RADIOIMMUNOASSAY OF IRIDOID GLUCOSIDES : PART 1 GENERAL METHODS FOR PREPARATION OF THE HAPTENS AND THE CONJUGATES WITH A PROTEIN OF THIS SERIES OF GLUCOSIDES ¹³

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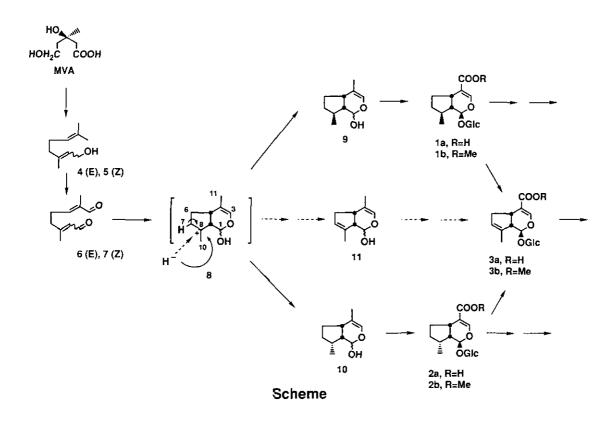
<u>Abstract</u> — In order to obtain antibodies specific to three key intermediates, 7-deoxyloganin (<u>1b</u>), 7-deoxy-8-epi-loganin (<u>2b</u>) and 10-deoxygeniposide (<u>3b</u>), in biosynthesis of iridoid glucosides in behalf of their microdetermination by a radioimmunoassay technique in various plant extracts, we have synthesized, starting with suitable natural iridoid glucosides, three optically pure intermediates from which we in turn prepared their haptens and conjugates to bovine serum albumin.

Though it is known that iridoid glucosides are distributed widely in dicotyledons²⁰, the iridoid constituents of many plants have not been yet investigated. In fact, during the past decade the number of new glucosides increased rapidly.^{3, 4, 5)} The vast majority of these glucosides belong to a group with a carboxy group at C-4 position of iridane as in 7-deoxyloganic acid (<u>1a</u>) or its decarboxylated one which is biosynthetically derived from the former. Their biosynthetic pathway (Scheme) was proposed as follows⁶⁻⁶⁹: 1) Three iridoid isomers (<u>9, 10, 11</u>) having the iridane skeleton are formed via cyclization of 10-oxogeranial (<u>6</u>) or its isomer, 10-oxoneral (<u>7</u>), generated through respective oxidation of geraniol (<u>4</u>) or nerol (<u>5</u>) which are derived from mevalonic acid (MVA). 2) These isomers (<u>9, 10, 11</u>) are oxidized followed by glucosidation to be led to three glucosidic intermediates, 7-deoxyloganic acid (<u>1a</u>), 7-deoxy-8-epi-loganic acid (<u>2a</u>) and 10-deoxygeniposidic acid (<u>3a</u>) or their methyl esters (<u>1b</u>, <u>2b</u>, <u>3b</u>) (Each free acid and its methyl ester are considered to be biosynthetically equivalent). 3) Each intermediate gives rise to various iridoid glucosides via several oxidation stages.

A cyclization of <u>6</u> or <u>7</u> provides iridodial cation (<u>8</u>), a supposed intermediate, which in turn forms three biosynthetic intermediates (<u>9</u>, <u>10</u>, <u>11</u>) by mode of quenching the cation: the attack

of a hydride to $\underline{9}$ and $\underline{10}$ or the deprotonation to $\underline{11}$. The two intermediates (<u>1b</u>, <u>2b</u>), of these intermediates, precede <u>3b</u> in iridoid biosynthesis, because <u>1b</u> and <u>2b</u> were incorporated to <u>3b</u> in plants and their calluses.^{B, 9)} However, the biosynthetic route through compound <u>11</u> to <u>3</u> is not negligible entirely.

The aim of this study is to investigate through which of these three intermediates $(\underline{1}, \underline{2}, \underline{3})$ iridoid glucosides having a carboxy group at C-4 position present in various plants were biosynthesized and furthermore the distribution of iridoid glucosides in dicotyledons through the detection of these biosynthetic intermediates.



As methods for determining a trace amount of a target intermediate in plant extracts, gas chromatographic (GC), GC - mass spectrometric, high performance liquid chromatographic and immunogenic techniques containing radioimmunoassay have been used. In this study, the radioimmunoassay technique was preferred for the determination of iridoid glucosides because of sensitivity and specificity as well as direct applicability to glucosidic extracts. The present paper deals with the preparation of haptens and conjugates with a protein of three optically pure intermediates (1, 2, 3) for establishing their radioimmunoassay system.

Design for the synthesis of immunogenic conjugates

The above-mentioned biosynthetic intermediates $(\underline{1}, \underline{2}, \underline{3})$ of iridoid glucosides could not be immunogenic by themselves because of their low molecular weight. Thus, in order to obtain antibodies against them, it is necessary to conjugate them covalently to an antigenic protein or synthetic polypeptide carrier, before immunization, for which we chose bovine serum albumin (BSA). Moreover, for the preparation of three antibodies specific to three compounds it is better to bind them to BSA at the remote positions from the position to be districtly recognized of their molecule. Thus two types of conjugates with regard to three glucosides ($\underline{1}, \underline{2}, \underline{3}$) were prepared. In the type I ($\underline{13}, \underline{20}, \underline{28}$), glucosides was bound to a BSA through the carboxy group at C-4 position of the aglucone moiety and in the type II ($\underline{16}, \underline{25}, \underline{31}$), through the carboxy group derived from the primary hydroxy group at C-6' position of the sugar moiety. In both cases γ -aminobutyric acid (GABA) was used as a spacer binding two parts.

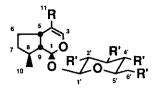
Preparation of six immunogenic conjugates with BSA

In general, a chemically and optically pure substance is necessary for the preparation of the specific antibody against it and for its radioimmunological analysis. Three above-mentioned biosynthetic intermediates $(\underline{1}, \underline{2}, \underline{3})$ of iridoid glucosides are different only in the geometry at C-8 position. So we paid our attention to the preparation of three optically or regiospecifically pure compounds $(\underline{1}, \underline{2}, \underline{3})$.

1) 7-Deoxyloganin-BSA conjugates 13 and 16

In the previous studies, 7-deoxyloganin (<u>1b</u>) was prepared through the hydrogenation including hydrogenolysis of geniposide (<u>37</u>)¹⁰, and the acetate (<u>38</u>).^{11,12} However, it had been proved from the detailed analysis of the high resolution NMR spectrum that the product contained 5 - 10% of 7-deoxy-8-epi-loganin (<u>2b</u>).

As the starting material for the alternative preparation of <u>1b</u>, now, verbenalin (<u>39</u>) was chosen because it has 8 β -methyl group which would be unaffected during its conversion to <u>1b</u> and furthermore is readily obtainable in large quantities from the leaves of <u>Symplocos glauca</u> Koidz. The tetraacetate (<u>40</u>) was treated with ethylenedithiol-BF₃ to give thioketal (<u>41</u>).



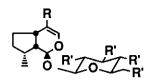
 1a. R=COOH,
 R'=OH

 1b. R=COOMe,
 R'=OH

 12. R=COOMe,
 R'=OAC

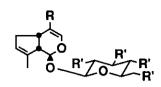
 13. R=CONH(CH₂)₃CO-BSA, R'=OH
 14. R=CONH(CH₂)₃COOMe, R'=OH

 15. R=CONH(CH₂)₃COOH,
 R'=OH



2a. R=COOH,	R'=OH
2b. R=COOMe,	R'=OH
20. R=CONH(CH2)3CO-BSA,	R'=OH
21. R=CHO,	R'=OH
22. R=CHO;	R'=OAc
23. R=COOH,	R'≞OAc

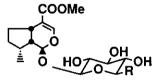
24 R=CONH(CH2)3COOH, R'=OH



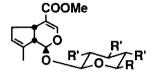
3a. R=COOH,	R'=OH
3b. R=COOMe,	R'=OH
28. R=CONH(CH ₂) ₃ CO	-BSA, R'=OH
29. R=COOMe.	R'=OAc

30. R=CONH(CH2)3COOH, R'=OH

- 16. R=CONH(CH₂)₃CO-BSA
- 17. R=COOH
- 18. R=CONH(CH₂)₃COOMe
- 19. R=CONH(CH2)3COOH



- 25. R=CONH(CH₂)₃CO-BSA 26. R=CONH(CH₂)₃COOH
- 27. R=COOH
- 27. H=COUH



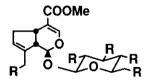
- 31. R=CONH(CH2)3CO-BSA, R'=OH

 32. R=COOH,
 R'=OH

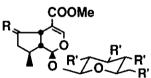
 33. R=CH2OTr,
 R'=OAC

 34. R=COOH,
 R'=OAC

 35. R=CONH(CH2)3COOH,
 R'=OH
- 36. R=CONH(CH2)3COOMe, R'=OAc



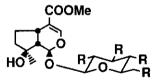
37. R±OH 38. R=OAc



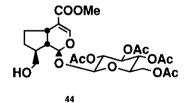
 39. R=O,
 R'=OH

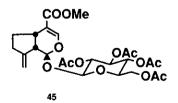
 40. R=O,
 R'=OAc

 41. R=-S(CH₂)₂S-,
 R'=OAc



42. R≃OH 43. R=OAc





Figure

Desulfurization of <u>41</u> with Raney Ni provided optically pure 7-deoxyloganin tetraacetate (<u>12</u>) which was led to the haptens of types I and II. 7-Deoxyloganic acid (<u>1a</u>) obtained through hydrolysis of <u>12</u> was condensed with methyl γ - aminobutyric acid (Me-GABA) using 1-ethyl-3-[3-(dimethyl-amino)propyl]-carbodiimide hydrochloride (EDCI) to afford 11-GABA amide methyl ester (<u>14</u>). Hydrolysis of <u>14</u> afforded 11-GABA amide (<u>15</u>), the hapten of type I. On the other hand, <u>12</u> was subjected to Zemplen deacetylation¹³⁾ to give 7-deoxyloganin (<u>1b</u>). Regioselective oxidation of <u>1b</u> with platinum oxide in the presence of alkali gave 6'-acid (<u>17</u>) in 75 % yield. In the same way as described for the preparation of <u>15</u>, <u>17</u> was converted, through 6'-GABA amide methyl ester (<u>18</u>), into 6'-GABA amide (<u>19</u>), the hapten of type II. Conjugation of two haptens (<u>15</u>, <u>19</u>) to BSA with EDCI gave two conjugates, <u>13</u> and <u>16</u>, respectively.

2) 7-Deoxy-8-epi-loganin-BSA conjugates 20 and 25

7-Deoxy-8-epi-loganin (2b) was prepared, as shown below, from boschnaloside (21)¹⁴⁾ in which the methyl group at C-8 position was established to be α -configuration. Oxidation of its acetate (22) with sodium chlorite and sodium dihydrogen phosphate in the presence of 2-methyl-2-butene afforded 7-deoxy-8-epi-loganic acid tetraacetate (23).¹⁵⁾ Methylation of 23 with diazomethane followed by Zemplen deacetylation afforded 7-deoxy-8-epi-loganin (2b).

