for laureline6; this observation likewise suggested a greater degree of conjugation in the molecule, resulting in increased rigidity and closer alignment of the planes of the aromatic rings. Attempts to methylate the nitrogen with formaldehyde and borohydride failed, and the only product obtained had added on two hydrogens; however, the nitrogen was readily methylated and quaternised with methyl icdide to give a prodmt with only one nwre cartan. The original alkaloid is thus a tertiary **base,** and the above-mentioned data together with the molecular formula, which from ms corresponds to norlaureline with *two* less hydrogens, pointed to structure 7. That the compound was indeed 6.6-dehydronorlavreline was confirmed **by** the fad that **the quaternary** methicdide referred to above could be reduced with borohydride to laureline (5); it thus forms another member of the small but increasing sub-group of aporphine-type alkaloids with a conjugated azomethine $\arccos^7 r^8$. The methicdide of 7 appeared to undergo ready oxidation on exposure to air; the *uv* spectrum of the product resembled that of an oxoaporphine , and an additional downfield double doublet **1 appsarej** in the matic region of the **H** *m* spectrun (at 8.75 and 8.35 ppn, J=8 Hz). These signals suggested that ring B had been aromatised to produce a structure such as 8; analogousreactions in the aporphine series involving N-demethylation and aromatisation have been recorded,

7 6,6a-Dehydronorlaureline

$$
R_1-R_2=CH_2
$$
, $R_3=H$, $R_4=CH_3$

although their reaction mechanism is not fully understood 10 .

Amongst the previously unrecorded bases was one that from high resolution ms had the same nolecular formula as dehydronorlaureline (?) except for an extra oxygen, which appeared to **be** in an hydroxyl group from the ir spectrum. I ike 7, the new base had a methoxyl and a methylene-
dioxy group from the ¹H rmr spectrum, which also showed the presence of extra aromatic protons as

compared to the other alkaloids described above; two of these protons produced a double doublet at the low-field end of the aromatic region, and suggested the presence of an isoquinoline The uv spectrum was in good accord with a benzylisoguinoline structure, and in nucleus. general the observations indicated that the alkaloid was an isomer and close structural analogue of sevanine 11 (9). The remaining aromatic protons showed the same splitting pattern and similar chemical shifts as for sevanine in the 1 H nmr spectrum: positions 6,7,3', and 4' must thus be substituted in both cases. The new alkaloid, which has been named isosevanine, gave a positive Gibbs reaction¹², indicating that its phenolic group must have an unsubstituted para position, and must thus be attached at $C-3'$. The tentative structure 10 put forward on the basis of these data has been confirmed by X-ray crystallographic analysis 13 .

One of the glaucine fractions contained a previously undescribed base whose uv spectrum showed it to have a phenanthrene nucleus 14 . The 1 H nmr spectrum revealed the presence of a methylenedioxy, a methoxy, and a dimethylamino group; the latter evidently forms part of an ethanamino chain as indicated by the base peak at m/z 58 in the ms, together with a strong complementary ion at m/z 265, and also by a pair of signals in the $^{\mathrm{1}}$ H mmr spectrum corresponding to protons in two methylene groups that are attached to nitrogen and to an aromatic ring respectively. From biogenetic considerations, the ethanamine chain and the methylenedioxy

- 11 Uvariopsine $R_1=H$, $R_2=OCH_2$
- 12 Isouvariopsine $R_1 = OCH_3$, $R_2 = H$

15 α-Eudesmol

- R
- 13 Cinnamolaurine **R=H**
- 0-Methylcinnamolaurine 14 R=Me

ß-Eudesmol $16¹$

group may be tentatively assigned to positions 1,3 and 4 respectively of ring **A** in the 14 1 phenanthrene system . **A broad** daun-field singlet at 8.6 **ppn** in the H mu **spec-** can be attributed to the bay proton $H-5^{14}$, and the methoxyl group in consequence must be located at C-6. **The** resulting structure is closely analogous to that of the !amm alkaloid wariopsine **⁴** *(11)*, and the new alkaloid, which has been named isouvariopsine, was confirmed as having structure 12 by comparison with the methine base formed by Hofmann degradation of laureline **(2).**

