

STRUCTURE OF 6- OR 8-ISOPRENOID SUBSTITUTED FLAVANONE:
 CHEMICAL SHIFT OF THE HYDROGEN-BONDED HYDROXYL GROUP¹

Toshio Fukai and Taro Nomura*

Faculty of Pharmaceutical Sciences, Toho University,
 2-2-1, Miyama, Funabashi, Chiba 274, Japan

Abstract — The proton signal of the hydrogen-bonded hydroxyl group of the 6-isoprenoid substituted flavanone was shifted more downfield than that of 6-nonsubstituted flavanone. By comparison of the chemical shifts of the hydrogen-bonded hydroxyl groups of 8-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone (3) and its 6-isomer (4) isolated from *Wyethia helenioides*, the proposed structures 3 and 4 were reversed each other. Comparison of the chemical shift of the hydroxyl group is a facile method to discriminate the 6-isoprenoid substituted flavanone and the 6-nonsubstituted one.

In the previous paper,² we reported that the proton signal of the hydrogen-bonded hydroxyl group at the C-5 position of the 6-isoprenoid substituted isoflavone was observed in the range of δ 13.0-13.35 in acetone-d₆, while the relevant proton signal of the 8-substituted isomer in the range of δ 12.7-13.0. On the basis of the above observation and other spectral evidence, we proposed the structures of the 6- or 8-prenylated isoflavones.² In this paper, we report the observation on the chemical shift of the hydrogen-bonded hydroxyl group at the C-5 position of the 6- and/or 8-isoprenoid substituted flavanones, and also describe the revision of the structures of 8-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone and its 6-isomer isolated from *Wyethia helenioides* (tribe Heliantheae, subtribe Helianthinae).³

We examined the ¹H nmr spectra of 48 kinds of the known 6- and/or 8-isoprenoid substituted flavanones. Except for the seven kinds of flavanones, the proton signal of the hydrogen-bonded hydroxyl group of the 6- and/or 8-isoprenoid substituted flavanone could be classified into the following two types according to the location

of the isoprenoid moiety: 1) The 6-isoprenoid substituted flavanones and the 6,8-disubstituted ones showed the proton signal of the hydroxyl group in the range of δ 12.4-12.5 in acetone- d_6 , while the relevant signal was observed in the range of δ 12.2-12.5 in $CDCl_3$. 2) The 8-isoprenoid substituted 6-nonsubstituted flavanones showed the relevant proton signal in the range of δ 12.0-12.2 in acetone- d_6 , and of δ 12.0-12.1 in $CDCl_3$ (Table 1).

On the other hand, Bohlmann et al.³ reported that 8-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone and its 6-isomer isolated from W. helenioides, showed the proton signals of the hydrogen-bonded hydroxyl groups at δ 12.47 and 12.13 in acetone- d_6 , respectively, and the results described in the report are inconsistent with our observations. To elucidate the inconsistency we synthesized 6-prenylated 5,7,3',4'-tetrahydroxyflavanone (1) and its 8-isomer (2) by using the unambiguous method (Scheme 1).⁴ The locations of the prenyl groups in the structures (1) and (2) were further confirmed by long-range selective proton decoupling (LSPD) experiment (Table 2). Identifications of the synthetic flavanones (1 and 2) with the natural compounds were carried out on the following facts. The 1H nmr spectra of 1 and 2 were in good agreement with the relevant spectra of Bohlmann's 8-prenylated flavanone (3) and 6-prenylated flavanone (4), respectively.⁵ Furthermore, heating of 1 in benzene with p-toluenesulfonic acid afforded 6 and a minor product (9). Heating of 2 under the same conditions afforded 9 and a minor product (6). The locations of the dimethyldihydropyrane moieties in the products (6 and 9) were confirmed by LSPD experiment (Table 2). The compounds (6) and (9) were identified as compounds (10) and (12) by comparison of the 1H nmr spectra of 6 and 9 with the reported spectra of compounds (10) and (12) (Table 3), respectively, which were derived from 3 and 4 by treatment with the same acidic conditions (Scheme 2).³ The above results obtained by the acidic treatment of 1 and 2 suggested that an isomerization must have taken place, and that the only conceivable possibility is a Wessely-Moser rearrangement⁶ in the acidic solution. From the above results the proposed structures (3) and (4) for the two natural compounds isolated from W. helenioides should be reversed each other. While several papers were reported on the isolation of 6-prenylated 5,7,3',4'-tetrahydroxyflavanone and its 8-isomer,⁷⁻¹² no detailed physico-chemical data of the compounds were described except the following an example:¹² The 8-isomer isolated from Encelia stenophylla Greene was identical with 2 by comparison of the 1H nmr spectrum of 2 with the reported data.¹² From the above results it is obvious that the hydrogen-bonded hydroxyl group of

