SYNTHESIS OF ANOMALOUSLY COUPLED NUCLEOSIDES BY ADDITION OF PURINES TO UNSATURATED SUGAR ALDEHYDES

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<u>Abstract</u> - α , β -Unsaturated sugar aldehydes were reacted in a Michael type of reaction with purines in the presence of an organic base to give isonucleosides.

Recently, we have shown that unprotected 2-deoxy-D-ribose is anomalously coupled with nucleobases when reacted with a tributylammonium polyphosphate mixture in chloroform¹. The reaction of the 2-deoxysugar at C-3 with purines was assumed to take place *via* a dehydration/addition process as follows:



To prove this assumption the attention was turned to the addition of bases 1 and 2 to unsaturated sugar aldehydes 3 and 4 as well as hemiacetal 5.



Compound 3 reacted with 1 in the presence of triethylamine (TEA) in DMF at 40° C for 7 days to produce the three adduct 6 in 20% yield. The hemiacetal 5 reacted similarly, but not in a clean reaction according to silica TLC. The aldehyde 4 easily available from tri-O-acetyl-D-glucal², was treated with 1 and TEA in DMF at 40° C for 8 days and the α -D-glucopyranose anomer 7 could be isolated in 21% yield by flash chromatography on silica with methanol/dichloromethane (1:25). In an addition reaction of theophylline (2) with the aldehyde 4 in DMF a large increase was observed in reaction rate when 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) was used instead of TEA. After 3 h at room temperature an adduct was isolated by adding water to the reaction mixture, neutralizing to pH 7, extraction with dichloromethane, and silica chromatography with methanol/ether (1:20). The primary adduct was deacetylated by treatment with 2% HCl in methanol at 60° C for 15 h to give a mixture of the methyl glycosides 8 in 31% overall yield form 4.



Pu = 6-(4-Chlorophenylamino)-2-methyl-9-purinyl
Th = 7-Theophyllyl

The structures and their corresponding conformations were assigned on the basis of their ¹H, ¹³C-NMR and mass spectra. 500 MHz ¹H-NMR spectra were recorded for **6** and **8**³. The assignment of the shift values in the ¹H-NMR spectra was based on the use of ¹H-¹H homonuclear shift correlated 2-D NMR. With this technique it was possible to assign the axial 4a-H in the β -anomer of **6** as a quartet with J = 12 Hz. This can be explained by assuming couplings to one geminal proton and two vicinal axial protons which prove three

configuration of 6 in the Cl conformation. This is confirmed by large couplings of the axial 3a-H to vicinal and axial orientated 2a-H and 4a-H for both α and β -anomers. In the same manner the coupling constants of 3-H in 7 and 8 prove the arabino configuration in the Cl conformation with the bulky purin-yl group at C-3 always in an equatorial position.

Characterization of 6, 7 and 8 as follows:

6: mp 200-201°C; m/z (EI): 431 (M⁺, 11); NMR (DMSO-d₆) from the mixture of α and β -anomers (3:2).

 $\beta-\text{anomer:} {}^{4}\text{H-NMR} (500 \text{ MHz}) \delta = 1.89 (q, J_{4a,4e} = J_{3,4a} = J_{4a,5} = 12\text{Hz}, 4a-\text{H}), 1.92-2.04 (m, 4e-\text{H}, 2a-\text{H}), 2.03 (s, OAc, 2.14 (m, J_{2e,2a} = 12\text{Hz}, 2e), 2.50 (s, CH_{3}), 3.89 (m, \Sigma [J] = 24\text{Hz}, 5-\text{H}), 4.03-4.10 (m, 6, 6'-\text{H}), 4.79 (tt, J_{2a,3} = J_{3,4a} = 12\text{Hz}, J_{2e,3} = J_{3,4e} = 4\text{Hz}, 3\text{H}) 4.87 (ddd, J_{1,OH} = 6\text{Hz}, J_{1,2e} = 2\text{Hz}, J_{1,2a} = 10\text{Hz}, 1-\text{H}), 6.91 (d, J = 6\text{Hz}, OH), 7.35 (d, J = 9\text{Hz}, Ar\text{H}), 8.01 (d, J = 9\text{Hz}, Ar\text{H}), 8.35 (s, 8'-\text{H}), 9.88 (s, \text{NH}). {}^{13}\text{C-} \text{NMR} (63 \text{ MHz}) \delta = 20.62 (Ac), 25.87 (CH_{3}), 32.87 (C-4), 38.36 (C-2), 49.51 (C-3), 65.86 (C-6), 70.01 (C-5), 94.01 (C-1), 118.06 (C-5'), 181.73 (C-2"), 125.67 (C-4"), 128.10 (C-3"), 138.97 (C-1"), 139.66 (C-8'), 150.05 (C-4'), 151.29 (C-6'), 160.37 (C-2"), 170.21 (C=0).$