Finally, an alkaloid that had not previously been reported was separated frm one of the laureline fractions; its formula, uv and mass spectra suggested that it belonged to the benzyltetrahydroisoquinoline group. From the ¹H mmr spectrum, the new base had a methylenedioxy, an N-methyl, **and** a methoxy **Trap.** Conplenentary and intense ions at m/z 190 and 121 in *the* **ms** pointed to the rnethylenedioxy grow king attached to ring **B** and the rnethoxy to ring C; the former group was located by the presence of two one-proton singlets in the aromatic region of **¹**the H rmr **spectrrnn** correspndinq to **H-5** and H-8, and **the** latter by the splitting pattern of the remaining aromatic protons, which corresponded to those of a p-disubstituted benzene ring. The **¹⁵**new alkaloid is thus the methyl ether of charmlaurine **U3). ard** its stmcture has teen confirmed as 14 by a comparison between the properties of the diazomethane reaction product of 12 and those of the new base, named O-methylcinnamolaurine. 13 and those of the new base, named 0-methylcinnamolaurine.

The persistent contaminants associated with the alkaloid fraction of the plant, which appaxed to alter the solubility relationships of the **bases** and render then difficult to separate and purify, were also investigated. It was difficult conversely to separate the contaminants £run the alkaloids **and** to purify then conpletely but this was eventually acconplished by dccc19 and by recrystallisation. **The** material **was** obtained in substantial amounts as a colourless crystalline substance that analysed for $C_{10}H_{18}O_2$. Since no double tends could te **detected by.ir** or 13c rmr **spedmscopy,** the nucleus was evidently bicyclic. The 1 H and 13 C nmr spectra indicated the presence of three methyl, two methylene, three methine and two quaternary carbons; furthermore, the ir and 13 C rmr spectra showed that the two oxygens were present as hydroxyl groups. Further studies designed to fix the structure and **stereochanistry** of this hiqclic terpenoid diol are mder way and will te reported elsewhere.

17 Elemol

18 Hedycaryol

Some volatile terpenoid material was also obtained from the solvent used in the initial extraction of the plant material: on evaporation of the methanol extract, the material distilled over with the solvent and eventually crystallised from it. The crystalline material was identified as a mixture of α - **(15)** and β - **(16)** eudesmols 16 by a comparison of the 1 H rmr, glc and **ms** data with **those** of authentic simples. Previous studies on the leaf oil of H. angustifolia resulted in the isolation of the sesquiterpene alcohols $elemol^{17}$ (17) and hedycaryol¹⁸ (18), and it was shown¹⁸ that the latter was the biogenetic precursor of 17 . It is likely that hedycaryol is related biosynthetically to the eudesmls 15 **and** 15 as well.

EXPERIMENTAL

Droplet countercurrent chromatography (dccc)¹⁹ was carried out on an instrument consisting of 100 glass tubes, each 4 mn in diameter and 1 mn long. **~thanol:chlorofonn:water** 15:5:3) were first equilibrated, and the lower layer was used as a stationary phase, while the upper layer, to which sufficient sulphuric acid was added to give a concentration of N/1000, was used as the mobile phase. Thin-layer chromatography (tlc) and preparative thin-layer chromatography (ptlc) were performed with Merck silica gel $GF_{25,4}$ or CAMAG silica gel DSF-5, and the α mpounds were visuallsed by spraying with iodoplatinate reagent or by exmination **under** w liaht. The rreltin? points (mp) were recorded on a Yanagimoto Seisakusho micro-melting point apparatus and are uncorrected. Ultraviolet (w) absorption spectra were recorded on methanol solutions with a ~itachi-perk-~lmer 124 spgtrapho-ter, **and** the logarithm of the extinction ccefficients are given in parentheses. Infrared spectra were recorded with a Beckman IR-33 spectrometer on chloroform solutions unless otherwise specified. Proton magnetic resonance $(^1\text{H}$ mm) spectra were recorded on deuterochloroform solutions unless otherwise snecified at 270 1Hz with a Bruker HX-270 spectrometer. Tetramethylsilane was used as the internal standard. Chemical shifts are given in **ppm** and the coupling constants in hertz (Hz). Peaks are described as singlets (s), doublets (d), triplets (t), quartets (q) or multiplets (m). Mass spectra were run on a Vacuum General micromass 7070 F spectrometer by the direct insertion technique at 200° and 70 eV. Intensities are given in parentheses as percentages of base peak intensity. htraction of plant naterial - **Iraves,** twigs **and** bark of Hedycarya angmtifolia collected **around** Little Grassy Creek, King Island (Map reference BR514678) in March 1978 were air-dried, milled to a fine powder (22.4 kg) and exhaustively extracted with methanol until a sample gave a negative alkaloid test with Mayer's reagent. The methanol extract was concentrated to 2 & in vaguo and dissolved in glacial acetic acid (1.5 ℓ). The solution was poured in a fine stream into rapidlystirred water (20 ℓ) and left overnight, then the non-alkaloid precipitate was filtered off through celite. The filtrate was evaporated to dryness in vacuo below 40° and the residue redissolved in 5% aqueous sulphuric acid. The acid solution was extracted with ether $(5 \times 100 \text{ ml})$, then basified with mmnia **and** extracted with chloroform (10 x 100 ml). The chloroform layer was dried (Na₂SO₄) and evaporated to dryness in vacuo to give the crude alkaloid extract as a yellow *amorphous* powder (12.5 g, 0.056%).