Table 1. Chemical shifts of C-5-OH of isoprenoid substituted flavanones



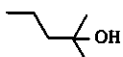
trivial name	chemical shift of 5-OH	isoprenoid group(s) on A-ring	OH-positin	others	sol. ⁺	ref.
cudraflavanone A	12.49(270 MHz, 23 °C)* 12.41(90 MHz, 35 °C)	6-prenyl [§]	5,7,2'	4',5'-Dmp	A	4
kushenol B	12.47(n.r.)	6-prenyl	5,7,2',4'	8-lavandulyl	A	18
kushenol E	12.48(90 MHz)	6,8-diprenyl	5,7,2',4'		A	19
norkurarinone (kushenol F)	12.50(90 MHz)	6-lavandulyl	5,7,2',4'		A	19,20
isokurarinone	12.52(60 MHz)	6-lavandulyl	5,7,4'	2'-OMe	A	21,22
antiarone H	12.47(400 MHz, 23 °C)	6-prenyl	5,7,3'	4'-OMe 2'-prenyl	A	23
(euchrestaflavanone B)	12.35(60 MHz)	6-prenyl	5,7,2',4'	5'-prenyl	A	26
1	12.47(400 MHz, 23 °C) 12.45(400 MHz, 35 °C) 12.42(400 MHz, 50 °C) 12.33(90 MHz, 35 °C)	6-prenyl	5,7,3',4'		A	
5	12.48(400 MHz, 23 °C)	6-(3''-OH)-isoamyl	5,7,3',4'		A	
flemiflavanone A	12.30(100 MHz)	6,8-diprenyl	5,7,2'	4'-OMe	C	24
amorinin	12.31(90 MHz)	6,8-diprenyl	5,7,3'	4',5'-Dmp	C	27
methyllinderatone	12.34(n.r.)	6-p-M	5	7-OMe	C	15,28
isolinderatone	12.34(n.r.)	6-p-M	5,7		C	15,28
no name	12.40(n.r.)	6-geranyl	5,7		C	29
bonannione A	12.40(80 MHz)	6-geranyl	5,7,4'		C	14
lonchocarpol A	12.28(360 MHz) 12.32(270 MHz)	6,8-diprenyl	5,7,4'		C	30 39
lonchocarpol B	12.29(360 MHz)	6-prenyl	5,7,4'	8-(2'',3''-di-OH)-isoamyl	C	30
lonchocarpol C	12.10(360 MHz)	6-prenyl	5,4'	7,8-Dif	C	30
hiravanone	12.34(270 MHz)	6,8-diprenyl	5,7,4'	3'-OMe	C	31
no name	12.33(270 MHz)	6,8-diprenyl	5,7,3'	4'-OMe	C	31
euchrenone a ₄	12.32(n.r.)	6,8-diprenyl	5,7	3',4'-Dmp	C	32
neolinderatone	12.48(n.r.)	6,8-di-p-M	5,7		C	33
6-C-prenyl-8-C-methylpinocembrin	12.29(300 MHz)	6-prenyl	5,7	8-Me	C	34
no name	12.41(270 MHz)	6-prenyl	5,7		C	35
no name	12.33(400 MHz)	6,8-diprenyl	5,7		C	36
flulvinervin A	12.38(300 MHz)	6-prenyl	5	7,8-Dmp	C	37
fleminone	12.28(270 MHz)	6-prenyl	5,2'	4'-OMe, 7,8-Dmp	C	38
no name	12.40(270 MHz)	6-prenyl	5,7,4'		C	39
(eriodictyol)	12.18(400 MHz, 23 °C)	none	5,7,3',4'		A	
sophoraflavanone A	12.10(100 MHz)	8-geranyl	5,7,4'		A	13
sophoraflavanone B	12.15(100 MHz)	8-prenyl	5,7,4'		A	13
sophoraflavanone G	12.20(270 MHz)	8-lavandulyl	5,7,2',4'		A	20

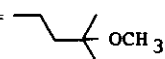
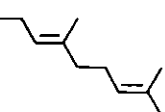
Table 1 (continued)

euchrestaflavanone A	12.10(100 MHz)	8-prenyl	5,7,4'	3'-prenyl	A	13
euchrestaflavanone B	12.20(100 MHz)	8-prenyl	5,7,2',4'	5'-prenyl	A	13
euchrestaflavanone C	12.20(100 MHz)	8-prenyl	5,7,2'	4',5'-Dmp	A	13
gancaonin E	12.14(400 MHz, 23°C)	8-prenyl	5,7,3',4'	5'-prenyl	A	40
antiarone G	12.14(400 MHz, 23°C)	8-prenyl	5,7,4'	3'-OMe 2'-prenyl	A	23
2	12.14(400 MHz, 23°C) 12.11(400 MHz, 35°C) 12.08(400 MHz, 50°C) 12.01(90 MHz, 35°C)	8-prenyl	5,7,3',4'		A	
7	12.12(400 MHz, 23°C) 12.00(90 MHz, 35°C)	8-(3''-OMe)-isoamyl	5,7,3',4'		A	
8	12.11(400 MHz, 23°C) 12.00(90 MHz, 35°C)	8-(3''-OH)-isoamyl	5,7,3',4'		A	
glabranin	11.98(80 MHz) 12.00(270 MHz)	8-prenyl	5,7		C	41 35
no name	12.10(270 MHz)	8-prenyl	5,3',4'	7-OMe	C	42
tephrowatsin C	12.05(80 MHz)	8-(3''-OH)-isoamyl	5	7-OMe	C	43
no name	12.00(270 MHz) 12.00(90 MHz, 35°C)	8-prenyl	5,7,4'		C A	39
lespedezaflavanone B	12.00(400 MHz)	8-prenyl	5,7,4'	3'-prenyl	C	25
lespedezaflavanone D	12.01(90 MHz)	8-prenyl	5,7,2',4'	5'-O-prenyl	C	44
no name	12.02(270 MHz)	8-prenyl	5,7,4'	3',5'-diprenyl	C	45
no name	12.02(270 MHz)	8-isoamyl	5,7,4'	3',5'-di- isoamyl	C	45
kushenol A	12.56(n.r.)	8-lavandulyl	5,7,2'		A	18
no name	12.40(250 MHz)	8-geranyl	5,7		C	46
no name	12.32(250 MHz)	8-geranyl	5	7-OMe	C	46
no name	12.30(400 MHz)	8-prenyl	5	7-O-prenyl	C	36

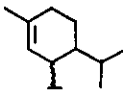
*: observed frequency of instrument and temperature of sample. n.r.: not reported.

