

LANGUIDULINE, A DITERPENOID WITH AN UNUSUAL STRUCTURE FROM SALVIA LANGUIDULA

Jorge Cárdenas,* Baldomero Esquivel, Rubén A. Toscano, and Lydia Rodríguez-Hahn
 Instituto de Química, Universidad Nacional Autónoma de México,
 Circuito Exterior, Ciudad Universitaria, Coyoacán 04510, México, D. F.

Abstract - The unusual structure of languiduline, a new diterpenoid isolated from Salvia languidula Epl (Labiatae), was deduced from the spectral data and confirmed by X-ray diffraction analysis.

Salvia genus has been shown to be a rich source of diterpenoids, mainly of the ent-clerodane¹ and abietane type.² It is represented in Mexico by over 275 species³ mostly of the Salvia, subgenus Calosphaea.⁴

As a result of our chemosystematic studies on Mexican Salvias, several ent-clerodane diterpenoids have been isolated.¹ Some of them presented a rearranged ent-clerodane skeleton, for example salvigenolide from Salvia fulgens⁵ showed an 8(9→11) abeo ent-clerodane skeleton. Recently we have isolated from S. puberula⁶ puberulin and iso-puberulin, two diterpenoids with novel benzonorcaradiene and benzocycloheptatriene skeletons of clerodanic origin.

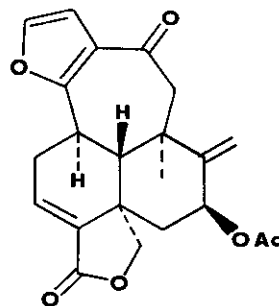
In continuation of our systematic study of Mexican Salvias we have analysed Salvia languidula Epl (Salvia, Section Angulatae, Subsection Glumaceae). From the acetonic extract of the aerial parts of the plant, a diterpenoid languiduline 1, was isolated after extensive chromatographic purification. Languiduline 1, showed mp 243-245°C, $[\alpha]_D^{20} = -193.43$ (c 0.2, CHCl₃). Its mass spectrum was consistent with a molecular formula C₂₂H₂₂O₆ (M⁺ 382), the base peak at m/z 339 (M⁺-43) and a peak at 322 (M⁺-60) indicated that it contained an acetate group. The ir spectrum of 1 showed the presence of an α,β -unsaturated γ -lactone function (1774 cm⁻¹) and acetate carbonyl at 1734 cm⁻¹. A strong band at 1654 cm⁻¹ was attributed to an α,β -unsaturated ketone, the absorptions at 1565, 1528 and 830 cm⁻¹ were assigned to a disubstituted furan ring. The uv spectrum of 1 [λ_{max} 208, 220 and 265 nm ($\epsilon = 28000, 17000$ and 5500)] indicated that the ketonic group is conjugated with the furan ring. These data suggested that the furan ring is fused to a tricyclic hydrocarbon skeleton which contains an α,β -unsaturated γ -lactone and an additional double bond.

The 300 MHz nmr spectrum of languiduline 1 and adequate double resonance experiments allowed for a complete assignment of all the protons in the molecule (Table 1). Thus it showed signals of an α,β -substituted furan ring and an α,β -unsaturated 18:19 olide. The C-19 methylene was observed as an

ABX system at δ 4.14 (H-19, pro-R, d, $J=8.1$ Hz) and 4.02 (H-19, pro-S, dd, $J=8.1$ and 2.4 Hz). The long-range coupling exhibited by H-19 pro-S suggested an α -axial orientation for C-19 and the absence of a β -substituent at C6.⁷ The multiplicity of H-3 showed that it is bound to a methylene group. A singlet at δ 2.05 was ascribed to the acetate group; its geminal proton appeared as a doublet ($J=8$ Hz) at δ 5.72 and was shown to be coupled to H-6 (Table 1), therefore the acetate group must be bound to C7 and β -pseudo-axial. The ¹H nmr spectrum of 1 did not show the secondary methyl group doublet usually present in the protonic resonance spectra of clerodanic diterpenoids,^{1,7} instead two vinylic singlets were observed at δ 5.43 and 5.36 and were assigned to the exocyclic C17 methylene double bond. This double bond must be spacially oriented in such a way as to exert a protective effect on the 19-pro-R proton (δ 4.14).⁷