The ethereal extract on evaporation gave a pale yellow crystalline non-alkaloidal material (6 g). The methanol recovered from the initial extract deposited a somewhat volatile white $crystalline non-alkaloidal material (0.7 q).$

^Asecoid batch of plant material (25 kg) collected fmn the sarre locality in july 1979 failed to give any alkaloids when extracted by the same method, but a third batch (23 kg) collected in March the following year gave a further 5.9 g of crude alkaloid extract. The conbined alkaloid extracts were dissolved in chloroform and extracted with dilute hydrochloric acid (0.05%. 4 **x** 25 ml). **The aqueous** acid solution was exhaustively extracted with ether in a liquid/liquid extractor for 30 h, basified with ammonia (pH = 13) and extracted with chloroform (6 x 25 **ml)** . The canbind chlorofom so1"tions were dried and evaporated to give a residue of mixed alkaloids $(0.8 g, 1.76 \times 10^{-3}$ %).

kcc Separation of Mixed Alkaloids - The mixture, which frcm tlc contained at least **seven** alkaloids together with a considerable munt of nm-basic material, was **subjected** to droplet countercurrent chromatography 19 . The mobile phase as it emerged from the apparatus was monitored by a **w** detector (254 **mn),** and 127 **x** 3 ml aliwots were collected autcmatically. Every fifth aliquot was basified with mmnia and extracted with chloroform; each extract was examined by tlc, and the aliquots were bulked accordingly into nine subfractions. Methanol and chloroform were removed from each subfraction by careful evaporation under vacuum below 30°C, and the **aqueous** residues were hsified and extracted with chloroform, then the extracts were dried (Na_2SO_4) and evaporated to give nine partially purified alkaloid fractions, which were further separated **and** purified by ptlc.

Fraction 1: Corydine and Laurotetanine - The ptlc purification of the brown gum $(0.098 \sigma,$ aliquots 8-15) gave corydine (0.057 g), mp 148°C from MeOH/CHCl₃ (lit.⁴ 148°C), $\left[\alpha\right]_D^{20}$ + 204° $(C = 0.5, C_2H_5OH)$, (lit. 4 + 204°), λ_{max} 218 (4.19), 262 (3.73), 270 (3.70), 302 rm (3.40), **V-** 3450, 1590, 1570, 1500, 1480 &'; 'H rmr 6 7.07 (lH, d, **J** = 8 Hz), 6.83 (lH, d, **J** = 8 Hz), 6.70 (IH, s), 3.88 (6H, s, 2 x OCH₃), 3.73 (3H, s, OCH₃), 2.54 (3H, s, N-CH₃); ms m/z 341 (M^+ , 100%), 340 (85), 326 (68), 324 (55), 310 (48), 298 (40), 183 (30), 267 (42), 170.5 (M^{++} 10); identical with an authentic sample of corydine⁴; and laurotetanine (0.026 g), amorphous, Amax 220 (4.25). 280 (3.22). 308 rn (3.15), on addition Of OH-, 315 (3.35); **umax** 3350, 1580, 1500 cm⁻¹; ¹H mm \circ 6 7.88 (1H, s), 6.55 (2H, m), 3.83 (6H, s, 2 x α H₃), 3.52 (3H, s, α H₃); ms

 m/z 327 (M^+ , 68), 326 (100), 312 (26), 310 (18), 298 (12), 296 (21), 183 (8), 267 (10), 163.5 $(M⁺$, 8); identical with an authentic sample of laurotetanine⁴.

