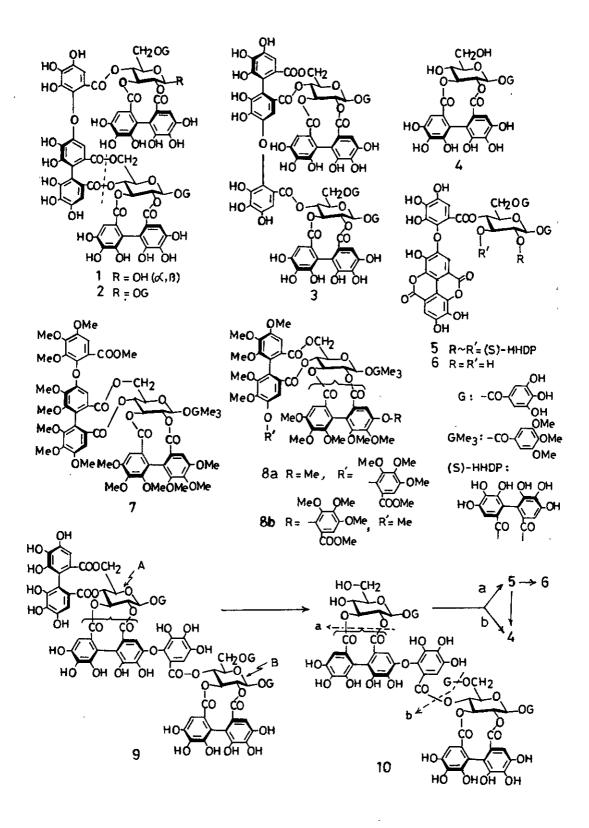
REVISED STRUCTURE OF NOBOTANIN B, A DIMERIC ELLAGITANNIN OF <u>TIBOUCHINA SEMIDECANDRA</u>

Takashi Yoshida,<sup>a</sup> Kumiko Haba,<sup>a</sup> Tetsuro Shingu,<sup>b</sup> and Takuo Okuda<sup>\*,a</sup> Faculty of Pharmaceutical Sciences, Okayama University,<sup>a</sup> Tsushima, Okayama 700, Japan and Faculty of Pharmaceutical Sciences, Kobe Gakuin University,<sup>b</sup> Ikawadani, Nishi-ku, Kobe 673, Japan

<u>Abstract</u> — The structure of nobotanin B, a dimeric ellagitannin of <u>Tibouchina semidecandra</u>, has been revised to 9 on the basis of structural analysis of a partial hydrolysate.

Nobotanin B is a dimeric ellagitannin isolated from Tibouchina semidecandra (Melastomataceae) along with nobotanin A (1) and nobotanin F (2). The structure 3 was proposed for nobotanin B in our previous communication,<sup>1</sup> based on the following evidence: (i) Nobotanin B is a structural isomer of nobotanin F (2), as it gave three partial hydrolysates, 4-6, upon the hydrolysis similar to that of nobotanin F.<sup>1</sup> Each glucose core in nobotanin B was therefore presumed to have a hexahydroxydiphenoyl (HHDP) group at  $0-2 \sim 0-3$  as in nobotanin F. (ii) Methylation of nobotanin B with dimethyl sulfate and potassium carbonate in acetone gave a partially degraded product, having eighteen methoxyl groups, and it was presumed to have the structure 8a which is isomeric to the structure of 7 derived from 2. However, upon detailed examination by HPLC during the hydrolyses of nobotanins B and F, we have found that in case of the former, 4 and 5 have been produced most probably from a dimeric partial hydrolysate of a retention time a liitle shorter than that of the starting material, although they can be directly produced from nobotanin F by the cleavage illustrated by a dotted line in the structure (2). The observation of this difference prompted us to reinvestigate the structure of nobotanin B.