Table 1. 300 MHz NMR SPECTRUM OF 1

H	δ	J in Hz
1	2.97 td	10, 3.1
2 α	3.31 ddd	17, 7.9, 3.1
2 β	2.41 ddd	17, 10, 2.4
3	6.88 dd	7.9, 2.4
6 α	2.65 dd	15, 8
6 β	1.68 dd	15, 2.4
7	5.72 d	8
10	2.94 d	10
11 α	2.66 d	16.8
11 β	3.35 dd	16.8, 0.74
14	6.77 d	2.1
15	7.35 d	2.1
17	5.43 s	- - -
17'	5.36 s	- - -
19 pro-R	4.14 d	8.1
19 pro-S	4.02 dd	8.1, 2.4
20	1.23 d	0.74
Ac	2.05 s	- - -



1

H-1 and H-10 were localised at δ 2.97 (td, $J=10$ and 3.1 Hz) and 2.94 (d, $J=10$ Hz), respectively, which indicated a trans-axial relationship between them and the allylic nature of H-1, therefore C1 must be bound to C16 in languiduline 1. An ABX₃ system observed at δ 2.66 (d, $J=16.8$ Hz) and 3.35 (dd, $J=16.8$ and 0.74 Hz) was assigned to the C11 methylene which must be bound to the ketonic group and to a fully substituted carbon atom. The 11 β proton showed a small long-range coupling with the C20 methyl which was observed at δ 1.23 (d, $J=0.74$ Hz).

The ¹³C nmr spectrum of languiduline (Table 2) was consistent with the structure 1 proposed for it. A singlet at δ 192.96 was ascribed to the ketonic carbon atom. This carbon resonance is adequate for an α,β -unsaturated cycloheptenone.⁸ It is noteworthy of mention the presence of only three

non oxygen bearing sp^3 triplets assigned to C2, C6 and C11. The chemical shifts and multiplicity of C1 (δ 36.22, d) confirmed that it is bound to the α furan carbon 16.

Table 2. ^{13}C NMR SPECTRUM OF 1

C	δ	C	δ
1	36.22 d	12	192.96 s
2	32.37 t	13	123.28 s
3	132.69 d	14	110.14 d
4	136.20 s	15	142.06 d
5	45.11 s	16	158.72 s
6	40.97 t	17	118.18 t
7	73.98 d	18	168.04 s
8	151.14 s	19	72.17 t
9	39.33 s	20	21.40 q*
10	48.57 d	OCOCH ₃	169.16 s
11	61.89 t	OCOCH ₃	20.04 q*

Recorded at 20 MHz. TMS as internal standard SFORD multiplicity.

* These values may be interchanged.