Fraction 2: Corydine and Boldine - Ptlc purification of this fraction (0.120 q, aliquots 16-17) gave corydine (0.035 g) and boldine (0.072 g), mp 161°C (lit.⁴ 161°C), [a]_D²⁰ + 108° (C = 1, C₂H_{_COH), (lit.⁴ + 111^o), λ_{max} 220 (4.6), 183 (4.21), 304 nm (4.23); ¹H nmr 6 (CD₃OD); 7.99 (1H,} s), 6.90 (1H, s), 6.60 (1H, s), 3.92 (3H, s, OCH₃), 3.61 (3H, s, OCH₃), 2.58 (3H, s, N-CH₃); ms m/z 327 $(M^+, 85)$, 326 (100), 312 (32), 310 (38), 296 (29), 184 (68), 269 (72), 253 (52), 163.5 (5); identical with an authentic sample of boldine⁴.

Fraction 3: Isosevanine - Ptlc of the gum $(0.055 g,$ aliquots 18-28) gave isosevanine $(0.016 g)$, colourless needles from methanol/chloroform mp 148°C; λ_{max} 236 (4.12), 270 (3.71), 314 (3.41), 330 mm (3.32); v_{max} 3400, 1640, 1610, 1580 cm⁻¹; ¹H mmr δ 8.28 (1H, d, J = 4.5 Hz), 7.36 (1H, d, $J = 4.5$ Hz), 7.35 (1H, s), 7.05 (1H, s), 6.7 (3H, m), 6.05 (2H, s), 4.45 (2H, s), 3.83 (3H, s, OCH₂); ms m/z 309 (M⁺, 60), 308 (100), 294 (23), 278 (8), 137 (11), 83 (56), 77 (22). Found: 309.0987; calculated for $C_{1.8}H_{1.5}NO_A$: 309.1017.

Fraction 4: Dehydronorlaureline and Laureline -- Ptlc purification of fraction 4 (0.085 q, aliguots 29-36) gave laureline (0.020 g) as a brown gum, λ_{max} 218 (4.20), 264 (3.95), 273 (4.1), 315 (3.57), 325 rm (3.25); ¹H rmx 6 7.66 (1H, d, J = 2.5 Hz), 7.15 (1H, d, J = 10 Hz), 6.75 (1H, d, J = 10 Hz), 6.53 (1H, s), 6.03 (1H, d, J = 1.5 Hz), 5.88 (1H, d, J = 1.5 Hz); ms m/z 309 $(\text{M}^{\dagger}, 65)$, 308 (100), 294 (53), 266 (48), which was identified by comparison with an authentic sample; and $6, 6a$ -dehydronorlaureline as a brown gum $(0.043 g)$, λ_{max} 248 (4.32) , 278 (4.08) , 317 (3.83) and 330 nm (3.85); v_{max} 1690, 1640 and 1600 cm⁻¹; ¹H nmr δ 7.35 (1H, s, 11-H), 7.2 (1H, s, 3-H), 7.1 (1H, d, J = 10 Hz, 9-H), 6.75 (1H, d, J = 10 Hz, 8-H), 6.1 (2H, s, O-CH₂-O), 4.49 (2H, m, 7-H), 3.7 (3H, s, OCH₂); ms m/z 293 (M⁺, 55), 292 (100), 288 (30), 262 (18), 249 (2), 149 (20), 146.5 (M⁺⁺, 2). Found: 293.1043; calculated for $C_{18}H_{15}NO_3$: 293.1052. Fraction 5: Laureline and O-Methylcinnamolaurine -- Ptlc purification of this fraction (0.05 g, aliquots 37-41) gave laureline, and a more polar compound as a brown qum which could not be crystallised, λ_{max} 1680, 1650, 1600, 1510 cm⁻¹; ¹H nmr & 7.05 (2H, d, J = 8 Hz, 2'-H, 6'-H), 6.81 (2H, d, J = 8 Hz, 3'-H, 5'-H), 6.55 (1H, s, 5-H), 6.2 (1H, s, 8-H), 5.86 (2H, dd, J = 8, 1.5 Hz), O-CH₂-O), 5.1 (1H, m, 1-H), 3.78 (3H, s, OCH₂), 2.5 (3H, s, NCH₂); ms m/z 311 (M⁺, 0.2%), 310 (0.2), 296 (0.2), 191 (12), 190 (100), 188 (7), 174 (13), 160 (7), 144 (12), 132 (8),

