REACTIVITIES OF GIBBERELLINS WITH A CATIONIC EXCHANGE RESIN, DOWEX-50W

Hideharu Seto,* Hideki Hayasaka, Hiroyuki Koshino, Shozo Fujioka, and Shigeo Yoshida

The Institute of Physical and Chemical Research (RIKEN), Hirosawa 2-1, Wako-shi, Saitama 351-0198, Japan

Abstract—A cationic exchange resin containing sulfonic acid functional groups, Dowex-50W, controlled the reactions of gibberellins (GAs) catalyzed by acid: by optimizing the reaction temperature and solvent, GA_1 (1) furnished gibberellin C (7) quantitatively, while GA_4 (2) gave *endo*- GA_4 (10) or 16-O-methyl GA_2 (12) in moderate to good yields, respectively; in a refluxing methanol, GA_3 methyl ester (14) gave triene carboxylic acid (15) (47%), *iso*- GA_3 methyl ester (16) (22%) and diene carboxylic acid (17) (11%). The formation of 15 and 16 was explained by a reaction mechanism *via* a common intermediate, 2β , 3β -epoxide (18).

INTRODUCTION

In previous studies on the structure identification of natural gibberellins (GAs) and their synthesis, the susceptible nature of GAs to acidic conditions has been well documented, which comes from their high functionalities and strained carbon skeleton.¹ The exocyclic $\Delta^{16(17)}$ -double bond of 13-hydroxyGAs, represented by GA₁ (1) and GA₃ (3), undergoes a Wagner-Meerwein rearrangement with inversion of the C/D ring configuration, *e.g.*, 1 to gibberellin C (7) in Scheme 1,² while that of 13-deoxyGAs represented by GA₄ (2) undergoes hydration and double bond migration to the endocyclic 15(16)-position, *i.e.*, 2 to GA₂ (11) and *endo*-GA₄ (10) in Scheme 2.³ On the other hand, the $\Delta^{1(2)}$ -double bond on 3 is much more susceptible. Even in unbuffered aqueous solution at room temperature, 3 is autocatalytically transformed to gibberellenic acid (4), although the rate is very slow,⁴ and under more vigorous or acidic conditions, 3 affords a variety of products including gibberic acid (5) through a complex degradation cascade.⁵ Since controlling these reactions is so difficult, under most acidic conditions, we always face the formation of an awkward complex mixture of rearrangement and degradation products along with the starting material and target compound.

In our early work,⁶ we found that Dowex-50W resin efficiently removed the methoxymethyl (MOM) protecting group of 3- and 13-hydroxyl functions on 3,13-dihydroxyGAs in refluxing aqueous methanol without rearrangement of the C/D ring system. In contrast, the rearrangement was observed to a

HO 3 H
$$CO_{2H}$$
 1 H CO_{2H} 4 CO_{2H} 5

considerable extent when p-toluenesulfonic acid was used. This suggested that Dowex-50W, a cationic exchange resin containing sulfonic acid functional groups attached to a styrene-divinylbenzene copolymer, was mild compared with the corresponding monomeric congener. With a view to elucidating the scope and limitation for controlling the reactions of GAs under acidic conditions, we examined the reactivities of typical natural GAs, 1 and 2, and GA₃ methyl ester (14) with Dowex-50W resin.

RESULTS AND DISCUSSION

As reported earlier,⁶ on treatment of 3,13-di-*O*-MOM-GA₁ (6)⁷ with Dowex-50W resin in a mixed solution of methanol (MeOH) and water (H₂O), 5:1, at 70 °C for 6 h, hydrolytic removal of the MOM groups occurred smoothly to give GA₁ (1) in 91% yield, where the formation of gibberellin C (7) was not detected at all. Elongation of the reaction time to 16 h did not affect this reaction, pointing to the completely stable nature of the C/D ring system on 13-hydroxyGAs under these conditions. However, elevating the reaction temperature gradually caused the Wagner-Meerwein rearrangement of the C/D ring system: when 1 was treated with the resin in MeOH and H₂O, 2:5, at 105 °C for 7 h, the rearrangement was completed to give 7 (84%) along with a considerable amount of its methyl ester (8) (8.6%). As the better conditions for preparation of 7, GA₁ (1) was treated with the resin in dioxane-methanol-H₂O (2:1:2) at 105 °C for 22 h, affording 7 in 96% yield.