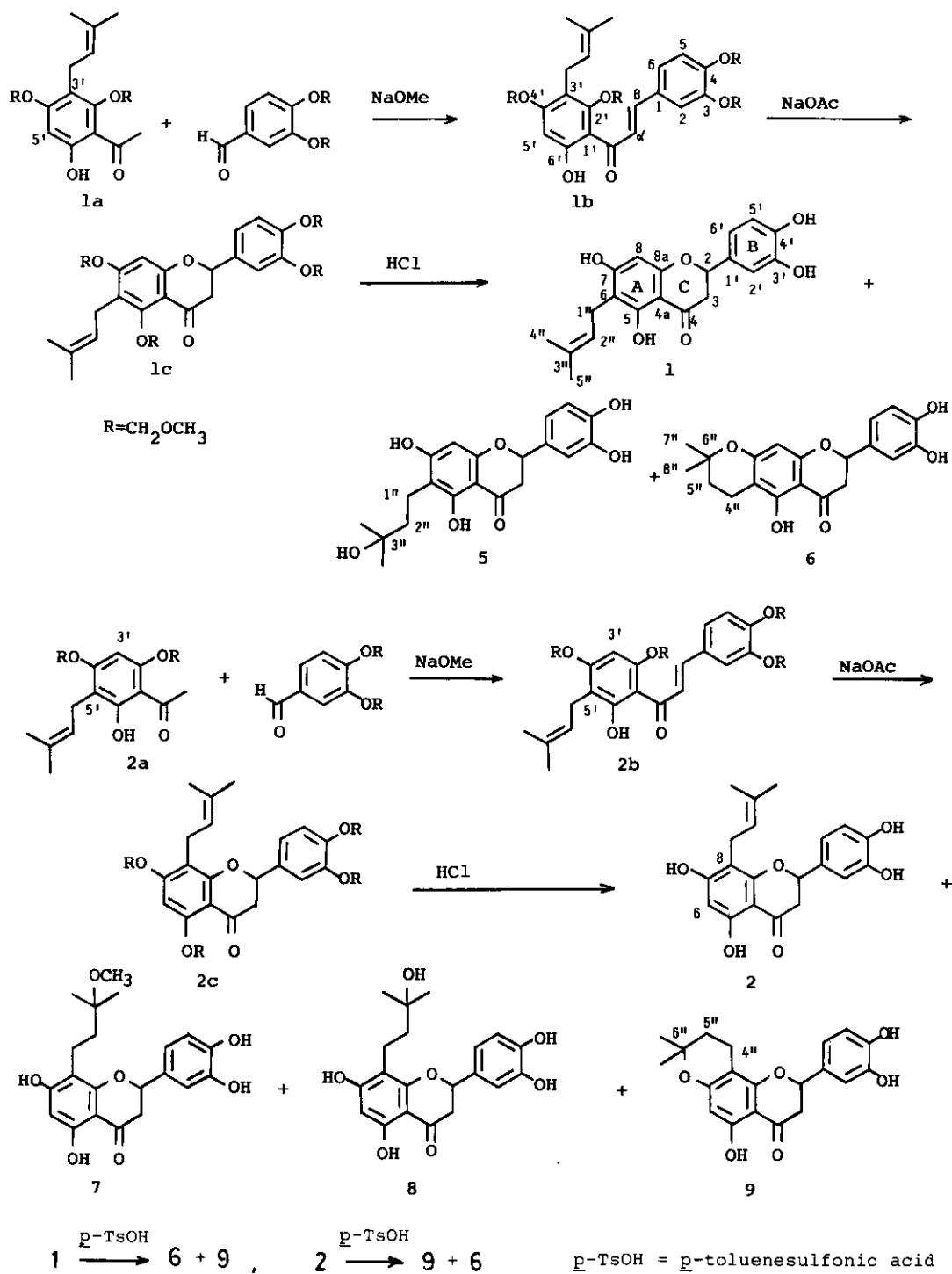
+: solvent: A=acetone-d₆, C=CDCl₃.

§: prenyl= , isoamyl= , (3''-OH)-isoamyl= ,

(3''-OMe)-isoamyl= , geranyl= ,

lavandulyl= , Dmp=6'',6''-dimethylpyrane,

Dif=3'',4''-dihydro-4''-(1'''-hydroxyisopropyl)furan, p-M (p-menthene)= 



Scheme 1.

Table 2. ^{13}C Nmr data of 1, 2, 6, and 9 in acetone- d_6 §

C	1	2
2	79.89 (br D, $^1J = \text{ca. } 148 \text{ Hz}$)*	79.75 (br D, $^1J = \text{ca. } 148 \text{ Hz}$)
3	43.74 (DD, $J = 128$ and 133 Hz)	43.52 (DD, $J = 128$ and 133 Hz)
4	197.14 (br S)	197.55 (br S)
4a	103.04 (br S)	103.29 (br S)
5	162.28 (Std, $^3J_{\text{H1}''-\text{C5}} = 4$, $^3J_{\text{5OH}-\text{C5}} = 4 \text{ Hz}$)	162.95 (Sdd, $^2J = 4$, $^3J = 5 \text{ Hz}$)
6	109.06 (Sm)	96.33 (Dd, $^1J = 161$, $^2J_{\text{5OH}-\text{C6}} = 7 \text{ Hz}$)
7	165.19 (Std, $^2J = 2$, $^3J = 4 \text{ Hz}$)	164.56 (Sm)
8	95.42 (D, $J = 162 \text{ Hz}$)	108.30 (Sm)
8a	161.98 (Sdd, $J = 1$ and 4 Hz)	161.04 (Std, $J = 4$ and 2 Hz)
1'	131.11 (Sm)	131.22 (Sm)
2'	114.70 (Ddd, $^1J = 157$, $^3J = 4$ and 7 Hz)	114.62 (Ddd, $^1J = 157$, $^3J = 4$ and 6 Hz)
3'	146.42 (Sm)	146.26 (St, $J = 6 \text{ Hz}$)
4'	146.07 (Sm)	146.05 (Std, $J = 4$ and 9 Hz)
5'	116.03 (D, $J = 159 \text{ Hz}$)	115.99 (D, $J = 157 \text{ Hz}$)
6'	119.14 (Ddd, $^1J = 160$, $^3J = 4$ and 7 Hz)	119.01 (Ddd, $^1J = 160$, $^3J = 4$ and 6 Hz)
1''	21.66 (Td, $^1J = 128$, $^2J = 4 \text{ Hz}$)	22.26 (Td, $^1J = 128$, $^2J = 4 \text{ Hz}$)
2''	123.71 (Dm, $^1J = \text{ca. } 155 \text{ Hz}$)	123.67 (Dm, $^1J = \text{ca. } 155 \text{ Hz}$)
3''	131.74 (Sm)	131.88 (Sm)
4''	17.84 (Qm, $^1J = \text{ca. } 125 \text{ Hz}$)	17.87 (Qm, $J = \text{ca. } 124 \text{ Hz}$)
5''	25.85 (Qm, $J = \text{ca. } 125 \text{ Hz}$)	25.90 (Qm, $J = \text{ca. } 125 \text{ Hz}$)