121 (80), 77 (15), 59 (21), 43 (36). Found: 311.1515; calculated for $C_{10}H_{21}NO_3$: 311.1521. The compound was identified as O-methylcinnamolaurine by comparison with an authentic sample prepared by diazomethane methylation of cinnamolaurine¹⁵.

Fraction 6: Isouvariopsine and Glaucine - This fraction (0.045 g, aliguots 44-47) yielded two

compounds on ptlc. The less polar compound, isouvariopsine, was obtained as a yellow solid, mp 155-157°, λ_{max} 218 (3.93), 250 (4.35), 260 (4.35), 313 (3.88), 325 (3.91), 360 (3.62), 378 mm (3.65); v_{max} 1610, 1590, 1500, 1450 σm^{-1} ; m m δ 8.6 (1H, m), 7.8-7.15 (5H, m), 6.25 (2H, s, *WC3-0).* 4.0 (3H, s, CCH31, 3.27 12H, m) , 2.65 (2H, m) , 2.43 (6H, N-M3 **x** 2) ; **ms** m/z 323 (M', 60). 308 (12). 292 (161, 278 (30). 265 (42), 247 (81, 222 (la), 205 (7). 176 (27). 163 (36). 58 (100). Found: 323.1356; calculated for $C_{20}H_{21}NO_3$: 323.1539. The more polar compound was identified as glaucine⁴ by comparison with an authentic sample, colourless crystals from methanol, mp 120°C(lit.⁴ 120-121°C); λ_{max} 218 (4.58), 281 (4.18), 303 nm (4.16); v_{max} 1590, 1575, 1510, 1460 cm^{-1} ; ¹H mm s 7.98 (1H, s), 6.81 (1H, s), 6.68 (1H, s), 3.97 (6H, s, 0CH3 **^x**2). 3.92 (3H, s, OX3), 3.72 (3H. **s,** CCH3), 2.59 (3H. s, N-M3) ; **ms** m/z 355 (M', 80). 354 (100), 340 (26), 338 (23), 324 (14), 297 (10), 281 (8), 177.5 (M^{++}) . Fraction $7 - (0.141 \text{ g})$, aliquots 48-96) consisted mainly of glaucine.

Fractions 8 and ⁹- (aliquots 97-127) gave negative Mayerrs tests, **and** on keeping depsited white crystals that pmed identical with **those** obtained previously frm the acid solution of crude alkaloids by ether extraction; needles from methanol, mp 253-254°C.

Volatile non-alkaloid fraction -- The methanol recovered from the extraction deposited white crystals (0.7 g, 3.15 x 10⁻⁵%); recrystallised from methanol, white flakes mp 62°C, v_{max} (KBr) 2900 (CH), 1430 cm⁻¹; m/z, 204 (M⁺, 10), 189 (9), 164 (7), 161 (12), 149 (22), 135 (5), 122 (8), 109 (14), 108 (12), 81 (12), 79 (8). The material was identified as a mixture of α - and β eudesmol by comparison of its glc and spectroscopic data with those of authentic samples. Attempted N-Methylation of $6,6a$ -Dehydronorlaureline - $6,6a$ -Dehydronorlaureline (0.10 g) in methanol (3 ml) was stirred with formaldehyde (37%, 0.2 ml) at room temperature for 5 h, then **scdium** bomhydride (0.05 g) was added in mall portions. The solvents were rawved, the residue dissolved in dilute **aqueous** hydrochloric acid (10 ml), and the solution was basifid with amnonia and extracted with chloroform. The product obtained gave no evidence of an N-CH₃ with ammonia and extracted with chloroform. The product obtained gave no evidence of an M
group (¹H mmr and ms) but the molecular weight was found to have increased by 2 amu. The $1\mathbf{I}_{\mathbf{H}}$ compound was later identified as norlaureline.