7: mp 200-201°C, m/z (EI): 489 (M⁺, 1.4); ¹H-NMR (DMSO-d₆, 250 MHz) δ = 1.67 (s, 4-OAc), 2.03 (s, 6-OAc), 2.08 (m, 2e-H), 2.57 (s, CH₃), 2.82 (t, J_{2a,3} = J_{2a,2e} = 13.1 Hz, 2a-H), 4.04 (dd, J_{5,6} = 1.6, J_{6,6} = 12 Hz, 6-H), 4.21 (dd, J_{5,6} = 5.1 Hz, 6'-H), 4.30 (m, J_{4,5} = 9.9 Hz, 5-H), 5.05 (m, J_{3,4} = 9.9 Hz

 $J_{2a,3} = 13.1 \text{ Hz}, J_{2e,3} = 4.4 \text{ Hz}, 3-\text{H}, 5.25 (t, 4-\text{H}), 5.42 (m, 1-\text{H}), 6.99$ $(d, J_{1,OH} = 3.7 \text{ Hz}, OH) 7.37 (d, J_{2",3"} = 8.9 \text{ Hz}, 3"-\text{H}), 8.05 (d 2"-\text{H}), 8.45$ $(s, 8'-\text{H}), 9.89 (s, NH). ¹³C-NMR (DMSO-d_6, 63 MHz) <math>\delta = 19.88, 20.55$ (2Ac), 25.86 (CH₃), 35.20 (C-2), 50.71 (C-3), 62.67 (C-6), 67.36, 69.68 (C-4,C-5), 89.81 (C-1), 117.8 (C-5'), 121.67 (C-2"), 125.67 (C-4"), 128.12 (C-3"), 138.94 (C-1"), 140.38 (C-8'), 150.70 (C-4'), 151.17 (C-6'), 160.44 (C-2'), 168.80, 170.05 (2 C=0).

8: mp 269-271°C; m/z (EI): 340 (M⁺, 21); NMR (DMSO-d₆) from the mixture of α and β -anomers (2:1).

 $\beta \text{-anomer: } ^{1}\text{H-NMR} (500 \text{ MHz}) \delta = 2.12 (ddd, J_{1,2e} = 2 \text{ Hz}, J_{2a,2e} = 11\text{Hz}, J_{2e,3} = 4 \text{ Hz}, 2e-\text{H}), 2.26 (q, J_{1,2a} = J_{2a,3} = 10 \text{ Hz}, 2a-\text{H}), 3.25 (s, CH_3), 3.27 (m, 5-\text{H}), 3.31 (CH_3), 3.42 (s, CH_3), 3.54 (m, 6'-\text{H}), 3.74 (m, 6-\text{H}), 3.88 (dt, J_{3,4} = J_{4,5} = 10 \text{ Hz}, J_{4,0H} = 7\text{Hz}, 4-\text{H}), 4.55 (dd, 1-\text{H}), 4.59 (hidden by 6-0H, 3-\text{H}), 4.63 (t, J_{6,0H} = J_{6',0H} = 7 \text{ Hz}, 6-0\text{H}), 5.18 (d, 4-0\text{H}). ^{13}\text{C-NMR} (63 \text{ MHz}) \delta = 27.61, 29.27 (2 CH_3), 37.03 (C-2), 55.58 (CH_3), 59.47 (C-3), 60.98 (C-6), 67.25 (C-4), 78.23 (C-5), 100.32 (C-1), 106.04 (C-5'), 142.11 (C-8'), 148.78 (C-4'), 150.66 (C-2'), 154.04 (C-6').$

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