Scheme 1

MOMO
$$CO_{2H}$$
 CO_{2H} CO_{2H} CO_{2H} CO_{2Me} CO_{2Me}

a: Resin, MeOH-H2O (5:1), 70 °C, 6 h. b: Resin, dioxane-MeOH-H2O (2:1:2), 105 °C, 22 h.

In contrast, the application of Dowex-50W resin for selective removal of the MOM group on 3-O-MOM- GA_4 (9)⁷ [Dowex resin, MeOH-H₂O, 5:1, 70 °C, 10 h], unexpectedly failed, where the double bond migration to the endocyclic 15(16)-position and addition of the protic solvent to the exocyclic $\Delta^{16(17)}$ -double bond occurred to give a mixture of GA_4 (2) and its double bond isomer, *endo*- GA_4 (10), along

with hydrated GA₄, or GA₂ (11), and its 16-O-methyl ether (12). This evidenced that the exo-methylene of 13-deoxyGAs was more susceptible than that of 13-hydroxyGAs to acid. The low reactivity of exo-methylene of 13-hydroxyGAs may be ascribed to the induction effect of the 13-hydroxyl function which decreases the nucleophilicity of the double bond. Since compounds (10) and (12) are of interest with respect to their biological activity, optimization of the reaction conditions for the preparation of each compound was attempted.

On treatment of 2 with the resin in refluxing tetrahydrofuran (THF) for 15 h, a 3:7 mixture of 2 and 10 was obtained in 92% yield. Elongation of the reaction time to 24 h had no effect on the product ratio, and under the same conditions, 10 also gave the same mixture, indicating that the ratio was that in equilibrium. On the other hand, when the reaction was carried out in refluxing MeOH, 2 afforded 12 in 50% yields and a 3:7 mixture of 2 and 10 in 39% yield. Elongation of the reaction time was not effective for increasing the formation of 12, suggesting that addition and elimination of methanol were reversible under the conditions and the reaction was stationary at this stage.

a: Resin, MeOH-H₂O (5:1), 70 °C, 10 h. b: Resin, THF, reflux, 15 h. c: Resin, MeOH, reflux, 6 h.

Application of the Dowex resin for selective removal of the MOM groups on 3,13-di-O-MOM-GA₃ methyl ester (13)⁷ [Dowex resin, MeOH-H₂O, 5:1, reflux, 6 h] also failed, giving a product mixture speculated to mainly consist of triene carboxylic acid (15), *iso*-GA₃ methyl ester (16) and diene carboxylic acid (17) along with a considerable amount of unidentified products. To simplify the reaction, GA₃ methyl ester (14) was treated with the resin in MeOH at refluxing temperature for 4 h, giving 15, 16 and 17 in 47, 22 and 11% yields, respectively, where 17 was obtained as a single compound but the stereochemistry at the C-2 remains unclear (*vide infra*). These compounds were not interchangeable under the same condition.

Scheme 3

a: Resin, MeOH, reflux, 4 h.

The formation of compounds (15) and (16) may be plausibly explained by a reaction mechanism via a common intermediate, 2β , 3β -epoxide (18), as illustrated in Scheme 4. The C(10)-O(10a) bond fission of 14, facilitated by acid, initiates the intramolecular anti-S_N2' displacement of the acyloxyl group of the bridged lactone by the 3-hydroxyl group, leading to the formation of 18. The subsequent intramolecular nucleophilic attack of the carboxyl function to the oxirane ring, catalyzed by acid, takes place in two competitive ways leading to reaction paths a and b. In path a, the carboxyl function attacks the C-3 position to give β -lactone (19), which is then converted to 20 by dehydration. The terminating step of path a is the S_N2 attack of methanol to the C-3, furnishing the end product (15). On the other hand, in path b, the attack of the carboxyl function at the C-2 furnishes 16.