C	6	9
2	79.78 (Dm, $^1J = \text{ca. } 148 \text{ Hz}$)	79.85 (br D, $^1J = \text{ca. } 148 \text{ Hz}$)
3	43.73 (DD, $J = 128$ and 133 Hz)	43.39 (DD, $J = 128$ and 134 Hz)
4	197.42 (br S)	197.42 (br S)
4a	103.03 (br S)	103.32 (br S)
5	162.23 (Std, $^3J_{\text{H4}''-\text{C5}} = 3$, $^3J_{\text{5OH}-\text{C5}} = 5 \text{ Hz}$)	162.37 (St, $J = 4 \text{ Hz}$)
6	102.55 (br S)	97.48 (Dd, $^1J = 163$, $^2J_{\text{5OH}-\text{C6}} = 7 \text{ Hz}$)
7	163.51 (Sm)	163.58 (Std, $J = 3 \text{ Hz}$)
8	96.59 (D, $J = 164 \text{ Hz}$)	101.51 (br S)
8a	161.57 (S br d, $J = 5 \text{ Hz}$)	161.04 (br S)
1'	131.75 (br S)	131.71 (Sm)
2'	114.72 (Ddd, $^1J = 158$, $^3J_{\text{H2}-\text{C2}'} = 3$, $^3J_{\text{H6}'-\text{C2}'} = 6 \text{ Hz}$)	114.57 (Ddd, $^1J = 157$, $^3J_{\text{H2}-\text{C2}'} = 4$, $^3J_{\text{H6}'-\text{C2}'} = 6 \text{ Hz}$)
3'	146.39 (St, $J = 7 \text{ Hz}$)	146.35 (St, $J = 7 \text{ Hz}$)
4'	146.07 (br S)	146.10 (Sdd, $J = 4$ and 7 Hz)
5'	116.03 (D, $J = 159 \text{ Hz}$)	116.08 (D, $J = 159 \text{ Hz}$)
6'	119.20 (Ddd, $^1J = 160$, $^3J_{\text{H2}-\text{C6}'} = 4$, $^3J_{\text{H2}'-\text{C6}'} = 8 \text{ Hz}$)	119.00 (Ddd, $^1J = 160$, $^3J_{\text{H2}-\text{C6}'} = 4$, $^3J_{\text{H2}'-\text{C6}'} = 7 \text{ Hz}$)
4''	16.32 (Tt, $^1J = 132$, $^2J = 3 \text{ Hz}$)	16.95 (Tt, $^1J = 131$, $^2J = 3 \text{ Hz}$)
5''	32.37 (Tm, $^1J = \text{ca. } 130 \text{ Hz}$)	32.43 (Tm, $^1J = \text{ca. } 130 \text{ Hz}$)
6''	76.96 (br S)	76.80 (br S)
7''	26.62 (Qm, $^1J = \text{ca. } 128 \text{ Hz}$)	26.61 (Qm, $^1J = \text{ca. } 128 \text{ Hz}$)
8''	26.80 (Qm, $^1J = \text{ca. } 128 \text{ Hz}$)	27.19 (Qm, $^1J = \text{ca. } 128 \text{ Hz}$)

§: The following results were observed with the LSPD experiments; 1, irradiation of the 5-OH proton, C4a(br S \rightarrow Sdd, $J = 2$ and 4 Hz), C6(changed), C5(Std \rightarrow St, $J = 4 \text{ Hz}$), 2; irradiation of the 5-OH proton, C4a(br S \rightarrow Sdd, $J = 2$ and 4 Hz), C5(Sdd \rightarrow Sd, $J = 4 \text{ Hz}$), C6(Dd \rightarrow D, $J = 161 \text{ Hz}$), 6; irradiation of the 5-OH proton; C5(Std \rightarrow St, $J = 3 \text{ Hz}$), 9; irradiation of the 5-OH proton, C4a(br S \rightarrow br Sd, $J = 5 \text{ Hz}$), C6(Dd \rightarrow D, $J = 163 \text{ Hz}$), C5(St \rightarrow Sd, $J = 4 \text{ Hz}$).

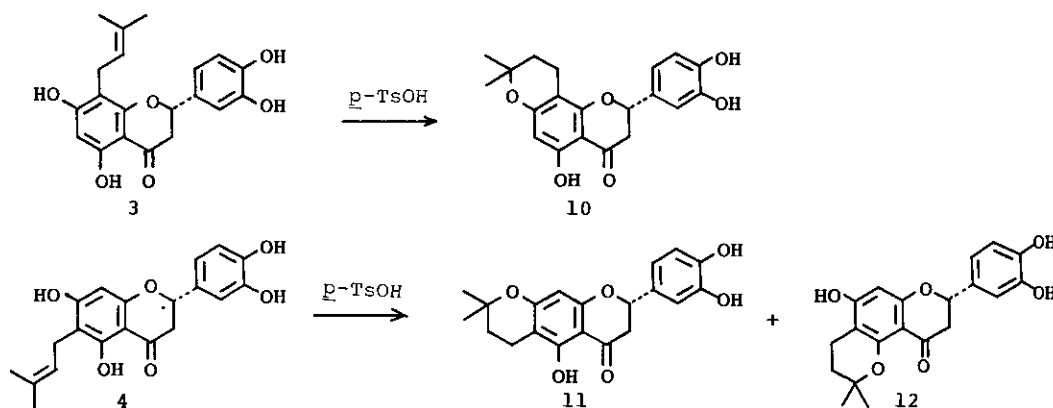
*: Capital letters refer to the pattern resulting from directly bonded proton(s) and lowercase letters to long-range ^{13}C - ^1H coupling.

6-isoprenoid substituted flavanones shifted more downfield than that of 6-non-substituted flavanones. In order to determine the location of the isoprenoids moiety in the A-ring of flavonoid, some methods have been reported such as the technique using the observation of ^{13}C - ^1H long-range coupling.¹³⁻¹⁵ The observation for the chemical shift of the hydroxyl group may be a facile method to estimate the isoprenoid moiety in the A-ring of flavanone.

