CONVERSION OF ARTEMISINIC ACID INTO (-)-FABIANANE

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Abstract - The conversion of (-)-fabianane from artemisinic acid was achieved in seven steps *via* photooxidative cyclization as a key step.

(-)-Fabianane (8) was isolated by Brown¹ from the aerial parts of Fabiana imbricata(Ruiz and Pavon) Romeo, a plant native to central Chile which is used by the Mapuche Indians to treat kidney and urinary afflictions. (-)-Fabianane (8) is novel seco-amorphane sesquiterpene. It shares several structural similarities such as overall stereochemistry and level of oxygenation at carbons **4.5** and 6 with deoxyartemisinin **(2),2** the metabolite of artemisinin, the antimalarial principle from Artemisia annua. The two compounds differ in that the 5, 12 ester linkage in deoxyartemisinin (2) is replaced by a 5, 11 ether linkage with an additional methyl at carbon 11 in fabianane (8). There have been no previous reports of synthesis of fabianane to the best of our knowledge.³ The novel seco-amorphane structure as well as natural scarcity of $(-)$ -fabianane (8) (0.001 % yield from F. imbricata) have prompted us to prepare the compound by synthesis. In this communication, we would like to report the first synthesis of (-)-fabianane.

(-)-Artemisinic acid **(I),** a versatile chiral starting material for the preparation of many novel analogs4 of artemisinin, was converted to dihydroartemisinyl aldehyde (3) by literature procedures4a in three steps in *65* % overall yield. The oxidative degradation⁵ of the aldehyde (3) with bubbling air in a dimethylformaldehyde solution of 1,4-diazabicyclo[2.2.2]octane (DABCO) and complex of cupric acetate with 2.2'-bipyridyl (70 °C, 1 h) afforded the ketone **(4)6** (mp 34-36 OC) in 44 % yield (Scheme 1). Treatment of the ketone (4) with methylmagnesium bromide in ether (reflux, 2 h) provided 4-amorphen-11-ol (5)⁷ (mp 80-82 °C) in 87 % yield. The tertiary alcohol (5) was also isolated⁸ from *Fabiana imbricata* (Ruiz and Pav.). Photosensitized oxygenation⁹ of 5 afforded new peroxofabianane $(7)^{12}$ (mp 79-80 °C) in 27 % yield via the intermediate (6)^{10,11}[irradiation of 5 under oxygen atomosphere and rose bengal as photosensitizer in CH₃CN/CH₂Cl₂

 $\overline{\mathbf{2}}$

OHC

 $\overline{\mathbf{3}}$

5

Reagents and Conditions: i, air, 1,4-diazabicyclo[2.2.2]octane (0.56 equiv.), cupric acetate (cat.)/2,2'bipyridyl(1:1), dimethylformamide, 70 °C, 1 h, 44 %; ii, methylmagnesium bromide (5.4 equiv.), anhydrous ether, reflux, 2 h, 87 %; iii, O₂, irradiation, rose bengal, CH₃CN/CH₂Cl₂(1:1), -23 °C, 2.5 h, 92 %; iv, copper triflate (0.41 equiv.), oxygen, -23 °C,1 h then room temperature, 2 h,27 % from 5; v, H₂, 5 % Pd/CaCO3, ethanol, room temperature, 22h then p-TsOH (0.4 equiv.), room temperature, 1 h, 91 %.

(1:1), -23 ^oC and subsequent treatment by copper triflate at -23 ^oC to room temperature]. Hydrogenation of 7 with 5 % Pd/CaCO₃ in ethanol and subsequent *in situ* treatment with p-TsOH afforded (-)-fabianane (8)¹³ (oil) in 91 % yield. (-)-Fabianane (8) synthesized from artemisinic acid was identical by comparision of specific rotation and spectral properties with those of natural $(-)$ -fabianane,¹ isolated from F. *imbricata*. This synthesis represents a demonstration of the potential biogenetic relationship between the tertiary alcohol (5) and fabianane (8). The intermediate (6)¹⁰ was obtained in 92 % yield from 4-amorphen-11-ol (5) and the stereoisomer was not detected in this reaction. Unfortunately, direct conversion² of the intermediate (6) with mCPBA in CHCl₃ to fabianane (8) was unsuccessful. The absolute configuration of natural fabianane has been assumed to be as shown in 8 by analogy with other amorphanes⁸ which were recently isolated from F. *imbricata*, such as the tertiary alcohol (5). The relative configuration of (-)-fabianane was confirmed as shown in the structure (8) by this conversion .

