1 H NMR SPECTRA OF PRENYLATED FLAVONOIDS AND PYRANOFLAVONOIDS 1

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Abstract — ¹H Nmr examination of prenylated flavonoids has shown that the presence of the prenyl group at C-3, C-6, or C-8 position of flavone can be deduced from the chemical shift of methylene protons of prenyl group measured in acetone-d₆. The chemical shifts of olefinic proton on pyran ring and A ring proton can be used to distinguish between linear type pyranoflavone and angular type pyranoflavone. The application of this ¹H nmr method to the identification of Atalantia flavone and Artocarpus flavones is discussed.

In our previous 1 H nmr studies on the isoprenoid substituted flavonoids 2 (isoflavone, flavone, flavanone, and flavonol), we reported that the signal of hydrogen-bonded hydroxyl proton (OH-5) of 6-isoprenoid substituted flavonoid appeared at more downfield (0.25-0.30 ppm) than that of the 6-nonsubstituted flavonoids having same B and C rings (measured in acetone-d₆). In contrast, the OH-5 signal of the 8-isoprenoid substituted flavonoid shows upfield shift (0.04-0.10 ppm) compared with that of the flavonoid having same B and C rings and no side chain. $^{3-6}$ The observation of the C-prenylation effects for the OH-5 signal is useful method for the characterization of 6- or 8-prenylated flavonoids, because the measurement

Figure 1. Revised structures of isoprenoid substituted flavonoids*

*: The structures were revised (or would be revised) to corresponding 6- or 8-isoprenoid substituted isomers (see Table 1).

	revised position of C ₅ -unit	chemical shift of OH-5	trivial name	refrences and notes
-			H	4.00
2	8-prenyl	12.14 (A) 12.47 (A)		4,28
3	6-prenyl 6-lavandulyl	12.47 (A) 12.56 (A)	kushenol A	4,28
4	6-geranyl	12.40 (C)	Rushenor A	29,30 31,32
5	6-prenyl	12.30 (C)		33
6	8-prenyl	11.98 (C)	flemiflavanone-D	34,35
7	6-prenyl	12.06 (A)		36,37
8	6-geranyl	13.25 (A)	albanin D	5,38
9	6-geranyl	13.32 (A)	albanin E	5,38
0	6-prenyl	13.30 (A)		12,39
1	6-prenyl	12,30 (A)	glepidotin A	6,40
2	6-(/-hydroxy- isoamyl)	12.40 (A)	2 .	6,36

Table 1. The revised structures of 6- or 8-isoprenoid substituted flavonoids

Table 2. Chemical shifts (δ) of methylene protons of prenyl (geranyl) group and A ring protons of 6- or 8-prenylated (geranylated) flavonoids in acetone-d₅.

	(6-prenyl	flavonoids)	(8-prenylflavonoids)		
(A ring)	CH ₂	H-8	CH ₂	H-6	
isoflavone	3.36-3.38	6.49-6.57	3.45-3.46	6.36-6.43	
flavone	3.35-3.37	6.57-6.63	3.56-3.58	6.33-6.35	
flavonol	3.36-3.37	6.53-6.65	3.52-3.58	6.26-6.39	
(methylene pr	otons on B ring)				
ortho-OH and	-H 3.30-3.43	(9 compounds)			
diortho-OH	3.44-3.45	(6 compounds)			

^{*:} The numbers of reference compounds are as follows; 6-prenylisoflavones (12 compounds), 8-prenylisoflavones (7), 6-prenylflavones (9), 8-prenylflavones (4), 6-prenylflavonols (4), and 8-prenylflavonols (4).

^{*:} The structures reported in the earlier literature were shown in Figure 1.