Scheme 4

This is the first description of the reaction mechanism of acidic degradation of GA₃ congeners by postulating 2β , 3β -epoxide like 18 as an intermediate to final products, where the observed selectivity in cyclization of 18 is consistent with the general tendency, *i.e.*, 4-exo cyclization (18 \rightarrow 19) predominated over 5-endo/7-exo one (18 \rightarrow 16). In this connection, it is well documented that the intermediary carboxylate of 2β , 3β -epoxide (18) is implicated in the transformation of 14 to 16 under weakly alkali condition (0.01 M aqueous KOH), where the alkoxide of 14 initiates the intramolecular anti-S_N2' reaction. However, the selectivity in the subsequent cyclization of the resulting carboxylate anion of 18 is entirely different from that under acidic conditions: 5-endo/7-exo cyclization takes place exclusively to give 16 only.

The structures of new compounds (12, 15 and 17) were verified by MS and NMR spectroscopy. NMR experiments on these compounds and 16 were carried out with 600 MHz by PFG-DQFCOSY, PFG-HMQC and PFG-HMBC, by which all resonances were completely assigned (see EXPERIMENTAL). The α -configuration of 16-OMe on 12 was attested to by the 1D selective PFG-ROESY experiment: methyl protons of 16-OMe [δ 3.15 (s)] showed ROE with 15 α -H [δ 1.82 (d, J_{gem} =13.7 Hz)], while 17-H₃ [δ 1.33 (s)] with 15 β -H [δ 1.57 (dd, J_{gem} =13.7 Hz and $J_{15\beta,14\alpha}$ =2.4 Hz)]. The β -configuration of 3-OMe on 15 was deduced from the NOE differential experiment: no NOE effect was observed between 3-H [δ 3.91 (d, $J_{3,2}$ =5.4 Hz)] and 5-H [δ 3.59 (dd, $J_{5,6}$ =8.8 and $J_{5,11\beta}$ =4.4 Hz)] with β -configuration. Also, no NOE effect between 3-H [δ 4.07 (d, $J_{3,2}$ =4.4 Hz)] and 5-H [δ 3.05 (br d, $J_{5,6}$ =6.8 Hz)] indicated the β -

configuration of 3-OH on 17. However, the stereochemistry at C-2 of 17 could not be defined by NMR analysis. The 2-H resonanced at δ 3.80 with coupling constants, $J_{2,3}$ =4.4, $J_{2,1}$ =3.4, $J_{2,9}$ =2.5 and $J_{2,5}$ =1.5 Hz, and NOEs were observed between 3-H and both of 2-H and methyl protons of the 2-OMe [δ 3.46 (s)]. These data did not rule out possibilities of α - and β -configurations of 2-OMe on 17 possessing A-ring with a half-chair conformation.

CONCLUSION

As described, the application of Dowex-50W resin for removal of the MOM protecting group of 3- and/or 13-hydroxyl function on GAs was found to be limited to GA_1 congeners only. However, the moderate acidity of the resin allowed us to confirm that the susceptibilities of acid-sensitive moieties typical of natural GAs were in the following order: $\Delta^{1(2)}$ -double bond of GA₃ congeners > $\Delta^{16(17)}$ -double bond of 13-hydroxyGAs. In a refluxing methanol solution, the resin effectively controlled the reaction of a GA₃ congener: 14 gave a mixture of 15-17 in a good combined yield without further degradation. This substantiated the implication of 2β ,3 β -epoxide intermediate like 18 in acidic degradation of GA₃ congeners. By varying the reaction temperature and/or solvent, gibberellin C (7) from 1 and *endo*-GA₄ (10) or 16-O-methylGA₂ (12) from 2 were obtained in moderate to good yields, respectively. Since these compounds are of interest with respect to the structure-activity relationships of GAs, the evaluation of a variety of GA activities of these compounds is being undertaken.