Table 3. ^1H Nmr data of 6, 9, and 10-12 (in CDCl_3 , 400 MHz)

	6	10*	11*	9	12*
2-H	5.26 (dd, J=3 and 12.5 Hz)	5.26	5.32	5.32 (dd, J=3 and 13 Hz)	5.33
3-H	2.76 (dd, J=3 and 17 Hz)	2.76	2.77	2.78 (dd, J=3 and 17 Hz)	2.77
3-H	3.03 (dd, J=12.5 and 17 Hz)	3.02	2.99	3.01 (dd, J=13 and 17 Hz)	3.00
6-H	—	5.92	—	5.97 (s)	—
8-H	5.93 (s)	—	5.93	—	5.97
2'-H	6.99 (br s)	6.98	6.99	7.01 (d, J=1.5 Hz)	7.02
5'-H	6.90 (d, J=8 Hz)	6.89	6.90	6.91 (d, J=1.5 and 7 Hz)	6.92
6'-H	6.87 (br d, J=8 Hz)	6.89	6.87	6.88 (dd, J=1.5 and 7 Hz)	6.89
4''-H ₂	2.61 (br t, J=6.5 Hz)	2.61	2.62	2.58 (m)	2.59
5''-H ₂	1.79 (t, J=6.5 Hz)	1.78	1.79	1.76 (m)	1.75
6''-CH ₃	1.338 (s)	1.33	1.33	1.33 (s)	1.32
6''-CH ₃	1.343 (s)	1.34	1.34	1.35 (s)	1.34
OH	12.37 (s)	12.37	12.34	11.74 (s)	11.74

*: Data from Bohlmann *et al.* (ref.3, J(Hz): 2,3=3 and 12.5; 3,3=17; 2',6'=1.5; 5',6'=8; 4'',5''=7)



Scheme 2. Cyclized reactions of 3 and 4 with *p*-toluenesulfonic acid by Bohlmann *et al.*³

EXPERIMENTAL

Abbreviations; s = singlet, d = doublet, dd = double doublet, t = triplet, m = multiplet, br = broad. The general procedures followed as described in our previous paper.¹⁶ The following instruments were used: melting points; Yazawa's micromelting point apparatus (hot-stage type), ^1H nmr spectra; JEOL JNM-GX-400 NMR Spectrometer and Hitachi R-900 NMR Spectrometer (90 MHz, CW mode), ^{13}C nmr spectra; JEOL JNM-GX-400 NMR Spectrometer, mass spectra: JEOL JMS-D-300 Mass Spectrometer and JEOL JMS-DX-303 Mass Spectrometer.

3'-(3'',3''-Dimethylallyl)-6'-hydroxy-3,4,2',4'-tetrakis(methoxymethoxy)chalcone (1b)

To a mixture of 2',4'-bis(methoxymethoxy)-3'-(3'',3''-dimethylallyl)-6'-hydroxyacetophenone (1a, 50 mg, 0.15 mmol)⁴ and 3,4-bis(methoxymethoxy)benzaldehyde (100 mg, 0.44 mmol)¹⁷ in methanol (2 ml) was added 28% NaOMe in methanol (1 ml). The mixture was allowed to stand at room temperature for 24 h, and then acetic acid (1 ml) was added to the solution. The reaction mixture was treated as usual and the product was purified by preparative tlc (silica gel, *n*-hexane:acetone=3:1) to give 1b (44 mg,

54%). Compound (1b) showed the following data: yellow amorphous solid. EI-MS, m/z (rel. int.): 533 $[M+1]^+$ (2%), 532 $[M]^+$ (5), 488 (2), 487 (3), 455 (3), 263 (3), 206 (6), 175 (11), 149 (7), 69 (10), 45 (100). HR-MS, m/z : 532.2318 $[M]^+$ ($C_{28}H_{36}O_{10}$ requires: 532.2308). 1H Nmr ($CDCl_3$, 400 MHz): δ 1.70, 1.79 (each 3H, br s, C-3"-CH₃), 3.37 (2H, br d, J = 7 Hz, C-1"-Hx2), 3.46, 3.49, 3.53, 3.54 (each 3H, s, OCH₂OCH₃), 4.89 (2H, s, OCH₂OCH₃), 5.19 (1H, br t, J = 7 Hz, C-2"-H), 5.23, 5.28, 5.29 (each 2H, s, OCH₂OCH₃), 6.50 (1H, s, C-5'-H), 7.19 (1H, d, J = 8.5 Hz, C-5-H), 7.29 (1H, dd, J = 2 and 8.5 Hz, C-6-H), 7.46 (1H, d, J = 2 Hz, C-2-H), 7.72 (1H, d, J = 15.5 Hz, C-8-H), 7.79 (1H, d, J = 15.5 Hz, C-8-H), 12.92 (1H, s, OH).

6-(3",3"-Dimethylallyl)-5,7,3',4'-tetrakis(methoxymethoxy)flavanone (1c)

A mixture of 1b (44 mg, 0.08 mmol) and sodium acetate (0.5 g, 6.1 mmol) in methanol (5 ml) was allowed to stand at 50°C for 3 h. The reaction mixture was treated as usual, and the product was purified by preparative tlc (silica gel, n-hexane:acetone=4:1) to give 1c (30 mg, 68%). Compound (1c) showed the following data: mp 82-83°C (colorless prisms, crystallized from methanol). EI-MS, m/z (rel. int.): 533 $[M+1]^+$ (1%), 532 $[M]^+$ (3), 500 (18), 488 (4), 487 (10), 455 (16), 441 (4), 379 (3), 263 (11), 219 (6), 175 (15), 148 (10), 69 (3), 45 (100). HR-MS, m/z : 532.2315 $[M]^+$ ($C_{28}H_{36}O_{10}$ requires: 532.2308). 1H Nmr ($CDCl_3$, 90 MHz): δ 1.66, 1.78 (each 3H, br s, C-3"-CH₃), 2.70 (1H, dd, J = 4 and 17 Hz, C-3-H), 3.01 (1H, dd, J = 13 and 17 Hz, C-3-H), 3.40 (2H, br d, J = 7 Hz, C-1"-Hx2), 3.44 (3H, s, OCH₂OCH₃), 3.52 (6H, s, OCH₂OCH₃x2), 3.60 (3H, s, OCH₂OCH₃), 5.18 (2H, s, OCH₂OCH₃), 5.22 (6H, s, OCH₂OCH₃), 5.32 (1H, dd, J = 4 and 13 Hz, C-2-H), 6.52 (1H, s, C-8-H), 7.00 (1H, dd, J = 2 and 8.5 Hz, C-6'-H), 7.17 (1H, d, J = 8.5 Hz, C-5'-H), 7.23 (1H, d, J = 2.5 Hz, C-2'-H): The olefinic proton signal of the 3",3"-dimethylallyl group overlapped with the signals of the methylene protons of the methoxymethoxy groups.