^{§: (}A); Measured in acetone-d₆, (C): measured in CDCl₃.

requires only a small amount (less than 500 micrograms) of sample. This method can also be used to confirm the structure postulated earlier for the prenylated flavonoid if the chemical shift of the OH-5 signal has been described. In the course of the ¹H nmr studies on the 6- or 8-isoprenoid substituted flavanones, flavones, and flavonols, twelve compounds (1-12) among fifty-seven are found to be necessary reinvestigated. These flavonoids were shown in Figure 1 and Table 1. Structures of the six flavonoids (1, 2, 8, 9, 11, and 12) were revised by synthesis. ⁴⁻⁶

In this paper, we report the chemical shifts of the other proton signals of the prenylated flavonoids and pyranoflavonoids for the confirmation of the location of isoprenoid residue.

1. Prenylated flavanone and isoflavone

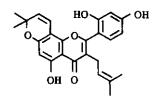
The chemical shifts of the methylene protons of prenyl group $(H_2-1")$ of synthetic 6-prenyl-3',4',5,7-tetrahydroxyflavanone $(1'=(\pm)-1)^4$ and 8-prenyl-3',4',5,7-tetrahydroxyflavanone $(2'=(\pm)-2)^4$ were almost same (1', 6 3.24; 2', 6 3.22) as well as those of aromatic proton on A ring (1', 6 6.02; 2', 6 6.01) measured in acetone-d₆. The 6-prenylated and 8-prenylated flavanones could not be discriminated each other with those chemical shifts as well as with the chemical shifts of C-6 and C-8 carbon signals.

The $\rm H_2$ -1" signal of 6-prenyl group of isoflavone (13)⁸ was observed at δ 3.37, and that of 8-prenyl group of 14^3 appeared at δ 3.45. The H-8 signal of 13 was observed at δ 6.50, and the H-6 signal of 14 appeared at δ 6.36. The chemical shifts of these signals of prenylated isoflavones were summarized in Table 2. These chemical shifts may discriminate between 6-prenylated and 8-prenylated isoflavones, but the additional observation of the OH-5 signal is more reliable for the structure determination of these compounds. The downfield shift (0.08 ppm) of the $\rm H_2$ -1" signal of 8-prenyl group of 14 compared with that of 6-prenyl group of 13 presumably reflects a localization of δ electron at C-8 position, rather than

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20 (atalantoflavone)



24 (morusin)

Figure 2

deshielding effect at the methylene protons as discussed below. In the 13 C nmr spectra of 13 and 14, the chemical shifts of these prenyl groups (C-1" - C-5") show remarkable resemblance, 3 thus, the electron densities of these methylene protons may be almost same. 9 Therefore, the downfield shift of the $\rm H_2$ -1" signal of 14 may be caused by ring current effect. Changes in electron distribution may be detected experimentally by using 13 C nmr spectroscopy (160-200 ppm/ π electron). 10 The C-8 carbon signal of 14 was observed at more upfield (4.06 ppm) in comparison between prenylated carbons of 14 (C-8) and 13 (C-6), therefore, the π electron density at C-8 carbon of 14 is higher (0.02-0.025 π electron) than the density at C-6 carbon of 13. Thus, it is likely that the downfield shift of the $\rm H_2$ -1" signal of the 8-prenyl group is caused by the A ring current effect.

In the case of flavanone derivatives, the chemical shift of C-8 carbon of 1' was similar with that of C-6 carbon of 2' (nonsubstituted carbons). The chemical shift of C-6 carbon of 1' was also similar to that of C-8 carbon of 2' (prenylated carbons). Therefore, the £ electron densities (effects of A ring current) are almost same at C-8 position of the 6-prenylated flavanone and C-6 position of the 8-prenylated flavanone as well as at C-6 position of the 6-prenylated flavanone and C-8 position of the 8-prenylated flavanone. Thus, the H₂-1" and aromatic proton signals of A rings of these 6- and 8-prenylflavanones may be observed at the same region, respectively.