EXPERIMENTAL

General.—Melting points were determined on Yanagimoto micromelting point apparatus and are uncorrected. NMR measurements were performed on JEOL JNM-A600 or Bruker AC-300 spectrometer. All spectra were recorded using standard pulse sequences. Chemical shifts were recorded as δ values in parts per million (ppm) relative to tetramethylsilane (δ 0 ppm) for ¹H and the solvent (δ 77.0 ppm) for ¹³C as an internal reference in CDCl₃ solution; or relative to the solvent: δ 2.06 ppm for ¹H and 29.80 ppm for ¹³C, in acetone- d_6 solution. All *J*-values are given in Hz. MS spectra, EI-MS and FAB-MS, were obtained with JEOL-SX102 and JEOL-HX-110 mass spectrometers. Analytical TLC was conducted on micro-slides coated with Merck Kieselgel KG60F-254; the developed plates were stained with 10% (w/v) vanillin in concentrated sulfuric acid at 180 °C. All reactions were carried out under a nitrogen atmosphere. Column chromatography was conducted using Wakogel C-300 [Wako Pure Chemical Industries] as the adsorbent. The ratios of mixed solvents were v/v.

General procedure for reaction of GAs with Dowex resin.—Purchased resin, Dowex-50W-X2 (H+ form), was used after successively washed with dist. H₂O, 2N NaOH, dist. H₂O and 2N HCl, then dist. H₂O on a glass-filter until the filtrate was neutral (wet resin); especially for anhydrous reactions, the resin was further washed with MeOH and Et₂O, and was dried *in vacuo* (dry resin). Resin was added to a solution of a substrate and the heterogeneous mixture was stirred with heating until the starting material disappeared or the reaction reached a stationary state, monitored by TLC. The resin was filtered off through a glass-filter and washed with methanol. The filtrate was concentrated *in vacuo* and the residue was purified by column chromatography on silica gel.

Removal of MOM protecting groups of 3,13-di-O-MOM-GA_I (6).—3,13-di-O-MOM-GA_I (6)⁷ (100 mg, 0.23 mmol) in MeOH (2.5 mL) and water (0.5 mL) was treated with wet resin (200 mg) at 70 °C for 6 h. Column chromatography using AcOEt as the eluent afforded GA_I (1) (73 mg, 91%), whose R_f value and 1 H and 13 C NMR spectra 11 were identical to those of authentic sample gifted by Prof. T. Sassa.

Reaction of GA₁ (1) [Preparation of Gibberellin C (7)].—GA₁ (1) (49 mg, 0.14 mmol) in a mixture of dioxane (1 mL), MeOH (0.5 mL) and H₂O (1 mL) was treated with wet resin (50 mg) at 105 °C for 22 h. Column chromatography using AcOEt-AcOH (50:1) as the eluent gave gibberellin C (7) (47 mg, 96%) as colorless prisms: mp 241-244 °C (decomp) (AcOEt) [lit., 2 mp 265-267 °C (decomp)]; 1H NMR (300 MHz, CDCl₃) δ 1.07 (3H, s, 18-H₃), 1.27 (3H, s, 17-H₃), 2.14 (1H, d, J=18.8 Hz, 14β-H), 2.68 (1H, d, J=6.7 Hz, 6-H), 3.01 (1H, dd, J=18.8 and 3.9 Hz, 14α-H), 3.22 (1H, d, J=6.7 Hz, 5-H), 3.91 (1H, m, 3-H); 13C NMR (75 MHz, acetone-d₆) δ 14.88 and 19.93 (C-17 and -18), 20.00, 26.71, 28.86, 36.19, 48.26 and 50.93 (C-1, -2, -11. -12, -14 and -15), 51.21, 53.28 and 54.23 (C-5, -6 and -9), 49.35, 51.37 and 54.34 (C-4, -8 and -16), 70.92 (C-3), 92.96 (C-10), 174.54 and 178.56 (C-7 and -19), 217.99 (C-13); EI-MS m/z 348 (M+, 74%), 330 (M+-H₂O, 49), 302 (100).