The two types of 7-deoxy-8-epi-loganin hapten were prepared according to the same procedure as in the above haptens, <u>15</u> and <u>19</u>. 11-GABA amide (<u>24</u>), the hapten of type I, was synthesized via condensation of <u>23</u> with Me-GABA followed by hydrolysis. 6'-GABA amide (<u>26</u>), the hapten of type II, was prepared by the regioselective oxidation of <u>2b</u> with platinum oxide and condensation of the resulting 6'-acid (<u>27</u>) with Me-GABA followed by hydrolysis. Condensation of two haptens (<u>24</u>, <u>26</u>) with BSA afforded two conjugates <u>20</u> and <u>25</u>, respectively.

3) 10-Deoxygeniposide-BSA conjugates 28 and 31

In the previous works, 10-deoxygeniposide (<u>3b</u>) and its acetate (<u>29</u>) were derived from geniposide (<u>37</u>), mussaenoside (<u>42</u>)¹⁶) and their acetates (<u>38</u>, <u>43</u>).¹⁷⁻¹⁹) However, hydrogenolysis of geniposide pentaacetate (<u>38</u>) gave a mixture of <u>29</u>, <u>12</u> and 7-deoxy-10-hydroxyloganin tetraacetate (<u>44</u>), besides <u>38</u>, which were separable in small scale by chromatography on silica gel and subsequently on silica gel impregnated with AgNO₃. On the other hand, dehydration of mussaenoside tetraacetate (<u>43</u>) gave <u>29</u> containing about 10 % of the 8,10-double bond isomer (<u>45</u>), which were unseparable. However, careful hydrogenolysis of free glucoside <u>37</u> in the presence of acid was found to give <u>3b</u> in high yield which was purified as its acetate (<u>29</u>) by chromatography on silica gel. Glucosides <u>3a</u> and <u>3b</u> were obtained through hydrolysis of <u>29</u> with 0.4N NaOH and Zemplen deacetylation, respectively. Condensation of <u>3a</u> with Me-GABA followed ·by hydrolysis gave 10-deoxygeniposide 11-GABA amide (<u>30</u>), the hapten of type I. Unfortunately, all attempts to oxidize <u>3b</u> with platinum oxide to 6'-acid (<u>32</u>) were unsuccessful. Thus we adopted an alternative route leading to 6'-acid (<u>32</u>) or its derivative. <u>3b</u> was subjected to tritylation with trityl chloride in pyridine but the reaction proceeded very slowly and the yield of the tritylated product also was very low. However, tritylation of <u>3b</u> with trityl pridininium tetrafluoroborate²⁰ in stead of TrCl resulted in the rapid progress of the reaction and the high yield of the tritylated product. The tritylation of <u>3b</u> followed by acetylation gave 2',3',4'-triacetyl-6'-trityl-10-deoxygeniposide (<u>33</u>) in 99.6 % yield. Detritylation of <u>33</u> with 80% acetic acid and subsequent oxidation with Jones reagent afforded 6'-acid triacetate (<u>34</u>). 6'-GABA amide (<u>35</u>), the hapten of type II, was derived from <u>34</u> via 6'-GABA amide methyl ester (<u>36</u>) according to the same procedure as mentioned above. Two haptens <u>30</u> and <u>35</u> were coupled with BSA to give immunogenic conjugates <u>28</u> and <u>31</u>, respectively.

On the basis of the colorimetrical analysis for sugar moieties in glucosides by a phenol-sulfuric acid reagent,⁽²¹⁾ the molar ratios of glucoside to BSA in six above-mentioned conjugates were as follows: 4.5 in <u>13</u>, 8.5 in <u>16</u>, 15.7 in <u>20</u>, 15.2 in <u>25</u>, 11.3 in <u>28</u> and 9.6 in <u>31</u>.

Preparation and properties of antibodies by the six immunogenic conjugates described in the present paper as well as their application to the determination of three key intermediates, $\underline{1}$, $\underline{2}$ and 3 in various plants will be given elsewhere.

ACKNOWLEDGEMENT

We are grateful to Prof. F. Murai and Dr. M. Tagawa, Aichi Medical University, for their kind gift of boschnaloside and also to Prof. T. Shingu, Kobe-Gakuin University, for the measurement of 400 MHz NMR spectra.

EXPERIMENTAL

Melting points were measured with a Yanagimoto micromelting point apparatus and are uncorrected. Infrared (IR) spectra were recorded on a Shimadzu IR-435 infrared spectrometer and ultraviolet (UV) spectra on a Hitachi model 200-20 spectrometer. Optical rotations were obtained at 25 $^{\circ}$ C on JASCO DIP-181 and DIP-300 degital polarimeters. Proton and carbon nuclear magnetic resonance (¹H-NMR and ¹³C-NMR) spectra were determined as solution in the indicated solvents with JEOL FX-200 which was used unless stated otherwise, Bruker AC-300 and AM-400 nuclear magnetic resonance spectorometers.

Merck silica gel GF_{254} or Merck 0.25 mm silica gel 60 F_{254} plates were used for thin layer chromatography (TLC) and Merck silica gel 60 PF_{254} for preparative thin layer chromatography (PLC). Column chromatography was carried out on M. Nagel silica gel 60 (70 - 200 mesh) or Wako

activated charcoal. Spots on TLC were visualized under UV light irradiation, by expodure to iodine vapor or by spraying with anisaldehyde - sulfuric acid followed by heating and bands on PLC under UV light irradiation. All nonaqueous reactions were carried out under nitrogen or argon. Anhydrous magnesium sulfate was used for drying organic solvent extracts.

6,6-Bis(ethylthio)-7-deoxyloganin tetraacetate (<u>41</u>)

Boron trifluoride etherate (2.0 ml) was added to an ice-cooled solution of <u>40</u> (2.317 g) and ethanedithiol (5.0 ml) in anhydrous tetrahydrofuran (THF) (7.0 ml). The mixture was stirred for 1 hr under ice-cooling and for an additional 2 days at room temperature. The reaction mixture was poured on iced water and extracted three times with $CHCl_3$. The extracts were washed successively with sat. aq. NaHCO₃ and water, and dried. Removal of the solvent gave a residue (8.235 g) which was chromatographed on silica gel with ether as eluant. The fractions showing Rf 0.48 on TLC (diethyl ether) were combined and concentrated <u>in vacuo</u>. The crystalline residue (2.617 g) was recrystallized from n-PrOH to yield colorless needles of <u>41</u>.

mp 135 - 136 °C. UV λ_{max} (MeOH) nm (log ε): 230(3.92). IR ν_{max} (KBr) cm⁻¹: 1758, 1717, 1636. ¹H-NMR (CDCl₃) δ : 7.28(1H, d, J=1.2 Hz, 3-H), 5.27(1H, d, J=5.1 Hz,1-H), 5.23(1H, t, J=9.3 Hz, 3'-H), 5.09(1H, t, J=9.5 Hz, 4'-H), 5.00(1H, dd, J=9.3 and 8.1 Hz, 2'-H), 4.85 (1H, d, J=8.1 Hz, 1'-H), 4.28(1H, dd, J=12.5 and 5.4 Hz, 6'-H), 4.13(1H, dd, J=12.2 and 2.4 Hz, 6'-H), 3.74(1H, ddd, J=9.5, 5.1 and 2.4 Hz, 5'-H), 3.71(3H, s, OCH₃), 3.53(1H, br. d, J=8.3 Hz, 5-H), 3.35 - 3.15(4H, m, $-SCH_2CH_2S-$), 2.48(1H, dd, J=11.7 and 5.4 Hz,7-H), 2.09, 2.03, 2.01 and 1.97 (each, 3H, s, 4 × OCOCH₃), 1.14(3H, d, J=6.1 Hz, 10-H₃). Anal. Calcd. for C₂₇H₃₆O₁₃S₂: C, 51.26; H, 5.74; S, 10.13. Found: C, 50.97; H, 5.69; S, 10.14.

7-Deoxyloganin tetraacetate $(\underline{12})$

A mixture of $\underline{41}$ (2.000 g) and Raney Ni (W-2) (20.0 g) in absolute EtOH (30 ml) was refluxed overnight. The catalyst was filtered off and washed with EtOH. The filtrate and washings were combined and concentrated <u>in vacuo</u>. The residue (1.746 g) was acetylated with acetic anhydride and pyridine (each 15 ml) in the usual way. The crude acetylated product (1.958 g) was chromatographed on silica gel (80 g) using diethyl ether as eluant. The combined fraction showing Rf 0.35 on TLC (diethyl ether) was concentrated <u>in vacuo</u> to give a crystalline residue (1.220 g) which was recrystallized from n-PrOH to yield colorless needles of 12.

mp 117-118.5 °C. [α]_D -80.83 ° (c, 0.98, MeOH). UV λ max (MeOH) nm (log ε): 233(3.04). IR ν max (KBr) cm⁻¹: 1758, 1742(sh),1715, 1640, 1630(sh). 'H-NMR (CDCl₃) δ : 7.30(1H, d, J=1.2 Hz, 3-H), 5.23(1H, t, J=9.3 Hz, 3'-H), 5.16(1H, d, J=3.4 Hz, 1-H), 5.10(1H, t, J=9.3 Hz, 4'-H), 4.99(1H, dd, J=9.0 and 8.1 Hz, 2'-H), 4.87 (1H, d, J=8.1 Hz, 1'-H), 4.31(1H, dd, J=12.5 and 4.6 Hz, 6'-H), 4.14(1H, dd, J=12.5 and 2.4 Hz, 6'-H), 3.74(1H, ddd, J=9.8, 4.6 and 2.4 Hz, 5'-H), 3.70(3H, s, OCH_{\exists}), 2.87 (1H, br. q, J=6.5 Hz, 5-H), 2.17(1H, m, 6-H), 2.09, 2.03, 2.00 and 1.93(each 3H, s, 4 × $OCOCH_{\exists}$), 1.90 - 1.76(2H, m, 8- and 9-H), 1.45(1H, d, J=15.0, 7.3 and 5.6 Hz, 6-H), 1.18(1H, m, 7-H), 1.06(3H, d, J=5.9 Hz, 10-H₃). ¹³C-NMR (CDCl₃) δ : 170.54, 170.15, 169.38 and 169.11(each s, 4 × COCH₃), 167.45(s, 11-C), 149.45(d, 3-C), 113.23(s, 4-C), 96.04(d, 1-C), 95.38(d, 1'-C), 72.56(d, 3'-C), 72.12(d, 5'-C), 70.69(d, 2'-C), 68.35(d, 4'-C), 61.81(t, 6'-C), 51.11(q, 11-OCH₃), 48.26(d, 9-C), 34.96(d, 5-C), 33.01(t, 6-C), 32.64(d, 8-C), 31.16(t, 7-C), 20.70, 20.58 and 20.21(each q, 4 × COCH₃), 19.63(q, 10-C). Anal. Calcd. for C_{z5}H₃₄O₁₃: C, 55.35; H, 6.32. Found: C, 55.34; H, 6.33.