2. Prenylated flavone and flavonol

The $\rm H_2-1"$ and $\rm H-8$ signals of 6-prenylated (6-geranylated) flavones and flavonols were observed in the range of $\rm d$ 3.35-3.37 and $\rm d$ 6.53-6.65 (except 3-prenylflavones), respectively, as shown in Table 2. In contrast, the H-6 signals of 8-prenylated (8-geranylated) flavones and flavonols were observed in the range of $\rm d$ 6.26-6.39, and the $\rm H_2-1"$ signals of these compounds except 3-prenylflavones appeared in the range of $\rm d$ 3.52-3.58. The chemical shifts described above are sufficiently consistent to permit their use in the structure determination of 6-prenylated or 8-prenylated flavone

(flavonol).

The A ring current effects for C-6 and C-8 positions of flavone and flavonol derivatives are probably similar to the effect for those of isoflavone derivatives, because the chemical shifts of C-6 and C-8 carbons are similar between these flavonoids having same substitution on A ring. 3,5 Considering the following result, the B ring current of flavone (flavonol) may cause the downfield shift of H-8 signal. The downfield shift (0.11 ppm) of H-8 signal of 6-prenylflavone (15) was observed to compare with that of 6-prenylisoflavone (13). The value of the B ring current shift for the H-8 of flavones and flavonols was +0.11 ppm which given by the following equation. 11

$$\Delta \delta(\text{ppm}) \approx p(1-\cos^2\theta)/r^3$$
 (see structure (16), in which $p=27$, $\theta=90^{\circ}$, $r=\underline{\text{ca.}} 6.3 \text{ Å}$)

The downfield shift (0.12 ppm) of the $\rm H_2$ -1" signal of 8-prenylflavone (10) compared with that of 8-prenylisoflavone (14) is also explained by the B ring current effect.

The prenylapigenin (10) isolated from Marshallia grandiflora is considered to be prenylated at C-8 position, 12 and the OH-5 signal of this compound appears at 6 13.30. By our previous 1 H nmr method (C-prenylation effect for OH-5), 5 the chemical shift indicates that the structure of the compound is 6-prenylapigenin (Table 1). The chemical shifts of the proton on A ring (6 6.63) and 1 H₂-1" signal (6 3.37) also indicate that the structure of the compound is 6-prenylapigenin (15).

3. 3-Prenylflavone

Several 3-prenylated flavones have been isolated from Moraceae plant. 13 The chemical shifts of 12 -1" signal of 6-prenyl group or H-6 signal of these flavones were similar to those of 3-nonsubstituted flavones, respectively. However, the signals of 12 -1" of 8-prenyl group or H-8 of 3-prenylflavones appeared at more upfield than those of 3-nonsubstituted flavone, respectively, as described below. By comparison of 12 -1" signal (13 3.36,

8-prenyl group) of 3,8-diprenylflavone $(17)^{14}$ with that of 3-nonsubstituted flavone (8-geranyl-2',4',5,7-tetrahydroxyflavone),⁵ the upfield shift (0.22 ppm) of the signal of 17 was found. The chemical shift of H-6 signal (δ 6.32) of 17 was similar to that of the 3-nonsubstituted 8-geranylflavone (δ 6.33). By comparison of H-8 signal of 3,6-diprenylflavone (18) with that of 3-nonsubstituted flavone (6-geranyl-2',4',5,7-tetrahydroxyflavone),⁵ the upfield shift (0.16 ppm) of the signal of 18 (H-8, δ 6.41)¹⁵ was also found. The chemical shift of H₂-1" signal of 6-prenyl group (δ 3.37) of 18 was same as that of the 6-geranylflavone. The upfield shifts may be caused by B ring current, because the 3-prenyl substituent forces the B ring out-of-plane with the chromone ring (A and C rings).¹⁶ The H-8 and H₂-1" of 8-prenyl group of the 3-prenylflavones exist above the B ring plane.