Preparation of 6-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone (1) and related compounds (5 and 6) from 1c

To a solution of 1c (29 mg) in methanol (2.5 ml) was added 3N HCl (0.5 ml). The mixture was refluxed for 15 min, and then treated as usual. The reaction product was purified by preparative tlc (silica gel, n-hexane:acetone=3:2) to give 1 (10 mg, 51.5%), 6-(3"-hydroxyisoamyl)-5,7,3',4'-tetrahydroxyflavanone (5, 3 mg, 15%) and 6",6"-dimethyl-4",5"-dihydropyrano(2",3":7,6)-5,3',4'-trihydroxyflavanone (6, 0.5 mg, 2.5%). Compound (1) showed the following data: mp 192-193°C (colorless prisms, crystallized from $CHCl_3$). EI-MS m/z (rel. int.): 357 $[M+1]^+$ (24%), 356 $[M]^+$ (100), 341 (23), 313 (20), 301 (38), 288 (6), 234 (9), 221 (14), 220 (18), 219 (20), 205 (53), 192 (35), 177 (30), 165 (100), 136 (28), 123 (16), 69 (16). Compound (5) showed the following data: mp 179-181°C (colorless prisms, crystallized from benzene-acetone). EI-MS, m/z (rel. int.): 375 $[M+1]^+$ (1%), 374 $[M]^+$ (5), 356 (36), 341 (8), 301 (49), 300 (67), 299 (9), 282 (14), 221 (10), 220 (9), 219 (5), 205 (17), 192 (11), 179 (14), 165 (100), 136 (26), 123 (15). HR-MS, m/z : 374.1328 $[M]^+$ ($C_{20}H_{22}O_7$ requires: 374.1366). 1H Nmr (acetone- d_6 , 400 MHz): δ 1.24 (6H, s, C-3"-CH₃x2), 1.65 (2H, m, C-2"-Hx2), 2.66 (2H, m, C-1"-Hx2), 2.71 (1H, dd, J = 3 and 17 Hz, C-3-H), 3.12 (1H, dd, J = 13 and 17 Hz, C-3-H), 5.36 (1H, dd, J = 3 and 13 Hz, C-2-H), 6.01 (1H, s, C-8-H), 6.86 (2H, br s, C-5'-H and C-6'-H), 7.03 (1H, br s, C-2'-H), 12.48 (1H, s, OH). Compound (6) showed the following data: amorphous solid. EI-MS, m/z (rel. int.): 357 $[M+1]^+$ (23%), 356 $[M]^+$ (100), 341 (7), 301 (22), 300 (7), 247 (15), 234 (29), 221 (34), 220 (28), 205 (18), 192 (19), 177 (11), 165 (76), 136 (21), 123 (13). HR-MS, m/z : 356.1242 $[M]^+$ ($C_{20}H_{20}O_6$ requires: 356.1260).

5'-(3'',3''-Dimethylallyl)-6'-hydroxy-3,4,2',4'-tetrakis(methoxymethoxy)chalcone (2b)

To a mixture of 2',4'-bis(methoxymethoxy)-5'-(3'',3''-dimethylallyl)-6'-hydroxyacetophenone (2a, 98 mg, 0.3 mmol)⁴ and 3,4-bis(methoxymethoxy)benzaldehyde (220 mg, 0.97 mmol)¹⁷ in methanol (2 ml) was added 28% NaOMe in methanol (1 ml). The mixture was allowed to stand at room temperature for 6 h, and then acetic acid (1 ml) was added to the mixture. The reaction mixture was treated as usual, and the product was purified by preparative tlc (silica gel, *n*-hexane:acetone=3:1) to give 2b (91 mg, 56.5%). Compound (2b) showed the following data: mp 111-112°C (yellow needles, crystallized from methanol). EI-MS, *m/z* (rel. int.): 533 [M+1]⁺ (4%), 532 [M]⁺ (13), 487 (23), 455 (5), 263 (16), 219 (8), 175 (19), 69 (3), 45 (100). HR-MS, *m/z*: 532.2324 [M]⁺ (C₂₈H₃₆O₁₀ requires: 532.2308). ¹H Nmr (CDCl₃, 90 MHz): δ 1.69, 1.80 (each 3H, br s, C-3''-CH₃), 3.34 (2H, br d, J = 7 Hz, C-1''-Hx2), 3.48 (3H, s, OCH₂OCH₃), 3.53 (9H, s, OCH₂OCH₃x3), 5.23 (2H, s, OCH₂OCH₃), 5.26 (6H, s, OCH₂OCH₃x3), 6.40 (1H, s, C-3'-H), 7.18 (2H, br s, C-5-H and C-6-H), 7.47 (1H, br s, C-2-H), 7.68 (1H, d, J = 16 Hz, C-α-H), 7.88 (1H, d, J = 16 Hz, C-β-H), 13.74 (1H, s, OH): The olefinic proton of the 3'',3''-dimethylallyl group overlapped with the signals of the methylene protons of the methoxymethoxy groups.

8-(3'',3''-Dimethylallyl)-5,7,3',4'-tetrakis(methoxymethoxy)flavanone (2c)

A mixture of 2b (90 mg, 0.17 mmol) and sodium acetate (0.5 g, 6.1 mmol) in methanol (5 ml) was allowed to stand at 50°C for 20 h. The reaction mixture was treated as usual, and the product was purified by preparative tlc (silica gel, *n*-hexane:acetone=2:1) to give 2c (46 mg, 51%). Compound (2c) showed the following data: amorphous solid. EI-MS, *m/z* (rel. int.): 533 [M+1]⁺ (2%), 532 [M]⁺ (7), 488 (11), 487 (10), 455 (4), 443 (4), 441 (6), 373 (3), 265 (3), 263 (16), 175 (8), 148 (8), 69 (6), 45 (100). HR-MS, *m/z*: 532.2339 [M]⁺ (C₂₈H₃₆O₁₀ requires: 532.2308). ¹H Nmr (CDCl₃, 90 MHz): δ 1.67 (6H, br s, C-3''-CH₃x2), 2.77 (1H, dd, J = 4 and 16 Hz, C-3-H), 3.03 (1H, dd, J = 12 and 16 Hz, C-3-H), 3.35 (2H, br d, J = 7 Hz, C-1''-Hx2), 3.49 (3H, s, OCH₂OCH₃), 3.52 (9H, s, OCH₂OCH₃x3), 5.24 (8H, s, OCH₂OCH₃x4), 5.35 (1H, dd, J = 4 and 12 Hz, C-2-H), 6.56 (1H, s, C-6-H), 7.02 (1H, dd, J = 2 and 8.5 Hz, C-6'-H), 7.18 (1H, d, J = 8.5 Hz, C-5'-H), 7.28 (1H, d, J = 2 Hz, C-2'-H): The olefinic proton signal of the 3'',3''-dimethylallyl group overlapped with the methylene proton signals of the methoxymethoxy groups.