7-Deoxyloganin (<u>1b</u>)

To a solution of <u>12</u> (228.3 mg) in anhydrous MeOH (4 ml) was added 0.26N NaOMe (0.2 ml) and the mixture was stirred for 2 hr at room temperature. After neutralization with Amberlite IR-120 resin (H⁺ form), the resin was filtered off and washed with MeOH. The filtrate and washings were combined and concentrated <u>in vacuo</u>. The residue (154.3 mg) was purified by PLC (MeOH - CHCl₃ 2 : 8) and the band around Rf 0.35 afforded a crystalline residue (139.9 mg) which was recrystallized from n-PrOH to yield colorless needles of <u>1b</u>.

mp 156 - 158 °C. [α]_D -92.24° (c, 0.98, MeOH) [lit. mp 157 - 157.5 °C from THF - hexane.

 $[\alpha]_{D} -90^{\circ}$ (c, 0.295, EtOH)].²²⁾ UV λ_{max} (MeOH) nm(log ε): 236(4.06),IR ν_{max} (KBr) cm⁻¹: 3510, 3350, 1685, 1645. 'H-NMR (CD₃OD) δ : 7.41(1H, d, J=1.2 Hz, 3-H), 5.21(1H, d, J=5.6 Hz, 1-H), 4.66(1H, d, J=7.8 Hz, 1'-H), 3.89(1H, dd, J=12.0 and 1.7 Hz, 6'-Ha), 3.69(1H, s, 11-OCH₃), 3.67(1H, dd, J=12.0 and 5.6 Hz, 6'-Hb), 3.38(1H, dd, J=9.0 and 7.8 Hz, 3'-H), 3.36 - 3.25(2H, m, 5'- and 4'-H), 3.19(1H, dd, J=8.8 and 7.8 Hz, 2'-H), 2.88(1H, br q, J=7.3 Hz, 5-H), 2.18(1H, dtdd, J=12.7, 7.6, 3.4 and 1.2 Hz, 6-Ha), 1.96(1H, br tq J=7.3 and 6.6 Hz, 8-H), 1.87(1H, dtd, J=11.5, 7.3 and 3.7 Hz, 7-H), 1.73(1-H, ddd, J=8.1, 6.1 and 5.6 Hz, 9-H), 1.38(1H, ddt, J=12.7, 9.3 and 7.3 Hz, 6-Hb), 1.18(1H, ddt, J=11.5, 9.3 and 3.7 Hz, 7-H), 1.08(3H, d, J=6.6 Hz, 10-H₃). ¹³C-NMR (CD₃OD, 75.5MHz) δ : 169.75(s, 11-C), 152.70(d, 3-C), 112.97(s, 4-C), 100.25(d, 1'-C), 97.90(d, 1-C), 78.40(d, 3'-C), 78.10(d, 5'-C), 74.81(d, 2'-C), 71.65(d, 4'-C), 62.82(t, 6'-C), 51.66(q, 11-OCH₃), 36.59(d, 5-C), 35.29(t, 6-C), 34.20(d, 8-C), 33.37(t, 7-C), 20.83(q, 10-C). Anal. Calcd. for C_{1.7}H₂₆O₉: C, 54.54; H, 7.00. Found: C, 54.51; H, 7.07.

7-Deoxyloganic acid (<u>1a</u>)

The compound <u>1b</u> (500 mg) was hydrolyzed with 0.5N aq. NaOH (7.0 ml) for 5 hr under stirring at room temperature. The mixture was neutralized with Amberlite IR-120 resin (H^+ form) and the

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resin was filtered off, then washed with water. The combined filtrate and washings were concentrated in vacuo to yield <u>la</u> as a colorless syrup.

 $[\alpha]_{\circ}$ -89.38 ° (c 1.07, MeOH). 'H-NMR (CD₂OD) δ : 7.42(1H, d, J=1.2 Hz, 3-H), 5.21(1H, d, J=5.9 Hz, 1-H), 4.68(1H, d, J=7.8 Hz, 1'-H), 3.90(1H, dd, J=11.7 and 1.5 Hz, 6'-H), 3.67(1H, t, J=8.8 Hz, 3'-H), 3.20(1H, dd, 8.8 and 7.8 Hz, 2'-H), 2.86(1H, tdd, J=8.0, 6.5 and 1.2 Hz, 5-H), 2.30 - 2.10(1H, m, 6-H), 2.07 - 1.85(2H, m, 7- and 8-H), 1.73(1H, ddd, J=8.3, 6.4 and 5.9 Hz, 9-H), 1.53 - 1.10(2H, m, 6- and 7-H), 1.09(3H, d, J=6.4 Hz 10-H₃). FAB-MS m/z: 361 [M+H]⁺

7-Deoxyloganin 11-GABA amide methyl ester (14)

A solution of <u>la</u> (245.1 mg), 1-hydroxybenzotriazole (HOBT) (183.8 mg) and EDCI (156.6 mg) in anhydrous THF (2.0 ml) was stirred for 1 hr under ice- cooling. To the mixture was added a solution of methyl γ -aminobutylate hydrochloride (Me-GABA·HCl) (114.1 mg) and dimethylaminopyridine (DMAP) (166.9 mg) in anhydrous acetonitrile (1.0 ml) and the mixture was stirred for 1 hr at the same temperature. Stirring was continued for an additional 2 days at room temperature. Evaporation <u>in vacuo</u> of the solvent gave a residue which was purified by PLC (MeOH - CHCl₃ - HCOOH 30 : 65 : 5). The band around Rf 0.50 afforded <u>14</u> (298 mg) as a colorless syrup.

[α]_P -87.00° (c, 1.01, MeOH). 1R ν_{max} (KBr) cm⁻¹: 3365.8, 2953.3, 2872.2, 1723.7, 1654.1, 1602.2, 1542.0. 'H-NMR (CD₃OD) δ : 7.67(1H, br. t, J=5.6 Hz, NH), 7.05(1H, d, J=1.2 Hz, 3-H), 5.20(1H, d, J=5.1 Hz, 1-H), 4.66(1H, d, J=7.6 Hz, 1'-H), 3.89(1H, brd, J=12.7 Hz, 6'-H), 3.71 - 3.63(1H, m, 5'-H), 3.66(3H, s, OCH₃), 3.60(1H, dd, J=12.7 and 5.6 Hz, 6'-H), 3.39(1H, dd, J=9.0 and 8.8 Hz, 4'-H), 3.33 - 3.23(3H, m, 4"-H₂ and 3'-H), 3.21(1H, dd, J=8.6 and 7.8 Hz, 2'-H), 2.97(1H, br q, J=7.1 Hz, 5-H), 2.36(2H, t, J=7.3 Hz, 2"-H), 2.17 - 1.70(4H, m, 9-, 8-, 7- and 6-H), 1.81(2H, q, J=7.3 Hz, 3"-H), 1.34 - 1.04(2H, m, 7- and 6-H), 1.09(3H, d, J=6.6 Hz, 10-CH₃). ¹³C-NMR (CD₃OD, 75.5MHz) δ : 175.48(s, 1"-C), 170.71(s, 11-C), 146.88(d, 3-C), 116.60(s, 4-C), 100.05(d, 1'-C), 97.13(d, 1-C), 78.36(d, 3'-C), 78.04(d, 5'-C), 74.84(d, 2'-C), 71.65(d, 4'-C), 62.80(t, 6'-C), 52.14(q, 11-0CH₃), 39.78(T, 6'-C), 36.33(d, 8-C), 34.87(d, 5-C), 34.05(t, 7-C), 32.53(t, 6-C), 32.20(t, 2"-C), 25.88(t, 3"-C), 20.70(q, 10-C). FAB-MS m/z: 460 [M+H]⁺

7-Deoxyloganin 11-GABA amide (15)

A solution of <u>14</u> (217.2 mg) in MeOH (4.8 ml) was added to 0.4N aq. NaOH solution (4.8 ml) and the mixture stirred for 2 hr at room temperature. After neutralization with Amberlite IR-120 resin (H⁺ form), the resin was filtered off and washed with MeOH. The filtrate and washings were combined and concentrated in <u>vacuo</u>. The residue was purified by PLC (MeOH - $CHCl_3 3:7$) and

the product obtained from the band around Rf 0.2 was chromatographed on activated charcoal (400 mg). Elution with MeOH and afforded a crystalline residue which was recrystallized from n-PrOH to yield colorless needles of $\underline{15}$.

mp 216-218 °C. UV λ_{mex} (MeOH) nm (log ε): 228 (4.01). IR ν_{mex} (KBr) cm⁻¹: 3501, 3425, 3250, 2963, 2915, 1711, 1649, 1613, 1528. 'H-NMR (CD₃OD) δ : 7.05(1H, d, J=1.2 Hz, 3-H), 5.21(1H, d, J=5.1 Hz, 1-H), 4.66(1H, d, J=7.6 Hz, 1'-H), 3.89(1H, br. d, J=12.0 Hz, 6'-Ha), 3.68(1H,dd, J=12.0 and 5.4 Hz, 6'-Hb), 3.46 - 3.15(5H, m, 2'-, 3'-, 4'-H and 4"-Hz), 2.99(1H, br q, J=7.6 Hz, 5-H), 2.33(2H, t, J=7.3 Hz, 2"-H), 2.15 - 2.03(1H, m, 6-Ha), 2.02 - 1.70(3H, m, 8-, 7- and 9-H), 1.81(2H, quint, J=7.3 Hz, 3"-Hz), 1.45 - 1.13(2H, m, 6- and 7-H), 1.09(3H, d, J=6.4 Hz, 10-H<u>3</u>). FAB-MS m/z: 446 [M+H]⁺

7-Deoxyloganin 11-GABA-BSA conjugate (13)

BSA (223.2 mg) was dissolved in water which was adjusted by the addition of 0.02N aq. NaOH. Upon addition of <u>15</u> (60.0 mg), lowering pH to 5.5 was observed, and EDCI (28.5 mg) was added subsequently. The mixture was stirred overnight at 25 °C and dialyzed at 4 °C against 10 × 5 l H₂O for 5 days. The inner solution was lyophilized to give <u>13</u> (230.1 mg). Coupling ratio was determined from the colorimetrical data by a phenol - sulfuric acid reagent for the sugar part of deoxyloganin in the conjugate <u>13</u> and the glucoside/protein ratio was calculated as 4.5.

6'-Carboxy-7-deoxyloganin (17)

The foregoing compound (<u>1b</u>) (258 mg) and Na₂CO₃ (430 mg) were added to the suspension of prehydrogenated PtO₂ (258 mg) in H₂O (2.5 ml) and the mixture was stirred for 7 days under an atmosphere of oxygen at room temperature. The reaction mixture was neutralized with Amberlite IR-120 resin (H⁺ form) and the resin was filtered off, then washed with H₂O. The combined filtrate and washings were concentrated <u>in vacuo</u> to afford the residue (246.9 mg) which was subjected to PLC (MeOH - CHCl₃ - HCOOH 20 : 80 : 0.5). The residue obtained from the band around Rf 0.25 was chromatographed on activated charcoal (720 g). The MeOH eluate yielded <u>17</u> (129 mg) as a syrup.