4. Pyranoflavonoid

The difference of the chemical shift was also found between the A ring protons of linear type and angular type pyranoflavones. Further, the H-4" (olefinic proton on pyran ring) signal of angular type pyranoflavone appears at more downfield than that of linear type pyranoflavone as described below. The H-8 signal of linear type pyranoflavone (19) was observed at δ 6.49, and H-4" signal appeared at δ 6.67. The chemical shifts of H-6 and H-4" signals of angular type pyranoflavone (20) have been reported at δ 6.19 and δ 6.93, respectively. 17

In addition, the downfield shift of H-8 signal of 19 (0.08 ppm) compared with that of linear type pyranoisoflavone (21, H-8; δ 6.41) ¹⁸ was found as described in the case of prenylflavone. The downfield shift of H-4" signal of 20 (0.19 ppm) compared with that of angular type pyranoisoflavone (22, H-4"; δ 6.74) ¹⁸ was also found. In contrast, the signal of H-4" of 19 (δ 6.67) was similar to that of 21 (δ 6.69). There is not much difference between the chemical shift of H-6 signal of 20 (δ 6.19) and that of 22 (δ 6.22).

The H-8 signal of linear type 3-prenylated pyranoflavone (23)19 $_{
m was}$

observed at 6 6.27, and H-4" signal of 23 appeared at 6 6.69. The H-6 and H-4" signals of angular type pyranoflavone (24)²⁰ were observed at 6 6.16 and 6 6.61, respectively. The upfield shifts of H-8 of 23 and H-4" of 24 compared with relevant signals of 3-nonsubstituted pyranoflavones (19, 20) were also observed, respectively.

5. Prenyl group of C-3 and B ring

The methylene proton (H_2-9) signals of 3-prenyl groups were observed in the range of 6 3.08-3.16 (13 compounds). On the other hand, the methylene proton signals of prenyl groups on B ring of flavones and isoflavones were observed in the range of 6 3.30-3.45. The prenyl groups could be classified into following two types according to number(s) of hydroxyl group(s) at the adjacent positions. When the both ortho-positions to the prenyl group were replaced by hydroxyl groups, the signals of the methylene protons were observed in the range of 6 3.44-3.45. On the other hand, when one of the ortho-positions to the prenyl group was replaced by a hydroxyl group, and another was nonsubstituted, the signals of the methylene protons were observed in the range of 6 3.30-3.43 (Table 2).

6. Erroneous structure of pyranoflavones

The structure of pyranoflavone isolated from Atalantia ceylanica has been reported as linear type flavone $(19)^{21}$ by comparison with the synthetic carpachromene $(19)^{22}$. The signals of an aromatic proton on A ring and H-4" of the naturally occurring flavone appear at δ 6.15 and δ 6.89, respectively. These chemical shifts indicate that the structure of the pyranoflavone is angular structure (20) which has been isolated from Atalantia racemosa by Banerji et al. 17

Artobilochromen has been isolated from Artocarpus nobilis, and the structure has been reported as formula (25). 23 Although artobilochromen is considered to be prenylated at C-3' position (diortho-substituted prenyl group), the $\rm H_2$ -1" signal of the prenyl group appears at $\rm 6$ 3.16 (in acetone-d₆). The chemical shift of the compound indicates that the prenyl

Figure 3

group exists at C-3 position. The $^1{\rm H}$ and $^{13}{\rm C}$ nmr data of artobilochromen are similar to those of artonin E²⁵ (KB-3, ²⁶ 26) obtained from <u>Artocarpus communis</u> (see Table 3). Thus, they are same compounds, and the formula (25) must be revised to the structure of artonin E (26).

(-)-Dihydrofuranoartobilochromen <u>a</u> has been also isolated from <u>A. nobilis</u> by Kumar <u>et al.</u>, and the structure of the compound was characterized as formula (27). 23 (\pm)-Dihydrofuranoartobilochromen <u>a</u> has been derived from artobilochromen by the treatment with DDQ. The spectroscopic data of (-)-dihydrofuranoartobilochromen <u>a</u> is similar to that of artobiloxanthone $(28)^{24}$ isolated from <u>A. nobilis</u> (see Table 3). Thus, the (-)-dihydrofuranoartobilochromen <u>a</u> is presumably (-)-artobiloxanthone. Further, artobilochromen has been converted to chromanoartobilochromen <u>b</u> (29, isolated from the same plant) with acidic condition, thus, the structure of the compound (29) must be reinvestigated. The chemical shifts of A ring proton and protons of C_5 units of the compound (29) resemble those of oxydihydromorusin $(30)^{14}$ (Table 3). Thus, the compound (29) may be 3-isoprenoid substituted angular type pyranoflavone.