Preparation of 8-(3'',3''-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone (2) and related compounds (7, 8, and 9) from 2c

To a solution of 2c (43 mg) in methanol (2.5 ml) was added 3N HCl (0.5 ml). The mixture was refluxed for 25 min, and then treated as usual. The reaction product was purified by preparative tlc (silica gel, *n*-hexane:acetone=2:1) to give 2 (12 mg, 42%), 8-(3''-methoxyisoamyl)-5,7,3',4'-tetrahydroxyflavanone (7, 7 mg, 32%), 8-(3''-hydroxyisoamyl)-5,7,3',4'-tetrahydroxyflavanone (8, 3 mg, 10%), and 6'',6''-dimethyl-4'',5''-dihydropyrano(2'',3'':7,8)-5,3',4'-trihydroxyflavanone (9, 4 mg, 14%). Compound (2) showed the following data: mp 199-202°C (colorless prisms, crystallized from CHCl₃). EI-MS, *m/z* (rel. int.): 357 [M+1]⁺ (24%), 356 [M]⁺ (100), 341 (23), 313 (21), 301 (29), 288 (6), 234 (8), 221 (12), 220(18), 219 (27), 205 (54), 192 (36), 177 (35), 165 (88), 136 (26), 123 (12), 69 (15). Compound (7) showed the following data: amorphous solid. EI-MS, *m/z* (rel. int.): 389 [M+1]⁺ (2%), 388 [M]⁺ (8), 356 (60), 341 (16), 313 (12), 301 (65), 300 (72), 282 (11), 247 (4), 234 (8), 221 (14), 220 (16), 205 (27), 192 (19), 179 (11), 177 (13), 165 (100), 136 (26), 123 (14), 73 (51). HR-MS, *m/z*: 388.1498 [M]⁺ (C₂₁H₂₄O₇ requires: 388.1522). ¹H Nmr (acetone-d₆, 400 MHz): δ 1.11 (6H, s,

C-3"-CH₃x2), 1.62 (2H, m, C-2"-Hx2), 2.56 (2H, t, J = 8 Hz, C-1"-Hx2), 2.77 (1H, dd, J = 3 and 17 Hz, C-3-H), 3.07 (1H, dd, J = 12.5 and 17 Hz, C-3-H), 3.07 (3H, s, C-3"-OCH₃), 5.42 (1H, dd, J = 3 and 12.5 Hz, C-2-H), 6.02 (1H, s, C-6-H), 6.87 (1H, d, J = 8.5 Hz, C-5'-H), 6.92 (1H, dd, J = 2 and 8.5 Hz, C-6'-H), 7.07 (1H, d, J = 2 Hz, C-2'-H), 12.12 (1H, s, OH). Compound (8) showed the following data: amorphous solid. EI-MS, m/z (rel. int.): 375 [M+1]⁺ (1%), 374 [M]⁺ (5), 356 (36), 341 (7), 299 (11), 300 (66), 282 (14), 221 (10), 220 (10), 219 (5), 205 (16), 192 (12), 165 (100), 136 (26), 123 (14). HR-MS m/z : 374.1372 [M]⁺ (C₂₀H₂₂O₇ requires: 374.1366). ¹H Nmr (acetone-d₆, 400 MHz): δ 1.19 (6H, s, C-3"-CH₃x2), 1.65 (2H, m, C-2"-Hx2), 2.65 (2H, t, J = 8 Hz, C-1"-Hx2), 2.79 (1H, dd, J = 3 and 17 Hz, C-3-H), 3.08 (1H, dd, J = 12 and 17 Hz, C-3-H), 5.42 (1H, dd, J = 3 and 12 Hz, C-2-H), 6.00 (1H, s, C-6-H), 6.86 (1H, d, J = 8 Hz, C-5'-H), 6.91 (1H, dd, J = 2 and 8 Hz, C-6'-H), 7.07 (1H, d, J = 2 Hz, C-2'-H), 12.11 (1H, s, OH). Compound (9) showed the following data: mp 238-239°C (colorless prisms, crystallized from benzene-acetone), EI-MS, m/z (rel. int.) 357 [M+1]⁺ (24%), 356 [M]⁺ (100), 341 (4), 301 (23), 300 (6), 247 (16), 234 (30), 221 (39), 220 (29), 205 (16), 192 (27), 177 (15), 165 (100), 136 (28), 123 (11), 78 (66). HR-MS m/z : 356.1247 [M]⁺ (C₂₀H₂₀O₆ requires: 356.1260).

Preparation of 6 and 9 from 6-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone (1)

A mixture of 1 (15 mg) and *p*-toluenesulfonic acid (10 mg) in benzene (4 ml) was refluxed for 30 min.³ The reaction product was purified by preparative tlc (silica gel, *n*-hexane:ether=1:1) to give 6 (8 mg, 53%) and 9 (less than 0.5 mg). Compound (6) thus obtained was identical with 6 derived from 1c on comparison of ¹H nmr spectra and tlc analysis. Compound (9) thus obtained was identical with 9 derived from 2c by comparison of the ¹H nmr spectra and tlc analysis.

Preparation of 9 and 6 from 8-(3",3"-dimethylallyl)-5,7,3',4'-tetrahydroxyflavanone (2)

A mixture of 2 (25 mg) and *p*-toluenesulfonic acid (16 mg) in benzene (6 ml) was refluxed for 30 min.³ The reaction product was purified by preparative tlc (*n*-hexane:ether=1:1) to give 9 (18 mg, 72%, mp 237-239°C) and 6 (3 mg, 12%). Compound (9) thus obtained was identical with 9 derived from 2c on comparison of the ¹H nmr spectra and tlc analysis. Compound (6) thus obtained was identical with 6 derived from 1c by comparison of the ¹H nmr spectra and tlc analysis.