A dimeric partial hydrolysate (10),  $C_{68}H_{50}O_{44}.7H_2O$ ,  $[\alpha]_D +31^{\circ}(MeOH)$ , from nobotanin B, was isolated as the main product after the treatment with boiling



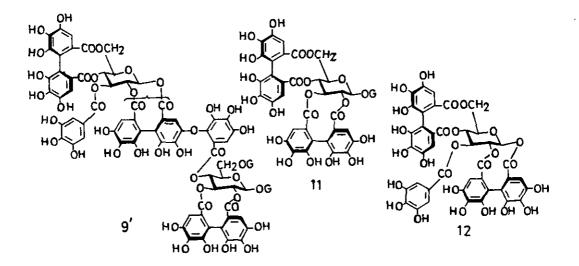


Table I. <sup>1</sup>H-Nmr Data of the Glucose Moieties of Nobotanin B (9), Partial Hydrolysate (10), Casuarictin (11) and Roxbin B (12) (400 MHz, acetone-dc)

	acetone-d <sub>6</sub> )						
	H-1	H-2	H-3	H-4	H-5	H-6	Н-б′
9 Gluc-A	6.20 d	5.10 dd	5.82 dd	5.17 t	4.67 dd	5.33 dd	3.92 d
	(J=8.5)	(J=8.5,9)	(J=9,10)	(J=10)	(J=6,10)	(J=6,13.5)	(J=13.5)
Gluc-B	6.02 d	5.18 dd	5.41 t	5.83 t	3.45 dd	4.92 d	3.91 dd
	(J=8.5)	(J=8.5,10)	(J=10)	(J=10)	(J=2,10)	(J=13)	(J=2,13)
10 Gluc-A	6.30 d	4.91 dd	5.50 dd	3.90 t	4.10 dd	3.81 dd	3,94 d
	(J=8.5)	(J=8,5,9)	(J=9,10)	(J=10)	(J≖6,10)	(J=6,13)	(J=13)
Gluc-B	5.96 d	5.14 dd	5.33 t	5.80 t	3.49 br.d	4.84 dd	3.97 d
	(J=8.5)	(J=8.5,10)	(J=10)	(J=10)	(J=10)	(J=2,13)	(J=13)
11	6.22 d	5.18 t	5.45 dd	5.17 t	4.50 dd	5.37 dd	3.88 d
	(J=9)	(J≈9)	(J=9,10)	(J=10)	(J=7,10)	(J=7,13)	(J=13)
12	6,08 d	4.90 dd	5,21 t	4.89 dd	4.40 br.dd	1 5.30 dd	3.80 dd
	(J=8.5)(	J=8.5,9.7)	(J=9.7)(J	=9.7,10)	(J=6.5,10)	(J=6.5,13)	(J=1.5, 13)
	Gluc-B Gluc-A	Gluc-A 6.20 d (J=8.5) Gluc-B 6.02 d (J=8.5) Gluc-A 6.30 d (J=8.5) Gluc-B 5.96 d (J=8.5) 6.22 d (J=9) 6.08 d	Gluc-A 6.20 d 5.10 dd (J=8.5) (J=8.5,9) Gluc-B 6.02 d 5.18 dd (J=8.5) (J=8.5,10) Gluc-A 6.30 d 4.91 dd (J=8.5) (J=8.5,9) Gluc-B 5.96 d 5.14 dd (J=8.5) (J=8.5,10) 6.22 d 5.18 t (J=9) (J=9) 6.08 d 4.90 dd	Gluc-A 6.20 d 5.10 dd 5.82 dd (J=8.5) (J=8.5,9) (J=9,10) Gluc-B 6.02 d 5.18 dd 5.41 t (J=8.5) (J=8.5,10) (J=10) Gluc-A 6.30 d 4.91 dd 5.50 dd (J=8.5) (J=8.5,9) (J=9,10) Gluc-B 5.96 d 5.14 dd 5.33 t (J=8.5) (J=8.5,10) (J=10) 6.22 d 5.18 t 5.45 dd (J=9) (J≈9) (J=9,10) 6.08 d 4.90 dd 5.21 t	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$

Table II. 13C-Nmr Data of the Glucose Moieties of Nobotanin B (9), Casuarictin (11) and Roxbin B (12) (100 MHz, acetone-d<sub>6</sub>)