IR ν_{mex} (KBr) cm⁻¹: 3431, 2954, 1735, 1696, 1640, 1442. ¹H-NMR (DMSO-de, 300 MHz) δ : 7.38(1H, d, J=1.13 Hz, 3-H), 5.19 - 5.11(1H, br s, COO<u>H</u>), 4.99(1H, d, J=6.1 Hz, 1-H), 4.60(1H, d, J=7.8 Hz, 1'-H), 3.68(1H, d, J=9.6 Hz, 5'-H), 3.35(1H, dd, J=9.5 and 9.1 Hz, 4'-H), 3.20(1H, t, J=8.9 Hz, 3'-H), 3.02(1H, dd, J=8.5 and 8.1 Hz, 2'-H), 2.79(1H, q, J=7.6 Hz, 5-H), 2.10(1H, ddd, J=12.6, 8.0 and 3.5 Hz, 6-H), 1.85(1H, sextet, J=6.7 Hz, 8-H), 1.76(1H, qd, J=7.6 and 3.5 Hz, 7-H), 1.66(1H, dt, J=8.1 and 6.1 Hz, 9-H), 1.26(1H, ddt, J=13.0, 8.8 and 7.8 Hz, 6-H), 1.20 - 1.15(1H, m, 7-H), 1.02(3H, d, J=6.5 Hz, 10-H). ^{1.3}C-NMR (DMSO-de, 75.5 MHz) δ : 169.95(s, 6'-C), 166.76(s, 11-C), 150.81(d, 3-C), 110.83(s, 10-H).

4-C), 99.29(d, 1-C), 96.46(d, 1'-C), 75.76(d, 3'-C), 75.50(d, 5'-C), 72.61(d, 2'-C), 71.17(d, 4'-C), 50.82(q, 11-OCH₃), 47.04(d, 9-C), 34.59(d, 5-C), 33.45(t, 6-C), 32.26(d, 8-C), 37.71(t, 7-C), 20.20(q, 10-C). FAB-MS m/z: 389 [M+H]⁺

7-Deoxyloganin 6'-GABA amide methyl ester (18)

A solution of <u>17</u> (94.6 mg), HOBT (44.9 mg) and EDCI (51.4 mg) in anhydrous acetonitrile and anhydrous THF (1:1) (2.0 ml) was stirred for 1hr in a ice bath. To the mixture was added a solution of Me-GABA·HCl (44.9 mg) and DMAP (72.5 mg) in anhydrous acetonitrile (1.0 ml) and the mixture was stirred for 1 hr at the same temperature. After stirring the solution for an additional 2 days at room temperature. Evaporation of the solvent <u>in vacuo</u> gave a residue which was purified by PLC (MeOH - CHCl₃ - HCOOH 20 : 80 : 0.5). The band around Rf 0.60 afforded <u>18</u> (91.7 mg) as a colorless syrup.

IR ν_{max} (KBr) cm⁻¹: 3370, 2954, 1728, 1709, 1663, 1636, 1546. 'H-NMR (CD₃OD) δ : 7.40(1H, d, J=1.0 Hz, 3-H), 5.14(1H, d, J=5.6 Hz, 1-H), 4.73(1H, d, J=7.8 Hz, 1'-H), 3.74(1H, d, J=9.5 Hz, 5'-H), 3.69(3H, s, 11- OCH₃), 3.65(3H, s, 4"-OCH₃), 3.53(1H, dd, J=9.3 and 8.5 Hz, 4'-H), 3.42(1H, dd, J= 8.8 and 8.5 Hz, 3'-H), 3.35 - 3.20(3H, m, 2'- and 4"-H₂), 2.89(1H, q, J=7.6 Hz, 5-H), 2.37(3H, t, J=7.3 Hz, 2"-H), 2.19(1H, br. dtd, J=12.2, 7.6 and 3.4 Hz, 6-H), 1.96(1H,br.tq, J=7.3 and 6.4 Hz, 8-H), 1.82(3H, quint. J=7.3 Hz, 3"-H), 1.76(1H, ddd, J=8.3, 6.11 and 5.9 Hz, 9-H), 1.37(1H, br.dtd, J=12.5, 7.6 and 3.4 Hz, 7-H), 1.19(1H, ddt, J=12.9, 7.6 and 3.4 Hz, 7-H), 1.08(3H, d, J=6.4 Hz, 10-H₃). ¹³C-NMR (CD₃OD) δ : 175.36(s, 4"-C), 171.44(s, 6'-C), 169.62(s, 11-C), 152.49(d, 3-C), 113.09(s, 4-C), 100.56(d, 1'-C), 98.18(d, 1-C), 77.55(d, 3'-C), 76.94(d, 5'-C), 74.26(d, 2"-C), 73.12(d, 4'-C), 52.13(q, 1"-OCH₃), 51.69(q, 11-OCH₃), 49.21(d, 9-C), 39.45(t, 4"-C), 36.63(d, 8-C), 35.22(d, 5-C), 34.13(t, 7-C), 33.40(t, 6-C), 32.01(t, 2"-C), 25.59(t, 3"-C), 20.82(q, 10-C). Anal. Calcd. for $C_{22}H_{33}O_{11}N \cdot 1/2H_{2}O$: C, 53.22; H, 6.90; N, 2.82. Found: C,53.10; H, 6.87; N, 2.83. FAB-MS m/z: 488 [M+H]⁺.

7-Deoxyloganin 6'-GABA amide (19)

Compound <u>18</u> (159.2 mg) was hydrolyzed in MeOH (4.0 ml) with 0.4N aq. NaOH (4.0 ml) for 1 hr by stirring at room temperature. The mixture was worked up as described above for the preparation of <u>15</u>. The crude products were chromatographed on activated charcoal (560 mg). Elution with MeOH afforded <u>19</u> (123 mg) as a white amorphous foam.

IR ν_{max} (KBr) cm⁻¹: 3450, 3410, 2920, 2850, 1625, 1573, 1528. 'H-NMR (CD₂OD) δ : 7.40(1H, d, J=1.2 Hz, 3'-H), 5.14(1H, d, J=5.9 Hz, 1-H), 4.73(1H, d, J=7.8 Hz, 1'-H), 3.74(1H, d, J=9.5 Hz, 5'-H), 3.69(3H, s, 11-OCH₃), 3.54(1H, dd, J=9.8 and 8.6 Hz, 4'-H), 3.42(1H, t, J=8.8 Hz, 3'-H), 3.36 - 3.18(3H, J=0.2 Hz, S)

m, 2'- and 4"- H), 2.89(1H, br. q, J=7.8 Hz, 5-H), 2.34(2H, t, J=7.3 Hz, 2"-H), 2.28 - 2.10(1H, m, 6-Ha), 2.05 - 1.70(3H, m, 8-, 7- and 9-H), 1.81(2H, quintet, 3"-H), 1.46 -1.12(2H, m, 6- and 7-H), 1.08(3H, s, 10-H₃). FAB-MS m/z: 474 [M+H]⁺.

7-Deoxyloganin 6'-GABA-BSA conjugate (16)

BSA (222.7 mg) was dissolved in water which was adjusted to pH 8.5 by the addition of 0.02N aq. NaOH. <u>19</u> (63.9 mg) and EDCI (28.5 mg) were added subsequently. The mixture was stirred overnight at 25 $^{\circ}$ C and dialyzed at 4 $^{\circ}$ C against 10 \times 5 l changes of H₂O for 5 days. The inner solution was lyophilized to give the conjugate <u>16</u> (236.3 mg). The conjugate <u>16</u> was determined from the colorimetrical method by phenol - sulfuric acid for the sugar moiety to contain 8.5 mole of <u>19</u> / mole of BSA.

7-Deoxy-8-epi-loganic acid tetraacetate (23)

The title compound was prepared according to the procedure of S. P. Bal et al.¹⁵⁾ To a solution of boschnaloside tetraacetate (22) (1.000 g) in t-BuOH (41.1 ml) was added 2-methyl-2-butene (9.9 ml) and subsequently dropwise a solution of sodium chlorite (1.641 g) and sodium dihydrogen phosphate (2.133 g) in water (16.5 ml) over a 15 min period by stirring at room temperature. Stirring was continued for 40 hr at the same conditions. The reaction mixture was concentrated <u>in vacuo</u> and the residue was partitioned between water and CHCl₃. The aqueous layer was further extracted with CHCl₃. The extracts were washed with sat. brine, dried and concentrated <u>in vacuo</u>. The residue (1.020 g) was chromatographed on silica gel (30 g) using CHCl₃-MeOH with the increasing ratio of MeOH. The 5% MeOH-CHCl₃ eluates were combined and concentrated <u>in vacuo</u> to give a crystalline residue (912.4 mg) which was recrystallized from aqueous EtOH to yield colorless needles of <u>23</u>.

mp 188 – 189.5 °C. [α]_D –98.89 ° (c, 1.00, MeOH). IR ν_{mex} (KBr) cm⁻¹: 3450, 2950, 1758, 1720, 1646. ¹H-NMR (CDCl_B) δ : 7.45(1H, br s, 3-H), 5.21(1H, br t, J=9.3 Hz, 3'-H), 5.10(1H, d, J=4.0 Hz, 1-H), 5.09(1H, br t, J=9.4 Hz, 4'-H), 5.06(1H, dd, J=9.0 and 8.1 Hz, 2'-H), 4.85(1H, d, J=8.1 Hz, 1'-H), 4.27(1H, dd, J=12.2 and 5.1 Hz, 6'-H), 4.14(1H, dd, J=12.2 and 2.4 Hz, 6'-H), 3.77(1H, ddd, J=9.8, 5.1 and 2.4 Hz, 5'-H), 2.90(1H, br q, J=7.1 Hz, 5-H), 2.40 – 2.10(2H, m, 8- and 9-H), 2.09, 2.04, 2.03 and 2.01(each 3H, s, 4 × OCOCH_B), 1.90 – 1.70(1H, m, 7-H), 1.55(1H, dtd, J=11.5, 6.6 and 4.9Hz, 6-H), 1.29(1H, dq, J=12.5 and 8.1 Hz, 7-H), 1.02(3H, d, J=6.6 Hz, 10-H_B). ^{1B}C-NMR (CDCl_B) δ :172.51(s, 11-C), 170.66, 170.35, 169.38, 169.18(each s, 4 × COOCH_B), 152.86(d, 3-C), 111.46(s, 4-C), 99.73(d, 1'-C), 99.49(d, 1-C), 73.12(d, 3'-C), 72.29(d, 5'-C), 71.56(d, 2'-C), 68.26(d, 4'-C), 62.13(t, 6'-C), 42.93(d, 9-C), 36.20(d, 5-C), 33.33(d, 8-C), 32.18(t, 6-C), 31.16(t, 7-C), 20.72(q, 2 × CO<u>C</u>H_B),

20.60(q, 2 × CO<u>C</u>H₃), 16.37(q, 10-C). Anal. Calcd. for C_{≥4}H_{3≥}O₁₃: C, 54.54; H, 6.10. Found: C, 54.67; H, 6.18.