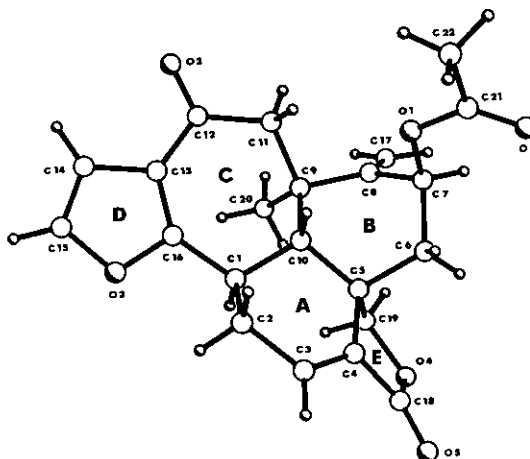


Fig. 2

The unusual structure of languiduline was confirmed by an X-ray analysis. Crystals of languiduline were obtained by slow crystallization from ethyl acetate. The crystal was orthorhombic in space group $P2_12_12_1$, with one molecule of composition $C_{22}H_{22}O_6$ per asymmetric unit ($\rho_{\text{calc}} = 1.29 \text{ g cm}^{-3}$), $Z=4$, $a=11.229(4)$, $b=11.394(5)$ and $c=15.388(6)$ Å; $V=1968.9(4)$ Å³. Intensities of 1496 independent reflections were measured on a Nicolet R3m diffractometer using MoK α radiation ($\lambda=0.7107$ Å) up to $2\theta \leq 45^\circ$ of which 1247 were considered to be observed [$I > 2.5\sigma(I)$]. The structure was solved by direct methods and refined anisotropically for the non-hydrogen atoms by least-squares. Hydrogen atoms were forced to ride on the corresponding carbon atoms with a fixed isotropic temperature factor $U = 0.06$ Å². Final $R=0.047$ ($R_w=0.044$) and no peaks greater than $\pm 0.19e$ Å⁻³. All calculations were performed using the program package SHELXTL⁹ on a Nova 4S computer. A computer-generated drawing is given in Fig. 2 and reveals the relative stereochemistry of languiduline. The ring A has a *sofa*^{10a} conformation: A/B and A/C rings are *trans*-fused with torsion^{10b} angles of $-33.2(5)$; $70.7(4)^\circ$ and $60.9(5)$; $-45.1(5)^\circ$, respectively. The junction A/E has torsion angles of $1.1(7)$; $36.8(4)^\circ$. The ring B has a twisted-boat conformation with the acetate group at C7 and the methyl group at C9 on axial and equatorial position, holding anticlinal [$\tau=121.5(5)^\circ$] and synperiplanar [$\tau=23.5(6)^\circ$] relation with the exocyclic C=C at C8. The B/C junction is *trans* [$-35.2(5)$; $80.3(5)^\circ$]. The C ring has a *sofa* conformation. The carbonyl group C12-O2 is conjugated with the furan ring D. The γ -lactone ring E adopts an envelope conformation with flap at C5. The molecular packing is governed by van der Waals interactions.¹¹

Languiduline 1 shows an unusual skeleton with a seven-membered ring as a result of the linkage of C1 to C16 in a clerodanic precursor. To our knowledge there are no precedents of this type of annulation in clerodanes of natural origin.

ACKNOWLEDGMENTS

We are grateful to Dr. T. P. Ramamoorthy, Botany Department, Instituto de Biología, UNAM, for botanical classification of the plant material. We thank warmly Dr. Benjamin Rodríguez, Instituto de Química Orgánica CSIC, Madrid, Spain, for the 300 MHz ¹H nmr spectrum of 1. The authors are indebted to Messrs. R. Villena, M. Torres, L. Velasco, A. Cuellar and R. Gaviño for technical assistance. This work was supported in part by the CONACYT (Project PCCBBNA 021142).

REFERENCES

1. A. A. Sánchez, B. Esquivel, A. Pera, J. Cárdenas, M. Soriano-García, A. Toscano, and L. Rodríguez-Hahn, Phytochemistry 1987, 26, 479 and references cited therein.
2. M. A. Galicia, B. Esquivel, A. A. Sánchez, J. Cárdenas, T. P. Ramamoorthy, and L. Rodríguez-Hahn, Phytochemistry, in press.
3. T. P. Ramamoorthy, personal communication.
4. C. Epling, Report. Spec. Nov. Beih., 1939, 110, 1.
5. B. Esquivel, J. Cárdenas, A. Toscano, M. Soriano-García, and L. Rodríguez-Hahn, Tetrahedron, 1985, 41, 3213.
6. L. Rodríguez-Hahn, B. Esquivel, A. A. Sánchez, J. Cárdenas, O. G. Tovar, M. Soriano-García, and A. Toscano, J. Org. Chem., in press.
7. B. Esquivel, A. Vallejo, R. Gaviño, J. Cárdenas, A. A. Sánchez, T. P. Ramamoorthy, and L. Rodríguez-Hahn, Phytochemistry, in press.
8. A. Ahond, A. M. Bui, P. Potier, E. W. Hagaman, and E. Wenkert, J. Org. Chem., 1976, 41, 1878.
9. G. M. Sheldrick. SHELXTL Revision 4. An integrated system for solving, refining and displaying structures from diffraction data. University of Göttingen (1983).
10. a) R. Bucourt. Topics in Stereochemistry. Vol. 8, p. 186, Ed. E. L. Eliel and N. L. Allinger, John Wiley and Sons, New York (1974);
b) ibid., p. 192.

Received, 1st March, 1988