	(11) and noxoin 5 (12) (100 mml, account -6)						
	C-1	C-2	C-3	C-4	C-5	C-6	
9 <sup>a)</sup> Gluc-A	92.34	76.77	76,98	69.61	73.37	63,37	
Gluc-B	92.34	75.38	78.07	66.89	73.92	63.57	
11 <sup>b)</sup>	92.4	76.0	77.3	69,3	73.5	63.1	
12	92.51	77.24	78.22	69.62	73.24	63.03	

a) Assigned by  ${}^{1}H^{-13}C$  shift correlation spectrum.

b) Data from Reference 2.

water for 7 h. The  $^{1}$ H-nmr spectrum (400 MHz, acetone-d<sub>4</sub>) of 10 indicates the presence of three galloyl groups (  $\delta$  7.27, 7.09 and 6.95), a valoneoyl and an HHDP group ( $\delta$  7.12, 6.72, 6.47, 6.45 and 6.08), suggesting that 10 lacks one of the HHDP groups in nobotanin B. Extensive spin-spin decoupling experiments for the glucose proton signals of 10 revealed significant upfield shifts of H-4 ( $\delta$  5.17  $\rightarrow$  3.90) and a part of C-6 methylene proton signals ( $\delta$  5.33  $\rightarrow$  3.81) of the glucose core A, from those of nobotanin B (Table I), to indicate that the elimination of HHDP group occurred at  $0-4 \sim 0-6$ . This evidence, and production of 4 and 5 upon prolonged hydrolysis of 10, led to a conclusion that a galloyl and a valoneoyl group are located at 0-1, 0-2 and 0-3 in the glucose core A of 10, and of nobotanin B. Between the assignable structures 9 and 9' of nobotanin B, the former is supported by comparison of its  $^{1}H$ - and  $^{13}C$ -nmr data with those of casuarictin  $(11)^{2,3}$  and roxbin B  $(12)^4$ ; As in Tables I and II, the proton and  $^{13}$ C signals, particularly C-1 and C-3 signals, of glucose core A in nobotanin B, coincide with those of 11. Further evidence for the presence of a galloyl group at 0-1 of each glucose core in nobotanin B was obtained by a reversed-phase HPLC analysis<sup>5</sup> of the reaction mixture of degalloylation with tannase: A peak of gallic acid and four peaks due to anomers were shown. The latter four peaks were replaced by a single peak of different retention time after treatment of the reaction mixture with NaBH<sub>l</sub>.<sup>6</sup> The structures of nobotanin B and the</sub>octadecamethyl derivative obtained on the methylation are therefore revised to 9 (orientation of the valoneoyl group at 0-2 and 0-3 may be reversed) and 8b, respectively.

## **REFERENCES AND NOTES**

- T. Yoshida, Y. Ikeda, H. Ohbayashi, K. Ishihara, Y. Ohwashi, T. Shingu, and T. Okuda, <u>Chem. Pharm. Bull.</u>, 34, 2676 (1986).
- 2) T. Yoshida, T. Hatano, T. Okuda, M. U. Memon, T. Shingu, and K. Inoue, <u>Chem.</u> <u>Pharm. Bull.</u>, 32, 1790 (1984).
- T. Okuda, T. Yoshida, M. Ashida, and K. Yazaki, <u>J. Chem. Soc., Perkin Trans</u>. 1, 1983, 1765.
- T. Yoshida, T. Hatano, Xin-Min Chen, M. Fukushima, and T. Okuda, <u>Chem. Pharm.</u> <u>Bull.</u>, 35, 1817 (1987).
- 5) HPLC: column, YMC A312 (ODS)(150 mm x 6 mm); eluent, 0.01M phosphate buffer-EtOH (100:5); detection, A<sub>280</sub>.
- 6) T. Yoshida, Y. Maruyama, M. U. Memon, T. Shingu, and T. Okuda, <u>Phytochemist-</u> <u>ry</u>, 24, 1041 (1985).

Received, 30th June, 1987