7-Deoxy-8-epi-loganin 11-GABA amide (24)

A solution of 23 (100 mg), HOBT (51.2 mg) and EDCI (39.9 mg) in anhydrous THF (1.5 ml) was stirred for 1 hr in a ice bath. To the solution was added a solution of Me-GABA·HCl (31.7 mg) and DMAP (46.4 mg) in anhydrous acetonitrile (1.0 ml) and the mixture was stirred for 1 hr at the same temperature. Stirring was continued for an additional 1 day at room temperature. The reaction mixture was diluted with water and extracted with CHCl₃. The extracts were washed, successively, with 1N HCl, sat. aq. NaHCO₃ and water, and dried. Removal of the solvent gave a residue which was purified by PLC (CHCl₃ - MeOH 95 : 5). The band around Rf 0.60 afforded a crystalline residue (112.0 mg) which was recrysallized from i-PrOH to give tetraacetyl 7-deoxy-8-epi-loganin 11-GABA amide methyl ester as colorless needles.

mp 139 - 140 °C. UV λ_{max} (MeOH) nm (log ε): 227.7 (4.02). IR ν_{max} (KBr) cm⁻¹: 3400, 2950, 1750, 1659, 1622. 'H-NMR (CD₃OD, 300 MHz) δ : 7.63(1H, br t, J=5.5 Hz, N<u>H</u>), 7.10(1H, d, J=1.0 Hz, 3-H), 5.31(1H, d, J=2.9 Hz, 1-H), 5.28(1H, d, J=9.5 Hz, 4'-H), 5.04(1H, d, J=8.0 Hz, 2'-H), 5.01(1H, d, J=9.5 Hz, 1'-H), 4.87(1H, dd, J=9.7 and 8.1 Hz, 4'-H), 4.29(1H, dd, J=12.4 and 4.3 Hz, 6'-H), 4.17(1H, dd, J= 12.4 and 2.4 Hz, 6'-H), 3.92(1H, ddd, J=10.0, 4.3 and 2.4 Hz, 5'-H), 3.66(3H, s, OCH₃), 3.21(2H, quint, J=6.8, 4"-H), 2.94(1H, td, J=7.7 and 4.1 Hz, 5-H), 2.36(2H, t, J=7.4 Hz, 2"-H), 2.39 - 2.22(2H, m, 8- and 9-H), 2.06, 2.01, 1.96 and 1.92(each 3H, s, OCOCH₃), 1.82(2H, quint, J=7.1 Hz, 2"-H), 1.99 - 1.73(2H, m, 6- and 7-H), 1.51(1H, ddd, J=11.7, 7.6 and 4.1 Hz, 6-H), 1.28(1H, dq, J=12.3 and 8.1, 7-H), 1.03(3H, d, J=6.8 Hz, 10-H). ¹³C-NMR (CD₃OD) δ : 173.90(s, 1"-C), 170.54 and 170.06(each s, OCOCH₃), 169.40(s, 2 × OCOCH₃), 166.70(s, 11-C), 146.27(d, 3-C), 115.47(s, 4-C), 95.84(d, 1'-C), 94.36(d, 1-C), 72.63(d, 3'-C), 72.12(d, 5'-C), 70.79(d, 2'-C), 68.47(d, 4'-C), 61.83(t, 6'-C), 51.74(q, 1"-OCH₃), 43.01(d, 9-C), 39.07(t, 4"-C), 35.15(d, 5-C), 33.13(t, 6-C), 31.89(d, 8-C), 31.65(t, 2"-C), 30.94(t, 7-C), 24.67(t, 3"-C), 20.70(q, OCOCH₃), 20.57(q, 2 × OCOCH₃), 20.41(q, OCOCH₃), 15.96(q, 10-C). Anal. Calcd. for $C_{29}H_{4,1}O_{1,4}N$: C, 55.50; H, 6.58; N, 2.23. Found: C, 55.53; H, 6.51; N, 2.10.

The above amide methyl ester (236.8 mg) was hydrolyzed in MeOH (4.7 ml) with 0.4N aq. NaOH (4.7 ml) for 1.5 hr by stirring at room temperature. The mixture was worked up as described above for the preparation of <u>15</u>. The crude products were purified by PLC (Rf 0.10, CHCl3 - MeOH - HCOOH 20 : 80 : 5) followed by column chromatography on activated charcoal (800 mg) with MeOH to afford <u>24</u> (179.4 mg) as a white amorphous foam.

IR ν_{mex} (KBr) cm⁻¹: 3360, 2920, 2860, 1710, 1655, 1595. ¹H-NMR (CD₃OD) δ : 7.08(1H, d, J=1.0 Hz, 3-H), 5.42(1H, d, J=3.9 Hz, 1-H), 4.67(1H, d, J=7.6 Hz, 3-H), 3.90(1H, dd, J=11.7 and 1.7 Hz, 6'-H),

3.71(1H,dd, J=11.7 and 5.4 Hz, 6'-H), 3.25(2H, t, J=6.6 Hz, 4"-H₂), 3.20(1H, dd, J=9.0 and 7.8 Hz, 2'-H), 2.97(1H, br td like Hz, 5-H), 2.33(2H, t, J=7.3 Hz, 2"-H₂), 2.16 - 2.35(2H, m, 8-H and 9-H), 1.99(1H, dq, J=11.7 and 8.1 Hz, 6-H), 1.90 - 1.61(1H, m, 7-H), 1.81(2H, quint, J=7.1Hz, 3"-H₂), 1.61 - 1.23(2H, m, 6-H and 7-H), 1.08(3H, d, J=6.6 Hz, 10-H₃). Anal. Calcd. for $C_{20}H_{31}O_{10}N$: C, 53.93; H, 7.01; N, 3.14. Found: C, 53.63; H, 6.98; N, 3.16. FAB-MS m/z: 446 [M+H]⁺

7-Deoxy-8-epi-loganin 11-GABA-BSA conjugate (20)

BSA (443.0 mg) and $\underline{24}$ (179.4 mg) were condensed using EDCI (66.2 mg) according to the same procedure as described for <u>13</u>. Dialysis followed by lyophilization provided <u>20</u> (489.9 mg). The <u>24</u> /BSA ratio of the conjugate <u>20</u> was determined by the aforementioned colorimetric method to give 15.7 mole of <u>24</u> / mole of BSA.

7-Deoxy-8-epi-loganin (2b)

Diazomethane in diethylether was added to a solution of <u>23</u> (100.2 mg) in MeOH (5 ml) in an ice bath until persistent yellow color. After leaving on an ice bath for 30 min, the mixture was concentrated <u>in vacuo</u>. The residue was purified by PLC (Rf 0.60, acetone - benzene 20 : 80) to give a crystalline residue (95.8 mg) which was recrystallized from i-PrOH to yield tetraacetyl 7-deoxy-8-epi-loganín as colorless needles.

mp 111 - 112 °C. [α]_D -111.12 ° (c, 0.71, MeOH) [lit. mp 150 - 152 °C. from AcOEt. [α]_D -128 ° (c, 0.4, MeOH)].²³ UV λ_{max} (MeOH) nm (log ε): 237.1(4.08). IR ν_{max} (KBr) cm⁻¹: 2950, 1758, 1705, 1640. ¹H-NMR (CDCl₃, 400MHz) δ : 7.35(1H, d, J=1.0 Hz, 3-H), 5.27(1H, d, J=3.5 Hz, 1-H), 5.22(1H, t, J=9.6 Hz, 3'-H), 5.10(1H, dd, J=9.8 and 9.5 Hz, 4'-H), 4.99(1H, dd, J=9.6 and 8.1 Hz, 2'-H), 4.87(1H, d, J=8.1 Hz, 1'-H), 4.27(1H, dd, J=12.4 and 4.5 Hz, 6'-H), 4.17(1H, dd, J=12.4 and 2.4 Hz, 6'-H), 3.73(1H, ddd, J=10.0, 4.4 and 2.5 Hz, 5'-H), 3.70(3H, s, COOC<u>H</u>₃), 2.87(1H,tdd, J=8.0, 5.0 and 1.0 Hz, 5-H), 2.31 (1H, td, J=8.6 and 3.6 Hz, 9-H), 2.24(1H, tdq, J=8.6, 7.2 and 7.0 Hz, 8-H), 2.09, 2.03, 2.00 and 1.94 (each 3H, s, 4 × COOC<u>H</u>₃), 1.75(1H, dtd, J=12.3, 7.3 and 4.4 Hz, 7-H), 1.60(1-H, ddt, J=12.6, 7.6 and 4.9 Hz, 6-H), 1.24(1H, dq, J=12.6 and 8.1 Hz, 7-H), 1.00(3H, d, J=7.0 Hz, 10-H₃). ¹³C-NMR (CDCl₃) δ : 170.57, 170.18, 169.38, and 169.16(each s, 4 × <u>C</u>OCH₃), 167.36(s, 11-C), 150.21(d, 3-C), 113.26(s, 4-C), 95.69(d, 1'-C), 94.70(d,1-C), 72.61(d, 3'-C), 72.08(d, 5'-C), 70.74(d, 2'-C), 68.35(d, 4'-C), 61.71(t, 6'-C), 51.11(q,O<u>C</u>H₃), 42.67(d, 9-C), 35.44(d, 5-C), 32.38(d, 8-C), 31.09(t, 7-C), 20.70(q, CO<u>C</u>H₃), 20.58(q, 2 × CO<u>C</u>H₃), 20.26(q, CO<u>C</u>H₃), 16.05(q, 10-C). Anal. Calcd. for C₂₅H₃₄O₁₂:C, 55.35; H, 6.32. Found: C, 55.35; H, 6.34.

To a solution of the above acetate (450 mg) in anhydrous MeOH (9 ml) was added 0.26N NaOMe (0.2 ml) and the mixture was stirred for 4 hr at room temperature. Work up in the same manner as

described for <u>1b</u> gave <u>2b</u> (301.0 mg) as colorless needles which was recrystallized from n-PrOH. mp 163-165 °C. [α]_D -115.13 ° (c, 0.72, MeOH). UV λ mex (MeOH) nm (log ε): 237.1(4.08). IR ν mex (KBr) cm⁻¹: 3750, 2960, 2900, 1682, 1642. ¹H-NMR (CD₃OD) δ : 7.42(1H, d, J=1.0 Hz, 3-H), 5.46(1H, d, J=4.6 Hz, 1-H), 4.68(1H, d, J=7.6 Hz, 1'-H), 3.90(1H, dd, J=11.7 and 1.7 Hz, 6'-Ha), 3.68(3H, s, 11-OCH₃), 3.64(1H, dd, J=11.72 and 5.62 Hz, 6'-Hb), 3.37(1H, t, J=8.6 Hz, 4'-H), 3.27(1H, dd, J=8.1 and 7.8 Hz, 3'-H), 3.19(1H, dd, J=9.0 and 7.8 Hz, 2'-H), 2.92(1H, td, J=7.8 and 6.1 Hz, 5-H), 2.40 - 2.15(2H, m, 9- and 8-H), 2.06(1H, dq, J=12.7 and 7.8 Hz, 6-Ha), 1.86 - 1.70(1H, m, 7-Ha), 1.63 - 1.46 (1H, m, 6-Hb), 1.39(1H, dq, J=12.2 and 7.8 Hz, 7-Hb), 1.08(3H, d, J=6.6 Hz, 10-CH₃). ¹³C-NMR (CD₃OD) δ : 169.59(s, 11-C), 152.81(d, 3-C), 113.38(s, 4-C), 99.81(d, 1'-C), 96.25(d, 1-C), 78.42(d, 3'-C), 78.06 (d, 5'-C), 74.85(d, 2'-C), 71.81(d, 4'-C), 63.00(t, 6'-C), 51.64(q, 11-0CH₃), 44.44(d, 9-C), 37.58(d, 8-C), 34.59(d, 5-C), 33.28(t, 7-C), 32.38(t, 6-C), 16.76(q, 10-C). Anal. Calcd. for C_{1.7}H₂₆O₆:C, 54.54; H, 7.00. Found: C, 54.70; H, 7.07.