Conversion of 6,6₃-Dehydronorlaureline to Laureline - 6,6₃-Dehydronorlaureline (0.020 g)

dissolved in acetone (5 ml) was **treated** with methyl icdide (0.05 ml) in a sealed tube and left overnight at room temperature. Removal of the solvent gave a brown solid, ms m/z 308, which was dissolved in methanol (5 ml) and treated with scdim torohydride (0.020 **q).** The usual work-up of the reaction mixture gave laureline (0.018 g) as a brown η **m**, λ_{max} 218 (4.20) , 264 (3.95) , 273 (4.1), 315 (3.57), 325 nm (3.25); 1 H nmx δ 7.66 (1H, d, J = 2.5 Hz), 7.15 (1H, d, J = 10 Hz), 6.75 (lH, d, J=lOHz), 6.53 (lH, **s),** 6.03 IlH, d, J= 1.5Hz), 5.88 (lH, d, J=1.5Hz); msmlz 309

 $(M⁺$, 65), 308 (100), 294 (53), 266 (48), which was identified by comparison with an authentic. sample of laureline.

ACKNOWLEDGEMENTS

We are grateful to Mr Paul Barnett for the identification and supply of plant material from King Island, and to Dr E. Gellert, University of Wollongong, for an authentic sample of cinnamolaurine. We thank also the University of Tasmania for Postgraduate Research Scholarships to YACPG and H-ML, and the Australian Research Grants Scheme for financial assistance for this work.

REFERENCES

- 1. A.W. Sangster and K.L. Stuart, Chem. Rev., 1965, 65, 69.
- I.R.C. Bick, J. Harley-Mason, N. Sheppard, and M.J. Vernengo, J. Chem. Soc., 1961, 1896. $2 -$
- $3.$ W.H. Barschers, R.R. Arndt, K. Pachler, J.A. Weisbach, and B. Douglas, J. Chem. Soc., 1964, 4778.
- H. Guinaudeau, M. Leboeuf, and A. Cave, J. Nat. Products, 1975, 38, 275; 1975, 42, 325. 4.
- M. Shamma, S.Y. Yao, B.R. Pai, and R. Charubala, J. Org. Chem., 1971, 36, 3253. 5.
- M. Shamma and W.A. Slusarchyk, Chem. Rev., 1964, 64, 59. 6.
- 7. H.B. Dutschewska, A.S. Orahovats, and N.M. Mollov, Compt. rend. Acad. bulg. Sci., 1973, 26, 899; Chem. Abstr., 1974, 80, 27410.
- H. Guinaudeau, J. Nat. Prod., 1983, 46, 761. 8.
- M. Shamma, "The Isoquinoline Alkaloids", Academic Press, New York, 1972, pp. 245-258. 9.
- 10. V.B. Chervenkova, N.M. Mollov, and S. Paszyc, Phytochem., 1981, 20, 2285.
- 11. V.A. Mnatsakanyan, V. Preinger, V. Šimánek, A. Klásek, L. Dolejš, and F. Šantavý, Tetrahedron Lett., 1974, 851.
- 12. H.D. Gibbs, J. Biol. Chem., 1927, 72, 649.
- 13. I.R.C. Bick, Y.A.G.P. Gunawardana, V.A. Patrick, and A.H. White, Austral. J. Chem., 1985, 38, 1571.
- 14. M. Shamma, "The Isoquinoline Alkaloids", Academic Press, New York, 1972, pp. 259-264.
- 15. E. Gellert and R.E. Summons, Austral. J. Chem., 1970, 23, 2095.
- 16. F.J. McQuillin and J.D. Parrak, J. Chem. Soc., 1956, 2573.
- 17. R.D. Hellyer, Austral. J. Chem., 1962, 15. 157.
- 18. R.V.H. Jones and M.D. Sutherland, Chem. Commun., 1968, 1229.
- 19. K. Hostettman, M. Hostettmann-Kaldas, and K. Nakanishi, J. Chromatography, 1979, 170, 355.

Received, 30th September, 1986