On the other hand, GA₁ (1) (39 mg, 0.11 mmol) in MeOH (2 mL) and water (5 mL) was treated with wet resin (1.0 g) at refluxing temperature for 7 h. Column chromatography using AcOEt as the eluent afforded gibberellin C methyl ester (8) (3.5 mg, 8.6%) as colorless prisms: mp 215-218 °C (AcOEt-hexane) [lit.,² mp 226-228 °C]; ¹H NMR (300 MHz, CDCl₃) δ 1.05 (3H, s, 18-H₃), 1.23 (3H, s, 17-H₃), 2.11 (1H, d, J=18.9 Hz, 14 β -H), 2.64 (1H, d, J=6.7 Hz, 6-H), 3.00 (1H, dd, J=18.9 and 3.9 Hz, 14 α -H), 3.22 (1H, d, J=6.7 Hz, 5-H), 3.74 (3H, s, -CO₂CH₃), 3.89 (1H, m, 3-H); EI-MS m/z 362 (M⁺, 94%), 344 (M⁺-H₂O, 88), 316 (100). Then, elution with AcOEt-AcOH (50:1) gave 7 (33 mg, 84%). The methyl ester of 7, prepared by esterification with ethereal diazomethane in methanol at 0 °C, gave the same R_f value and ¹H NMR spectra as those of 8.

Reaction of 3-O-MOM-GA₄ (9).—3-O-MOM-GA₄ (9)⁷ (20 mg, 0.053 mmol) in MeOH (0.5 mL) and H₂O (0.1 mL) was treated with wet resin (50 mg) at 70 °C for 10 h. Column chromatography using AcOEt–CH₂Cl₂ (5:1) as the eluent afforded a 3:7 mixture of GA₄ (2) and endo-GA₄ (10) (4.8 mg, 24%), and elution with AcOEt gave 16α -O-methylGA₂ (12) (5.3 mg, 27%). The further elution with AcOEt-AcOH (100:1) gave GA₂ (11) (3.4 mg, 18%) as a colorless amorphous powder [lit.,³ mp 256-258 °C (decomp)]: ¹H NMR (300 MHz, CDCl₃) δ 1.16 (3H, s, 18-H₃), 1.38 (3H, s, 17-H₃), 2.68 (1H, d, J=10.8 Hz, 6-H), 3.03 (1H, d, J=10.8 Hz, 5-H), 3.82 (1H, br s, 3-H); EI-MS m/z 350 (M⁺, 2%), 332 (M⁺-H₂O, 57), 270 (100). The R_f value and ¹H NMR spectra of 11 were identical to those of authentic sample prepared by the method described by Aldridge et al.³

Reaction of GA_4 (2).—(a) Preparation of endo- GA_4 (10): GA_4 (2) (100 mg, 0.30 mmol) in dry THF (5 mL) was treated with dry resin (100 mg) at refluxing temperature for 15 h. The crude mixture was shown to consist of 2 and endo- GA_4 (10) in a ratio of 29:71 by the ¹H NMR spectrum. Column chromatography using AcOEt-CH₂Cl₂ (2:1) as the eluent afforded endo- GA_4 (10) (56 mg, 56%) as colorless prisms: mp 228-232 °C (decomp) (AcOEt-MeOH-hexane) [lit., 3 mp 256-260 °C (decomp)]; ¹H NMR (300 MHz, acetone- d_6 with a few drops of D₂O) δ 1.09 (3H, s, 18-H₃), 1.26 (1H, ddd, J=13.6, 12.4 and 7.1 Hz), 2.14 (1H, d, J=10.1 Hz), 2.20 (1H, dd, J=7.8 and 4.7 Hz), 2.42 (1H, d, J=9.9 Hz, 6-H), 3.10 (1H, d, J=9.9 Hz, 5-H), 3.68 (1H, br d, J=2.8 Hz, 3-H), 5.46 (1H, br s, 15-H); ¹³C NMR (75 MHz, acetone- d_6) δ 15.12 and 15.33 (C-17 and -18), 18.93, 19.98, 27.11, 29.02 and 41.70 (C-1, -2, -11. -12 and -14), 41.89, 48.00, 52.22 and 52.77 (C-5, -6, -9 and -13), 55.55 and 56.20 (C-4 and -8), 70.27 (C-3), 94.15 (C-10), 133.51 (C-15), 149.30 (C-16), 174.19 and 178.78 (C-7 and -19); EI-MS m/z 332 (M⁺, 100%), 270 (88). The further elution gave a mixture of 2 and 10 (36 mg, 36%).