6'-Carboxy-7-deoxy-8-epi-loganin (27)

The compound <u>2b</u> (240 mg) and Na_2CO_3 (524.8 mg) were added to the suspension of prehydrogenated PtO₂ (240 mg) in water (2.4 ml) and the mixture was stirred under an atmosphere of oxygen for 6 days at room temperature. The same work up procedure as described for <u>17</u> afforded a residue (212.5 mg) which was subjected to PLC (MeOH - CHCl₃ - HCOOH 20: 80: 0.5). The product obtained from the band around Rf 0.30 was chromatographed on activated charcoal (800 mg). The MeOH eluate yielded a crystalline residue (186.5 mg) which was recrystallized from i-PrOH to afford colorless plates of <u>27</u>.

mp 111-118 °C. IR ν_{max} (KBr) cm⁻¹: 3420, 2951, 1734, 1697, 1684, 1648, 1636. 'H-NMR (CD₃OD, 300MHz) δ : 7.42(1H, d, J=0.9 Hz, 3-H), 5.34(1H, d, J=4.7 Hz, 1-H), 4.75(1H, d, J=7.8 Hz, 1'-H), 3.83(1H, d, J=8.8 Hz, 5'-H), 3.69(3H, s, OCH₃), 3.56(1H, t, J=9.0 Hz, 4'-H), 3.41(1H, t, J=8.9 Hz, 3'-H), 3.26(1H, dd, J=9.0 and 7.9 Hz, 2'-H), 2.92(1H, td, J=7.8 and 6.2 Hz, 5-H), 2.37 - 2.20(2H, m, 8- and 9-H), 2.07(1H, dq, J=13.1 and 8.4 Hz, 6-H), 1.79(1H, dtd, J=13.1, 8.6 and 5.6 Hz, 6-H), 1.32(1H, dq, J=12.4 and 8.3 Hz, 7-H), 1.08(3H, d, J=6.7 Hz, 10-H). ¹³-CNMR (CD₃OD, 75.5MHz) δ : 172.73(s, 6'-C), 169.54(s, 11-C), 152.64(d, 3-C), 113.48(s, 4-C), 100.13(d, 1'-C), 96.45(d, 1-C), 77.42(d, 3'-C), 76.79(d, 5'-C), 74.46(d, 2'-C), 73.04(d, 4'-C), 51.67(q, 11-OCH₃), 44.30(d, 9-C), 37.59(d, 8-C), 34.60(d, 5-C), 33.19(t, 7-C), 32.35(t, 6-C), 16.65(q, 10-C). Anal. Calcd. for C_{1.7}H_{2.4}O_{1.0} · 1/2H₂O: C, 51.38; H, 6.34. Found: C, 51.14; H, 6.54.

7-Deoxy-8-epi-loganin 6'-GABA amide (26)

A solution of 27 (100 mg), HOBT (43.1 mg) and EDCI (54.3 mg) in a mixture of anhydrous

acetonitrile and anhydrous THF (1:1) (2.0 ml) was stirred for 1hr under ice-cooling. To the mixture was added a solution of Me-GABA·HCl (43.1 mg) and DMAP (69.2 mg) in anhydrous acetonitrile (1.0 ml) and the mixture was stirred for 1 hr at the same temperature. Stirring was continued for an additional 2 days at room temperature. Evaporation in vacuo of the solvent gave a residue which was purified by PLC (MeOH - CHCl₃ - HCOOH 20:80:0.5). The band around Rf 0.60 afforded a crystalline residue (109.7 mg) which was recrystallized from CHCl₃ - petr. ether to afford colorless needles of 7-deoxy-8-epi-loganin 6'-GABA amide methyl ester.

mp 152-153 °C. UV λ_{max} (MeOH) nm (log ε): 237.6(4.10). IR ν_{max} (KBr) cm⁻¹: 3420, 3340, 2950, 1735, 1714, 1700, 1642. ¹H-NMR (CD₃OD) δ : 7.41(1H, d, J=1.0 Hz, 3-H),5.39(1H, d, J=4.6 Hz, 1-H), 4.74(1H, d, J=7.8 Hz, 1'-H),3.72(1H, d, J=9.5 Hz, 5'-H), 3.69(3H, s, 11-COOC<u>H</u>₃),3.65(3H, s, 1"-OC<u>H</u>₃), 3.53(1H, dd, J=9.5 and 9.0 Hz, 4'-H), 3.40(1H, dd, J=9.0 and 8.5 Hz, 3'-H), 3.29(2H, t, J=7.1 Hz, 4"-H), 3.25(1H, dd, J=8.5 and 7.8 Hz, 2'-H), 2.92(1H, br td, J=14.0 and 7.6 Hz, 5-H), 2.37(2H, t, J=7.3 Hz, 2"-H), 2.37 - 2.20(2H, m, 8- and 9-H), 2.07(1H, m, 6-H), 1.82(2H, quintet, J=7.3 Hz, 3"-H), 1.96 - 1.61(1H, m, 7-H), 1.54 (1H, dddd, J=12.7, 8.3, 5.9 and 4.6 Hz, 6-H), 1.32(1H, dtd, J=11.7, 8.3 and 7.3 Hz, 7-H), 1.08(3H, d, J=6.8 Hz, 10-H₃). ¹³C-NMR (CD₃OD) δ : 175.41(s, 1"-C), 171.54(s, 6'-C), 169.52(s, 11-C), 152.59(d, 3-C), 113.62(s, 4-C), 100.15(s, 1'-C), 96.52(d,1-C), 77.57(d, 5'-C), 76.75(d, 3'-C), 74.36(d, 2'-C), 73.17(d, 4'-C), 52.15(q, 4"-O<u>C</u>H₃), 51.67(q, 11-O<u>C</u>H₃), 44.39(d, 5-C), 39.50(t, 4"-C), 37.56(d, 9-C), 34.59(d,8-C), 33.37(t, 6-C), 32.38(t, 7-C), 32.08(t, 2"-C), 25.64(t, 3"-C), 16.81(q, 10-C). Anal. Calcd. for C₂₂H₃₃O₁₁N: C, 54.20; H, 6.82; N, 2.87. Found: C, 54.33; H, 6.70; N, 2.99.

The foregoing amide methyl ester (200.0 mg) was hydrolyzed with 0.4N aq. NaOH (4.0 ml) in MeOH (4.0 ml) for 20 min under stirring at room temperature. The mixture was worked up as described above for the preparation of <u>15</u>. The crude products were chromatographed on activated charcoal (800 mg). Elution with MeOH afforded <u>26</u> (175 mg) as a white amorphous foam.

UV λ_{max} (MeOH) nm (log ε): 236.5(4.01). IR ν_{max} (KBr) cm⁻¹: 3450, 3300, 2950, 1706, 1666, 1645. ¹H-NMR (CD₃OD) δ : 7.41(1H, d, J=1.0 Hz, 3-H), 5.39(1H, d, J=4.6 Hz, 1-H), 4.74(1H, d, J=7.6 Hz, 1'-H), 3.73(1H, dd, J=9.3 Hz, 5'-H), 3.69(3H, s, 11-OCH₃), 3.53(1H, dd, J=9.3 and 8.6 Hz, 4'-H), 3.40(1H, dd, J=9.0 and 8.6 Hz, 3'-H), 3.26(2H, t, J=7.3 Hz, 4"-H₂), 2.92(1H, td, J=7.8 and 6.1 Hz, 5-H), 2.34(2H, t, J=7.3 Hz, 2"-H₂), 2.38 - 2.22(2H, m, 9- and 8-H), 2.07(1H, dtd, J=12.7, 8.3 and 8.1 Hz, 6-Ha), 1.81(2H, quintet, J=7.3 Hz, 3"-H₂), 1.85 - 1.71(1H, m, 7-Ha), 1.54(1H, dddd, 12.7, 8.3, 6.1 and 4.9 Hz, 6-Hb), 1.32(1H, dq, J=12.0 and 8.1 Hz, 7-Hb), 1.08(3H, d, J=6.8 Hz, 10-H₃). Anal. Calcd. for C₂₁H₃₁O₁₁N: C, 53.27; H, 6.60; N, 2.96. Found: C, 53.02; H, 6.67; N, 2.95. FAB-MS m/z: 474 [M+H]⁺

7-Deoxy-8-epi-loganin 6'-GABA-BSA conjugate (25)

BSA (450 mg) and 26 (140 mg) were condensed using EDCI (66.2 mg) according to the same procedure as described for 13. Dialysis followed by lyophilization provided 25 (496.1 mg). The 26 /BSA ratio of the conjugate 25 was determined by the aforementioned colorimetric method to give 15.2 mole of 26 / mole of BSA.

10-Deoxygeniposide tetraacetate (29)

Geniposide (<u>37</u>) (327.9 mg) was hydrogenolyzed in MeOH (8 ml) with 5% Pd - C (50 mg) in the presence of 60% HClO₄ (6.5 μ l) under an atmosphere of hydrogen. After the uptake of 0.85 equivalent (16 ml) of hydrogen, the reaction was ceased. After removal of the catalyst by filtration, the filtrate was concentrated <u>in vacuo</u> to give a residue (312.3 mg) which was acetylated with acetic anhydride (1 ml) and pyridine (2 ml) overnight at 4 °C. The reaction mixture was poured on iced water and extracted with CHCl₃. The CHCl₃ layer was washed successively with 1N HCl, sat. aq. NaHCO₃ and water, dried and concentrated <u>in vacuo</u> to give a residue which was purified by PLC (diethyl ether). The band around Rf 0.65 gave a crystalline product which was recrystallized from i-PrOH to yield colorless needles of <u>29</u>.

mp 116 - 118 °C. [α]_D -19.83 ° (c, 1.01, MeOH). UV λ_{max} (MeOH) nm (log ε): 236(3.98). IR ν_{max} (KBr) cm⁻¹: 1756, 1707, 1636. 'H-NMR (CDCl₃) δ : 7.39(1H, d, J=1.0 Hz, 3-H), 5.45(1H, quint, J=1.7 Hz, 7-H), 5.18(1H, t, J=9.3 Hz, 3'-H), 5.11(1H, t, J=9.5 or 9.0 Hz, 4'-H), 5.02(1H, t, J=9.3, 7.8 Hz, 2'-H), 4.88(1H, d, J=8.1 Hz, 1'-H), 4.30(1H, dd, J=12.2 and 4.4 Hz, 6'-H), 4.14(1H, dd, J=12.2 and 2.4 Hz, 6'-H), 3.73(1H, ddd, J=10.0, 4.4 and 2.4 Hz, 5'-H), 3.70(1H, s, 11-COOC<u>H</u>₃), 3.14(1H,tdd, J=8.1, 4.6 and 1.0 Hz, 5-H), 2.84 - 2.64(2H, m, 9- and 6-H), 2.19 - 2.13(1H, m, 6-H), 2.08, 2.03, 2.01 and 1.97(each 3H, s, 4 × OCOC<u>H</u>₃), 1.77(3H, br s, 10-<u>H</u>₃). ^{1.3}C-NMR (CDCl₃) δ : 170.54, 170.18, 169.38 and 169.18(each s, O<u>C</u>OCH₃), 167.48(s, 11-<u>C</u>OOMe), 150.74(d, 3-C), 137.58(s, 4-C), 127.10(d, 7-C), 112.70(d, 8-C), 96.42(d, 1'-C), 98.48(d, 1-C), 72.61(d, 5'-C), 72.10(d, 3'-C), 70.79(d, 2'-C), 68.35(d, 4'-C), 61.76(t, 6'-C), 51.16(q, 11-O<u>C</u>H₃), 49.19(d, 9-C), 38.41(t, 6-C), 33.28(d, 5-C), 20.70(q, CO<u>C</u>H₃), 20.60(q, 2 × CO<u>C</u>H₃), 20.34(q, CO<u>C</u>H₃), 15.32(q, 10-C). Anal. Calcd. for C₂₅H₃₂O₁₃: C, 55.55; H, 5.97. Found: C, 55.29; H, 5.94.