Table 3. $^{13}\mathrm{C}$ Nmr and $^{1}\mathrm{H}$ nmr data of **25-30**

2 3 4		(C)	25 ^b		26 ^C			25 ^d
3	161.6		161.4	H-6	6.15	(d,J=0.7 Hz)	(H-8)	6.16 (s)
	119.9	(1')	120.3	H-3"		(d,J=10 Hz)		5.67 (d,J=10 Hz)
	181.8		182.4	H-4"	6.61	(dd,J=0.7 and 10 Hz)		6.65 (d,J=10 Hz)
4a	104.2		104.6	Me-2"	1.44	(6H,s)		1.46 (s)
5	160.9	(7)	162.2	H-3'	6.60	(s)	(H-6')	6.60 (s)
6	98.7	(8)	99.1	H-6'	6.88	(s)	(H - 3)	6.88 (s)
7	158.4	(8a)	158.9	H ₀ -9	3.16	(br d,J=7 Hz)		3.16 (d,J=7 Hz)
8	100.4	(6)	100.7	ਸ≤10	5.14	(m)		5.14 (t,J=7 Hz)
8a	151.7	(5)	152.2	Me-11	1.47	(br s)		1.46 (s)
9	23.6	(71)	23.8		1.58	(br s)		1.59 (s)
10	121.4	(81)	121.9	OH-5	13.25	(s)		13.22 (s)
11	131.2	(91)	131.7	-				d
12	17.3	(10')	17.4		30 ^e			29 ^d
13	25.4	(111)	25.5	<u> </u>				
2"-M	e 27.6		27.6	H-6	6.16		(H-8)	6.14 (s)
2"	78.0		78.2	H ₂ -9	2.50		(H ₂ -4"')	2.50 (m)
3"	127.5		127.9	H ₂ -10	1.64	• •	(H ² -3"')	
4"	114.1		114.6	Me-11	1.09		(Mē_2"')	1.10 (s)
1'	109.3	(31)	109.8	H-3"	5.65	(d,J=10 Hz)		5.65 (d,J=10 Hz)
21	148.7		149.2	H-4"	6.60	(d,J=10 Hz)		6.61 (d,J=10 Hz)
31	103.9	(3)	104.3	Me-2"	1.45	(s)		1.46 (s)
4'	148.5		149.0	OH-5	13.29	(s)		13.22 (s)
51	138.0		138.5					
6'	116.1		116.5					
	28 ^e				2'	7 ^d	···	
——- Н–6	6.13	(s)		(H-	-8)	6.12 (s)		
Me-2						1.46 (s)		
		(s)				1.48 (s)		
H-3"		(d,J=1	O Hz)			5.67 (d,J=10 Hz)		
H-4"		(d,J=1				6.93 (d,J=10 Hz)		

H-6	6.13 (s)	(H-8)	6.12 (s)
	1.44 (s)		1.46 (s)
	1.47 (s)		1.48 (s)
H-3"	5.67 (d,J=10 Hz)		5.67 (d,J=10 Hz)
H-4"	6.93 (d,J=10 Hz)		6.93 (d,J=10 Hz)
H - 9	2.45 (dd,J=8 and 16 Hz)	(H-7')	2.44 (dd,J=8 and 16 Hz)
	3.42 (dd, $J=2$ and 16 Hz)		3.44 $(d,J=16 Hz)$
H-10	4.01 (br d,J=8 Hz)	(H-8')	4.00 (d, J=8 Hz)
H-12	4.30 (br s)	(H-10')	4.64 (s)
	4.60 (br s)		4.66 (s)
Me-11	1.78 (br s)	(Me-9¹)	1.80 (s)
H-3'	6.59 (s)	(H-6')	6.60 (s)
0H-5	13.38 (s)		13.32 (s)

a: Measured in dmso-d₆, data from reference 25. b: Measured in acetone-d₆, data from reference 24. c: Measured in acetone-d₆, data from reference 25. d: Measured in acetone-d₆, data from reference 23. e: Measured in acetone-d₆.