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REFERENCES AND NOTES

1. Part 3 in the series 'NMR Spectra of Isoprenoid Substituted Phenols.' The reference 'T. Fukai and T. Nomura, *Heterocycles*, 1989, 29, 2379' is regarded as Part 1 in this series. Reference 2 is regarded as Part 2.

2. T. Fukai, Q.-H. Wang, M. Takayama, and T. Nomura, Heterocycles, 1990, **31**, 373.
3. F. Bohlmann, C. Zdero, H. Robinson, and R. M. King, Phytochemistry, 1981, **20**, 2245.
4. T. Fujimoto and T. Nomura, Planta Med., 1985, 190.
5. Direct comparison between the synthetic and the natural flavanones could not be performed since the specimens of the latter have been exhausted.
6. T. R. Seshadri, Tetrahedron, 1959, **6**, 169.
7. F. Bohlmann, C. Zdero, R. M. King, and H. Robinson, Planta Med., 1984, 195.
8. S. McCormick, K. Robson, and B. Bohm, Phytochemistry, 1985, **24**, 1614.
9. S. McCormick, K. Robson, and B. Bohm, Phytochemistry, 1986, **25**, 1723.
10. S. P. McCormick, K. A. Robson, J. Maze, and B. A. Bohm, Phytochemistry, 1987, **26**, 2421.
11. B. A. Bohm, J. B. Choy, and Y.-M. Lee, Phytochemistry, 1989, **28**, 501.
12. P. Proksch, U. Politt, E. Wollenweber, V. Wray, and C. Clark, Planta Med., 1988, 542.
13. Y. Shirataki, I. Yokoe, M. Endo, and M. Komatsu, Chem. Pharm. Bull., 1985, **33**, 444, and references cited therein.
14. M. Bruno, G. Savona, L. Lamartina, and F. Lentini, Heterocycles, 1985, **23**, 1147.
15. K. Ichino, H. Tanaka, and K. Ito, Heterocycles, 1990, **31**, 549.
16. T. Fukai and T. Nomura, Phytochemistry, 1989, **27**, 259.
17. T. Oyamada and H. Baba, Bull. Chem. Soc. Jpn., 1966, **39**, 507.
18. L.-J. Wu, T. Miyase, A. Ueno, M. Kuroyanagi, T. Noro, and S. Fukushima, Chem., Pharm. Bull., 1985, **33**, 3231.
19. L.-J. Wu, T. Miyase, A. Ueno, M. Kuroyanagi, T. Noro, and S. Fukushima, Yakugaku Zasshi, 1985, **105**, 736.
20. Y. Shirataki, I. Yokoe, M. Noguchi, T. Tomimori, and M. Komatsu, Chem. Pharm. Bull., 1988, **36**, 2220.
21. K. Kyogoku, K. Hatayama, and M. Komatsu, Chem. Pharm. Bull., 1973, **21**, 2733.
22. M. Mizuno, M. Iinuma, T. Tanaka, Y. Shirataki, I. Yokoe, and M. Komatsu, 'Abstract Papers of 36th Annual Meeting of the Japanese Society of Pharmacognocny, 1989, Kumamoto, p. 155.
23. Y. Hano, P. Mitsui, and T. Nomura, Heterocycles, 1990, **31**, 1315.
24. S. Sivarambabu, J. M. Rao, and K. V. J. Rao, Indian J. Chem., 1979, **17B**, 85.
25. M. Wang, J. Li, and W. Liu, Phytochemistry, 1987, **26**, 1218.
26. F. D. Monache, F. Ferrari, and M. Pomponi, Phytochemistry, 1984, **23**, 1489.

27. Z. Rózsa, J. Hohmann, J. Reisch, I. Mester, and k. Szendrei, Phytochemistry, 1982, 21, 1827.
28. K. Ichino, H. Tanaka, and K. Ito, Chem. Pharm. Bull., 1989, 37, 944.
29. J. Reisch, M. Gombos, K. Szendrei, and I. Novak, Arch. Pharmaz., 1976, 309, 152.
30. V. Roussis, S. A. Ampofo, and D. F. Wiemer, Phytochemistry, 1987, 26, 2371.
31. C. Ito, K. Sato, T. Oka, M. Inoue, M. Ju-Ichi, M. Omura, and H. Furukawa, Phytochemistry, 1989, 28, 3562.
32. M. Mizuno, K. Tamura, T. Tanaka, and M. Iinuma, Chem. Pharm. Bull., 1989, 37, 195.
33. K. Ichino, H. Tanaka, and K. Ito, Chem. Pharm. Bull., 1989, 37, 1426.
34. M. Kaouadji, P. Ravanel, and A.-M. Mariotte, J. Nat. Prod., 1986, 49, 153.
35. F. Bohlmann and W.-R. Abraham, Phytochemistry, 1979, 18, 1851.
36. F. Bohlmann and L. N. Misra, Planta Med., 1984, 271.
37. E. Venkata Rao, G. Venkataratnam, and C. Vilain, Phytochemistry, 1985, 24, 2427.
38. K. Nageswara Rao and G. Srimannarayana, Phytochemistry, 1983, 22, 2287.
39. C. Ito, T. Mizuno, M. Matsuoaka, Y. Kimura, K. Sato, I. Kajiura, M. Omura, M. Ju-Ichi, and H. Furukawa, Chem. Pharm. Bull., 1988, 36, 3292.
40. T. Fukai, Q.-H. Wang, and T. Nomura, Heterocycles, 1989, 29, 1369.
41. L. A. Mitscher, G. S. R. Rao, I. Khanna, T. Veysoglu, and S. Drake, Phytochemistry, 1983, 22, 573.
42. F. Bohlmann and J. Jakupovic, Phytochemistry, 1979, 18, 1189.
43. F. Gómez, L. Quijano, J. S. Calderón, C. Rodríguez, and T. Ríos, Phytochemistry, 1985, 24, 1057.
44. J. Li and M. Wang, Phytochemistry, 1989, 28, 3564.
45. P. Baruah, N. C. Barua, R. P. Sharma, J. N. Baruah, P. Kulanthaivel, and W. Herz, Phytochemistry, 1984, 23, 443.
46. T. Somleva and I. Ognyanov, Planta Med., 1985, 219.

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