10-Deoxygeniposide (<u>3b</u>)

The acetate (29) (2.000 g) was deacetylated in absolute MeOH (20 ml) with 0.26N NaOMe (0.7 ml) for 4 hr under stirring at room temperature. The same work up as described above for <u>1b</u> afforded a crystalline product (1.363 g) which was recrystallized from i-PrOH to yield colorless needles of <u>3b</u>.

mp 169 - 171 °C. [α]_b -5.50 ° (c, 1.00, MeOH). UV λ_{max} (MeOH) nm (log ε): 238(3.91). IR ν_{max} (KBr) cm⁻¹: 3540, 3420, 3380, 3250(sh), 1718, 1640. ¹H-NMR (CD₃OD) δ : 7.47(1H, d, J=1.2 Hz, 3-H), 5.47(1H, m, J=1.2 Hz, 7-H), 5.27(1H, d, J=6.6 Hz, 1-H), 4.71(1H, d, J=7.8 Hz, 1'-H), 3.88(1H, dd, J=12.0 and 1.7 Hz, 6'-Ha), 3.64(1H, dd, J=11.7 and 5.4 Hz, 6'-Hb), 3.39(1H, dd, J=9.0 and 8.8 Hz, 3'-H), 3.25(1H, dd like, J=9.0 and 8.8 Hz, 4'-H), 3.21(1H, dd, J=8.8 and 7.8 Hz, 2'-H), 3.12(1H, tdd, J=7.8, 6.6 and 1.2 Hz, 5-H), 2.72(1-H, br dd, J=15.9 and 7.8 Hz, 6-Ha), 2.63(1H, br dd like, J=7.8 and 6.6 Hz, 9-H), 2.06 (1H, ddq, J=15.9, 6.6 and 2.2 Hz, 6-Hb), 1.81(1H, br s, 10-H₃). ¹³C-NMR (CD₃OD) δ : 169.71(s, 11-C), 153.25(d, 3-C), 140.14(s, 8-C), 127.78(d, 7-C), 112.84(s, 4-C), 100.00(d, 1'-C), 97.54(d, 1-C), 78.35(d,5'-C), 77.94(d, 3'-C), 74.82(d, 2'-C), 71.59(d, 4'-C), 62.76(t, 6'-C), 51.74(q, 11-0CH₃), 50.28(d, 9-C), 39.70(t, 6-C), 35.71(d, 5-C), 16.22(q, 10-C). Anal. Caled. for C_{1.7Hz4}O₆: C, 54.83; H, 6.50. Found: C, 54.53; H, 6.60.

10-Deoxygeniposidic acid (3a)

The compound <u>3b</u> (1.817 g) was hydrolyzed with 0.5N NaOH (25 ml) under stirring overnight at room temperature. Workup as described above for <u>1a</u> afforded colorless needles of <u>3a</u> (1.538 g) which were recrystallized from n-PrOH.

mp 115.5-117 °C. [α]_D -3.84° (c, 0.99, MeOH). IR ν_{max} (KBr) cm⁻¹: 3400, 2960, 2910, 2850(sh), 2700, 2150, 1680, 1635. ¹H-NMR (CD₃OD) δ : 7.48(1H, d, J=1.0 Hz, 3-H), 5.48(1H, t-like, 7-H), 5.26(1H, d, J=6.4 Hz, 1-H), 4.72(1H, d, J=7.8 Hz, 1'-H), 3.89(1H, dd, J=12.0 and 1.7 Hz, 6'-Ha), 3.65(1H, dd, J=12.0 and 5.4 Hz, 6'-Hb), 3.40(1H, dd, J=9.0 and 8.8 Hz, 3'-H), 3.27(1H, dd, J=9.0 and 8.1 Hz, 4'-H), 3.23(1H, dd, J=8.1 and 7.8 Hz, 2'-H), 3.14(1H, br td, J=7.8 and 6.8 Hz, 5-H), 2.73(1H, br dd, J=15.9 and 7.8 Hz, 6-Ha), 2.62(1H, br dd, J=7.6 and 6.4 Hz, 9-H), 2.09(1H, ddq, J=15.9, 6.8 and 2.2 Hz, 6-Hb), 1.82(3H, br s, 10-H₃). ¹³C-NMR (CD₃OD) δ : 171.00(s, 11-C), 151.27(d, 3-C), 140.26(s, 8-C), 127.88(d, 7-C), 113.04(s, 4-C), 100.05(d, 1'-C), 97.62(d, 1-C), 78.42(d, 5'-C), 78.06 (d, 3'-C), 74.92(d, 2'-C), 71.71(d, 4'-C), 62.88(t, 6'-C), 49.77(d, 9-C), 39.82(t, 6-C), 35.93(d, 5-C), 16.30(q, 10-C). Anal. Calcd. for C₁₆H₂₂O₉·1/4n-PrOH: C, 53.63; H, 6.19. Found: C, 53.88; H, 6.48.

10-Deoxygeniposide 11-GABA amide (30)

A solution of <u>3a</u> (41.4 mg), HOBT (31.1 mg) and EDCI (26.4 mg) in anhydrous THF (1.0 ml) was stirred for 1 hr at room temperature. To the mixture was added a solution of Me-GABA·HCl (18.0 mg) and DMAP (30.9 mg) in anhydrous acetonitrile (1.0 ml) with ice-cooling. The mixture was stirred for 1 hr at the same temperature and subsquently overnight at room temperature. The reaction mixture was concentrated <u>in vacuo</u> and the residue purified by PLC (MeOH - CHCl_g -HCOOH 30 : 65 : 5). The band around Rf 0.50 gave a syrup of 10-deoxygeniposide 11-GABA amide methyl ester (48.1 mg).

¹H-NMR (CD₃OD, 300 MHz) $\delta_{\rm H}$: 7.14(1H, d, J=1.2 Hz, 3-H), 5.46(1H, quint, J=1.4 Hz, 7-H), 5.26(1H, d, J=6.1 Hz, 1-H), 4.69(1H, d, J=7.8 Hz, 1'-H), 3.88(1H, dd, J=12.3 and 1.8 Hz, 6'-H), 3.65(1H, s, 1"-OCH₃), 3.47(1H, dd, J=12.4 and 5.2 Hz, 6'-H), 3.38(1H, t, J=8.8 and 7.7 Hz, 3'-H), 3.32 - 3.17(7H, m, 2',4',5',5, 4"-H and NH), 2.74 - 2.62(2H, m, 6- and 9-H), 2.36(2H, t, J=7.4 Hz, 2"-H), 1.99(1H, ddquint, J=15.8, 4.1 and 1.9 Hz, 6-H), 1.82(2H, quint, J=7.1 Hz, 3"-H), 1.81(3H, br s, 10-H). FAB-MS m/z: 458 [M+H]⁺

The above methyl ester (280 mg) was hydrolyzed in MeOH (5.0 ml) with 0.4N aq. NaOH (5.0 ml) for 2 hr at room temperature. Neutralization, purification by PLC (Rf 0.20, MeOH - CHCl₃ 3 : 7) and by column chromatography on activated charcoal (720 mg) (MeOH as eluant) as described above for 15 afforded colorless needles of 30 (160 mg) which were recrystallized from n-PrOH. mp 181-182 °C. UV λ_{max} (MeOH) nm (log ε): 230.5(3.97). IR v_{max} (KBr) cm⁻¹: 3410, 3208(sh), 1710, 1656, 1610, 1512. 'H-NMR (CD₃OD) δ : 7.15(1H, d, J=1.0 Hz, 3-H), 5.46(1H, t-like, N-H), 5.26(1H, d, J=6.1 Hz, 1-H), 4.70(1H, d, J=7.8 Hz, 1'-H), 3.88(1H, dd, J=11.7 and 1.0 Hz, 6'-Ha), 3.65(1H, d, J= 11.7 and 5.1 Hz, 6'-Hb), 3.27(2H, br t, J=7.1 Hz, 1"-H), 3.23(1H, dd, J=8.1 and 7.8 Hz, 2'-H), 2.69(1H, brq, J=7.8 Hz, 6 β -H), 2.67(1H, br t, J=7.3 Hz, 9-H), 2.33(2H, t, J=7.1 Hz, 3"-H), 2.01(1H, br d quintet, J=15.6 and 2.2 Hz, 6 α -H), 1.81(3H, br s, 10-H₃). ¹³C-NMR (CD₃OD) δ : 177.06(s, 4"-C), 170.49(s, 11-C), 147.73(d, 3-C), 140.33(s, 4-C), 127.37(d, 7-C), 116.54(s, 8-C), 99.88(d, 1'-C), 96.89 (d, 1-C), 78.40(d, 5'-C), 78.01(d, 3'-C), 74.92(d, 2'-C), 71.69(d, 4'-C), 62.86(t, 6'-C), 50.60(d, 9-C), 39.94(t, 1"-C), 39.21(t, 6-C), 35.37(d, 5-C), 32.40(t, 3"-C), 25.91(t, 2"-C), 16.19(q, 10-C). Anal. Calcd. for C₂₀H₂₀O₁₀N: C, 54.17; H, 6.59; N, 3.16. Found: C, 53.99; H, 6.65; N, 2.96.

10-Deoxygeniposide 11-GABA-BSA conjugate (28)

BSA (223.2 mg) and <u>30</u> (60 mg) were condensed using EDCI (28.5 mg) according to the same procedure as described for <u>13</u>. Dialysis followed by lyophilization provided <u>28</u> (240.0 mg). The <u>30</u> /BSA ratio of the conjugate <u>28</u> was determined by the aforementioned colorimetric method to give 11.3 mole of <u>30</u> / mole of BSA.