EXPERIMENTAL

 $^1\mathrm{H}$ Nmr data of the prenylated flavonoids and pyranoflavonoids isolated or prepared in our laboratory were recorded at 23 $^\circ\mathrm{C}$ in acetone-d $_6$ (internal standard, TMS) using JEOL-NM-GX-400 or JEOL-NM-EX-400 nmr spectrometer.

REFERENCES AND NOTES

- Part 6 in the series 'NMR Spectra of Isoprenoid Substituted Phenols.'
 Part 5; see reference 6.
- 2. In this paper, isoprenoid means prenyl (3-methyl-2-butenyl), geranyl, /-hydroxyisoamyl, isoamyl, lavandulyl, and p-menthene groups.
- 3. T. Fukai, Q.-H. Wang, M. Takayama, and T. Nomura, Heterocycles, 1990, 31, 373.
- 4. T. Fukai and T. Nomura, Heterocycles, 1990, 31, 1861.
- 5. T. Fukai and T. Nomura, Heterocycles, 1991, 32, 499.
- 6. T. Fukai and T. Nomura, Heterocycles, 1992, 34, 1213.
- 7. Y. Shirataki, I. Yokoe, M. Endo, and M. Komatsu, Chem. Pharm. Bull., 1985, 33, 444.
- 8. T. Fukai, Q.-H. Wang, and T. Nomura, Heterocycles, 1989, 29, 1369.
- 9. The similarity of the chemical shifts indicates that the electron densities of these methylene carbons are almost same as well as those of C-2" C-5" (reference 10). Therefore, it is likely that the electron densities of these methylene protons are almost same.
- 10. I. I. Schuster, M. Parvez, and A. J. Freyer, <u>J. Org. Chem.</u>, 1988, 53, 5819 and references cited therein.
- 11. R. J. Abraham, J. Fisher, and P. Loftus, 'Introduction to NMR Spectroscopy,' John Wiley & Sons, New York, 1988, p.20.
- 12. F. Bohlmann, C. Zdero, R. M. King, and H. Robinson, Phytochemistry, 1979, 18, 1246.

- 13. T. Nomura, Fortschr. Chem. Org. Naturst., 1988, 53, 86.
- T. Nomura, T. Fukai, and M. Katayanagi, <u>Chem. Pharm. Bull.</u>, 1978, 26, 1453.
- 15. Y. Hano, Y. Matsumoto, K. Shinohara, J.-Y. Sun, and T. Nomura, Heterocycles, 1990, 31, 1339.
- 16. A. V. Rama Rao, M. Varadan, and K. Venkataraman, Indian J. Chem., 1971, 9, 7.
- 17. A. Banerji, D. L. Luthria, and B. R. Prabhu, Phytochemistry, 1988, 27, 3637.
- Y. Hashidoko, S. Tahara, and J. Mizutani, <u>Agric. Biol. Chem.</u>, 1986,
 50, 1797.
- T. Fujimoto, Y. Hano, T. Nomura, and J. Uzawa, <u>Planta Med.</u>, 1984,
 50, 161.
- 20. T. Nomura, T. Fukai, S. Yamada, and M. Katayanagi, Chem. Pharm. Bull., 1978, 26, 1394.
- 21. I. H. Bowen and Y. N. Patel, Planta Med., 1987, 73.
- 22. A. C. Jain, R. Khazanchi, and A. Kumar, Tetrahedron, 1978, 34, 3569.
- 23. N. S. Kumar, G. Pavanasasivam, M. U. S. Sultanbawa, and R. Mageswaran, J. Chem. Soc., Perkin Trans. 1, 1977, 1243.
- 24. M. U. S. Sultanbawa and S. Surendrakumar, Phytochemistry, 1989, 28, 599.
- 25. Y. Hano, Y. Yamagami, M. Kobayashi, R. Isohata, and T. Nomura, Heterocycles, 1990, 31, 877.
- 26. Y. Fujimoto, X.-X. Zhang, M. Kirisawa, J. Uzawa, and M. Sumatra, Chem. Pharm. Bull., 1990, 38, 1787.
- 27. Artobiloxanthone (KB-1, 28) is also isolated from Artocarpus communis by our group and Fujimoto et al., and the compound has been also isolated as an optically inactive compound (reference 26).
- 28. F. Bohlmann, C. Zdero, H. Robinson, and R. M. King, Phytochemistry, 1981, 20, 2245.