In conclusion, this first conversion of (-)-fabianane was stereoselectively established from readily available artemisinic acid in seven steps and provides this scarce natural product in quantities suitable for more extensive biological evaluation.

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- 5. For the proposed mechanism, see V. van Rheenen, Tetahedron Lett., 1969, 985.
- 6. Compound (4): mp 34-36 °C, $[\alpha]_{D}^{25}$ -53.5° (c 1.0, CHCl₃); nmr (300 MHz, CDCl₃): δ 4.82 (s, 1H, 5-H), 2.95 (br s, 1H.), 2.42 (dt, J=3.4,3.2 Hz, lH, 6-H), 2.15 (s, 3H, CH3CO), 1.58 (s, 3H, 14-CH3). 0.87 (d, J=6.20 Hz, 3H, 13-CH₃); ir (CHCl₃): 2923, 2867, 1706 (C=O), 1446, 1351, 1163 cm-¹; CIms mlz: 224 **(M+NH4+,** 100).
- 7. Compound (5): mp 80-82 °C, $[\alpha]_D^{29}$ -3.5 ° (c 1.0, CHCl₃), $[\text{lit.}8 [\alpha]_D$ -4.5 ° (c 1.54, CHCl₃)]; nmr (300MHz. CDC13): 6 5.65 (s, IH, H-5). 2.65 (s, lH, H-6), 2.63 (s, 3H, 15-CH3), 1.28 and 1.26 (s, 6H, 12 and 13-CH₃), 0.87 (d, J=6.6 Hz, 3H, 14-CH₃); ir (CHCl₃): 3428(OH), 2921, 2867, 1448, 1377 cm-I; CIms mlz: *240* (M+NHq+, 100), 222 (M+NH4+ -HzO, 46).
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- 10. Compound (6): oil, nmr (300 MHz, CDC13): 6 7.85 (br s), 5.69 (s, H-5). 1.34 (s, 3H, 15-CH3), 1.33 (s, 3H, 12-CH₃), 1.30 (s, 3H, 13-CH₃), 0.93 (d, J=6.0 Hz, 3H, 14-CH₃); ir (CHCl₃): 3389(OOH), 2944,2867, 1654, 1447, 1368, 1130,756 (C=C) cm-1.
- 11. For the proposed mechanisms, see M. Jung, H. N. EISohly, and J. D. McChesney, Planta Medica, 1990.5 6, 624 and (ref. 2).
- 12. The yield of oxidative cyclization of 5 to 7 was not optimized. Compound (7): mp 79-80 °C, [α]_D²³ +60 ° (c 0.85, CHCl₃); nmr (300 MHz, CDCl₃): δ 5.59 (s, 1H, H-5). 2.30 (m, IH, H-7). 2.10 (m, lH, 2a-H), 1.95 (dt, J=2.4, 4.0 Hz, 2H), 1.75 (m, lH), 1.58 (s, 3H, 15-CH₃), 1.44 (s, 3H, 12-CH₃), 1.24 (s, 3H, 13-CH₃), 0.97 (d, J=6.30 Hz, 3H, 14-CH₃); C-13 nmr (75 MHz, CDCl₃): δ 19.93, 24.41, 25.42, 25.74, 26.37, 30.17, 32.72, 36.91, 37.35, 49.15, 52.14, 83.72, 87.23, 96.32, 103.41; ir (CHCl3): 2928, 2872, 1453, 1379, 1208, 1034, 883, 828 cm-1; CIms m/z: 286 (M+NH₄+, 100).
- 13. Compound (8): (oil), $[\alpha]_{D}^{25}$ -34.5^o (c 0.29, CHCl₃); nmr (300 MHz, CDCl₃): δ 5.64 (s, 1H, H-5), 1.92 (m, 1H, H-7), 1.59 (s, 3H, 12-CH3), 1.55 (s, 3H, 4-CH3), 1.49 (m, 1H, H-1), 1.17 (s, 3H, 13-CH₃), 1.16 (m, 1H, H-10), 0.91 (d, J=6.20 Hz, 3H, 14-CH₃); C-13 nmr (75 MHz, CDCl₃): δ 110.61, 103.78, 94.17.87.66. 47.98, 43.89.35.10, 34.94, 32.92.29.82, 26.43, 25.56, 24.05, 23.49, 18.62; ir (CHCl₃): 2926, 1383, 1021 cm-¹; CIms m/z: 270 (M+NH₄+, 100).