2',3',4'-Triacetyl-6'-trityl-10-deoxygeniposide (33)

A solution of 10-deoxygeniposide (<u>3b</u>) (50 mg) and trityl pyridinium fluoroborate (152.4 mg) in a mixed solvent of anhydrous acetonitrile and THF (each 0.5 ml) was stirred overnight at room temperature. The mixture was in turn acetylated overnight at 4 % with acetic anhydride (0.5 ml) pyridine (1.0 ml). The resulting mixture was diluted with ice-cold water and extracted with CHCl₃. The organic layer was washed successively with 1N HCl, sat. aq. NaHCO₃ and water, dried

and concentrated in <u>vacuo</u>. The residue was purified by PLC (Rf 0.46, AcOEt - benzene 2 : 8) and the crystalline product was recrystallized from EtOH to yield colorless needles of <u>33</u>.

mp 190 - 193 °C. UV λ_{max} (MeOH) nm (log ε): 212.0(4.44), 219(4.31), 228.5(4.29). IR ν_{max} (KBr) cm⁻¹: 2940, 1758, 1704, 1635. 'H-NMR (CDCl₃) δ : 7.46 - 7.20(15H, m, 15 × aromatic H), 7.41(1H, d, J=1.5 Hz, 3-H), 5.52(1H, quintet-like, 7-H), 5.30 - 5.05(3H, m, 2'-, 3'-, and 4'-H), 5.21(1H, d, J=6.4 Hz, 1-H), 4.92(1H, d, J=7.6 Hz, 1'-H), 3.71(3H, s, 11-OCH₃), 3.59(1H, ddd, J=9.8, 4.6 and 2.0 Hz, 5'-H), 3.28(1H, dd, J=10.5 and 4.6 Hz, 6'-Ha), 3.19(1H, ddd, J=7.8, 6.4 and 1.5 Hz, 5-H), 3.06(1H, dd, J=10.5 and 4.6 Hz, 6'-Ha), 2.70(1H, m, 6-H), 2.69(1H, dd, J=7.8 and 6.4 Hz, 9-H), 2.20 - 2.13(1H, m, 6-H), 2.01 and 1.99(each 3H, s, OCOCH₃), 1.88(3H, br s, 10-H₃), 1.71(3H, s, OCOCH₃). Anal. Calcd. for C_{4z}H₄₄O_{1z}: C, 68.10; H, 5.99. Found: C, 67.87; H, 5.99.

6'-Carboxy-2',3',4'-triacetyl-10-deoxygeniposide (34)

A solution of <u>33</u> (900 mg) in 80% aqueous AcOH (35 ml) was stirred at 80 °C for 30 min. The reaction mixture was poured on iced water and was extracted with $CHCl_3$. The extracts were washed with sat. aq. NaHCO₃ and sat. brine and dried. Evaporation <u>in vacuo</u> of the solvent gave a residue which contained 2',3',4'-triacetyl-10-deoxygeniposide together with 2',3',6'-triacetyl-10-deoxygeniposide as a minor product. The mixture was employed without purification in the subsequent step.

To a solution of the detritylated products in acetone (10 ml) was added Jones reagent (1.36 ml) at $-70 \,^{\circ}$ with stirring for 10 min. Stirring was continued at room temperature for 6 hr. After decomposition of the excess reagent with i-PrOH, the reaction mixture was diluted with iced water and extracted with CHCl₃. The organic layer were extracted with sat. aq. NaHCO₃ and the alkaline layer was acidified with 1N HCl and extracted again with CHCl₃. The extracts were washed with brine, dried and concentrated in vacuo. The residue (445.7 mg) was recrystallized from benzene gave a colorless needles of 34.

mp 113 - 115 °C. UV λ_{max} (MeOH) nm (log ε): 234.3(3.90). IR ν_{max} (KBr) cm⁻¹: 3430, 1758, 1703, 1639. ¹H-NMR (CDCl₃) δ : 7.39(1H, d, J=1.0 Hz, 3-H), 6.70 - 6.10(1H, br m, 6'-O<u>H</u>), 5.44(1H, br t-like, 7-H), 5.37 - 5.19(2H, m, 3'- and 4'-H), 5.25(1H, d, J=4.6 Hz, 1-H), 5.10 - 4.98(1H, m, 2'-H), 4.96(1H, d, J=7.8 Hz, 1'-H), 4.18 - 4.05(1H, m, 5'-H), 3.71(3H, s, 11-OC<u>H</u>₃), 3.13(1H, td-like, J=8.1 and 4.4 Hz, 5-H), 2.83 - 2.72(1H, m, 9-H), 2.73(1H, br d, J=16.6 Hz, 6-Ha), 2.12(1H, br d, J=16.6 Hz, 6-Hb), 2.05, 2.03 and 1.97(each 3H, s, 3 × OCOC<u>H</u>₃), 1.76(3H, br s, 10-H₃). ¹³C-NMR (CDCl₃) δ : 170.15 and 169.84(each s, O<u>C</u>OCH₃), 169.76(s, 6'-C), 169.25(s, O<u>C</u>OCH₃), 167.67(s, 11-C), 150.74(d, 3-C), 137.34(s, 4-C), 127.15(d, 7-C), 112.80(s, 8-C), 96.11(d, 1'-C), 95.31(d, 1-C), 72.22(d, 5'-C), 71.86(d, 3'-C), 70.59(d, 2'-C), 69.20(d, 4'-C), 51.28(q, 11-O<u>C</u>H₃), 49.16(d, 9-C), 38.29(t, 6-C),

32.98(d, 5-C), 20.55(q, $OCO\underline{C}H_{\exists}$), 20.51, 20.26 and 15.25(each q, 3 × $OCO\underline{C}H_{\exists}$). Anal. Calcd. for $C_{2\exists}H_{\Xi B}O_{1\exists}$: C, 53.91; H, 5.51. Found: C, 53.70; H, 5.54.

2',3',4'-Triacetyl-10-deoxygeniposide 6'-GABA amide methyl ester (36)

A solution of <u>34</u> (855.6 mg), HOBT (451.2 mg) and EDCI (384.1 mg) in anhydrous THF (10 ml) was stirred for 1 hr at room temperature. To the ice-cold mixture was added a solution of Me-GABA·HCl (279.7 mg) and DMAP (489.5 mg) in anhydrous acetonitrile (5.0 ml) and the resulting mixture was stirred for 1 hr. Stirring was continued overnight at room temperature. Removal of the solvent gave a residue which was purified by PLC (Rf 0.50, acetone - $CHCl_a 2: 8$). The desired product <u>36</u> (549.6 mg) was obtained as colorless needles after recrystallization from aqueous MeOH.

mp 134 - 136 °C. [α]_D -28.25 ° (c, 1.03, MeOH). UV λ_{max} (MeOH) nm (log ε): 238.2(4.16). IR ν_{max} (KBr) cm⁻¹: 3400, 3360, 2950, 1758, 1710, 1705, 1667, 1645. 'H-NMR (CD₃OD) δ : 7.40(1H, d, J=1.2 Hz, 3-H), 6.52(1H, br t, J=5.7 Hz, CON<u>H</u>), 5.49(1H, br s, 7-H), 5.31(1H, t, J=9.3 Hz, 4'-H), 5.15(1H, d, J=5.4 Hz, 1-H), 5.11(1H, t, J=8.8 Hz, 3'-H), 5.0(1H, dd, J=9.2 and 8.1 Hz, 2'-H), 4.94(1H, d, J=8.1 Hz, 1'-H), 3.93(1H, J=9.8 Hz, 5'-H), 3.71(3H, s,11-OC<u>H</u>₂), 3.69(3H, s, 1"-OC<u>H</u>₃), 3.30(2H, qd, J=6.6 and 1.0 Hz, 4"-H₂), 3.16(1H, tdd, J=8.1, 5.1 and 1.0 Hz, 5-H), 2.87 - 2.67(2H, m, 6- and 9-H), 2.37(2H, t, J=7.3 Hz, 2"-H₂), 2.20 - 2.07(1H, m, 6-Hb), 2.06, 2.02 and 1.98(each 3H, s, 3 × OCOC<u>H</u>₃), 1.84(2H, quintet, J=6.8 Hz, 3"-H₂), 1.82(3H, br s, 10-H₃). ¹³C-NMR (CD₃OD) δ : 173.56(s, 1"-<u>C</u>OOCH₃), 169.81, 169.67, 169.18(each s, 3 × <u>C</u>OCH₃), 167.36(s, 11-<u>C</u>OOCH₃), 166.36(s, 6'-<u>C</u>ON), 150.72(d, 3-C), 137.58(s, 8-C), 127.44(d, 7-C), 112.62(s, 4-C), 96.59(d, 1-C), 96.30(d, 1'-C), 72.88 (d, 5'-C), 71.73(d, 3'-C), 70.66(d, 2'-C), 69.55(d, 4'-C), 51.69(q, 1"-O<u>C</u>H₃), 51.20(q, 11-O<u>C</u>H₃), 49.01 (d, 9-C), 38.51(t, 4"-C), 38.46(t, 6-C), 33.64(d, 5-C), 31.18(t, 2"-C), 24.30(t, 3"-C), 20.63, 20.55 and 20.31(each q, 3 × OCO<u>C</u>H₃), 15.12(q, 10-C). Anal. Calcd. for C₂₅H₃₇O₁₄N: C, 54.99; H, 6.10; N, 2.29. Found: C, 54.76; H, 6.12; N, 2.27.

10-Deoxygeniposide 6'-GABA amide (35)

Methyl ester <u>36</u> (400 mg) was hydrolyzed in MeOH (5.0 ml) with 0.4N aq. NaOH (5.0 ml) for 3 hr under stirring at room temperature. The mixture was worked up as described above for the preparation of <u>15</u>. The crude products were chromatographed on activated charcoal (1.2 g). Elution with MeOH afforded <u>35</u> (288 mg) as a white amorphous foam.

'H-NMR (CD₃OD) δ : 7.47(1H, d, J=1.0 Hz, 3-H), 5.50(1H, br quintet-like, 7-H), 5.17(1H, d, J=6.6 Hz, 1-H), 4.76(1H, d, J=7.8 Hz, 1'-H), 3.74(1H, d, J=9.3 Hz, 5'-H), 3.55(1H, dd, J=9.5 and 8.8 Hz, 4'-H), 3.43(1H, dd, J=9.0 and 8.6 Hz, 3'-H), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.16(1H, q, J=7.8 Hz, S'-R), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.16(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.16(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.16(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, dd), J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, dd), J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, dd), J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, dd), J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, dd), J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, m, 2'- and 4"-Hz), 3.35(1H, q, J=7.8 Hz), 3.35 - 3.24(3H, q, J=7.8 Hz), 3.35(1H, q, J=7.8 H

2.82 - 2.55(2H, m, 6- and 9-H), 2.33(2H, t, J=7.3 Hz, 2"-Hz), 2.01(1H, br d, J=16.1 Hz, 6-Hb), 1.90 - 1.68(5H, m, 3"-Hz and 10-Hz). FAB-MS m/z: 490 [M+H]⁺

10-Deoxygeniposide 6'-GABA-BSA conjugate (31)

BSA (178.5 mg) and $\underline{35}$ (51 mg) were condensed using EDCI (22.8 mg) according to the same procedure as described for $\underline{13}$. Dialysis followed by lyophilization provided $\underline{31}$ (190.6 mg). The $\underline{35}$ /BSA ratio of the conjugate $\underline{31}$ was determined by the aforementioned colorimetric method to give 9.6 mole of $\underline{35}$ / mole of BSA.

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