- 29. L.-J. Wu, T. Miyase, A. Ueno, M. Kuroyanagi, T. Noro, and S. Fukushima, Chem. Pharm. Bull., 1985, **33**, 3231.
- 30. The calculated chemical shift value for the OH-5 of the structure (3) was § 12.18 (based on 2',4',5,7-tetrahydroxyflavanone, § 12.22). Thus, the structure of kushenol A may be 6-lavandulylated isomer (calculated value for the 6-lavandulylated isomer was § 12.51).
- 31. T. Somleva and I. Ognyanov, Planta Med., 1985, 219.
- 32. T. Fukai, M. Takayama, and T. Nomura, 'Symposium Papers of 33rd Symposium on the Chemistry of Natural Products,' p. 549, October, 1991, Osaka, Japan.
- 33. F. Bohlmann and L. N. Misra, Planta Med., 1984, 50, 271.
- 34. L. A. Mitscher, S. R. Gollapudi, I. K. Khanna, S. D. Drake, T. Hanumaiah, T. Ramaswamy, and K. V. Jagannadha Rao, Phytochemistry, 1985, 24, 2885.
- 35. The OH-5 signal of 3',8-diprenyl-4',5,7-trihydroxyflavanone (euchrestaflavanone A) appears at & 12.10 (reference 4). In contrast, the OH-5 signal of 3',6-diprenyl-4',5,7-trihydroxyflavanone appeared at & 12.46 (reference 41). Thus, the chemical shift of the OH-5 of fleminoflavane-D (& 11.98) indicates that the structure of the compound is 8-prenylflavanone.
- 36. M. P. Souza, M. I. L. Machado, and R. Braz-Filho, Phytochemistry, 1989, 28, 2467.
- 37. The calculated chemical shift value for the OH-5 signal of the structure (7) was 6 ll.67 (based on 3,4',5,7-tetrahydroxyflavanone, 6 ll.71). Thus, the structure of the compound isolated from <u>Bursera leptopholeos</u> may be 6-prenyldihydrokaempferol (calculated value for the 6-prenylated isomer was 6 l2.00).
- 38. A. Shirata, K. Takahashi, M. Takasugi, S. Nagao, S. Ishikawa, S. Ueno,
 L. Muñoz, and T. Masamune, <u>Bull. Sericul. Exp. Sta.</u>, 1982, **28**, 793.
- 39. The OH-5 signal of 8-prenyl-4',5,7-trihydroxyflavone (10) was observed

- at δ 12.97. In contrast, the signal of 6-prenyl-4',5,7-trihydroxy-flavone (15) appeared at δ 13.28. Thus, the structure of the compound isolated from Marshallia grandiflora must be 6-prenylapigenin.
- 40. L. A. Mitscher, G. S. Raghav Rao, I. Khanna, T. Veysoglu, and S. Drake, Phytochemistry, 1983, 22, 573.
- 41. Unpublished data. The compound was isolated from the aerial parts of Glycyrrhiza uralensis.

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