(b) Preparation of 16-O-methyl GA₂ (12): GA₄ (2) (93 mg, 0.28 mmol) in dry MeOH (5 mL) was treated with dry resin (100 mg) at refluxing temperature for 6 h. Column chromatography using AcOEt as the eluent afforded a 3: 7 mixture of 2 and endo-GA₄ (10) (37 mg, 39%), and then elution with AcOEt-AcOH (100:1) gave 16-O-methyl GA₂ (12) (51 mg, 50%) as colorless needles: mp 219-221 °C (decomp) (AcOEt); ¹H NMR (600 MHz, CDCl₃) δ 1.17 (3H, s, 18-H₃), 1.33 (3H, s, 17-H₃), 1.49 and 1.81 (each 1H, each m, 12-H₂), 1.57 (1H, dd, J=13.7 and 2.4 Hz, 15 β -H), 1.57 and 1.73 (each 1H, each m, 11-H₂), 1.72 (1H, br d, J=11.2 Hz, 14 α -H), 1.75 and 1.98 (each 1H, each m, 1-H₂), 1.81 (1H, m, 9-H), 1.82 (1H, d, J=13.7 Hz, 15 α -H), 1.82 (2H, m, 2-H₂), 2.01 (1H, dd, J=11.2 and 4.9 Hz, 14 β -H), 2.19 (1H, dd, J=8.8 and 4.9 Hz, 13-H), 2.69 (1H, d, J=10.7 Hz, 6-H), 3.08 (1H, d, J=10.7 Hz, 5-H), 3.15 (3H, s, 16-OCH₃), 3.83 (1H, dd, J=2.9 and 2.4 Hz, 3-H); ¹³C NMR (150 MHz, CDCl₃) δ 14.66 (C-18), 16.02 (C-11), 21.40 (C-17), 22.06 (C-12), 27.22 (C-1), 27.92 (C-2), 35.91 (C-14), 39.35 (C-13), 49.68 (16-OCH₃), 50.83 (C-5), 51.60 (C-8), 52.16 (C-6), 52.54 (C-15), 54.56 (C-4), 54.64 (C-9), 70.28 (C-3), 86.86 (C-16), 94.15 (C-10), 175.20 (C-7), 178.27 (C-19); EI-MS m/z 364 (M⁺, 37%), 332 (M⁺-MeOH, 48), 270 (100); HR-EI-MS m/z [M]⁺: Found, 364.1893. Calcd for C₂₀H₂₈O₆, 364.1886.

Reaction with GA₃ methyl ester (14).—GA₃ methyl ester (14) (99 mg, 0.28 mmol) in dry MeOH (2.5 mL) was treated with dry resin (100 mg) at 70 °C for 4 h. Column chromatography using CH₂Cl₂-AcOEt (10:1) as the eluent afforded *iso*-GA₃ methyl ester (16) (21 mg, 22%), and elution with CH₂Cl₂-AcOEt (2:1) afforded triene carboxylic acid (15) (49 mg, 47%). The further elution with AcOEt-AcOH (100:1) gave diene carboxylic acid (17) (12 mg, 11%).

Triene carboxylic acid (**15**): colorless prisms, mp 220-222 °C (decomp) (AcOEt); ¹H NMR (600 MHz, acetone- d_6) δ 1.20 (3H, s, 18-H₃), 1.67 (1H, m, 12β-H), 1.68 (1H, m, 14α-H), 1.72 (1H, m, 12α-H), 2.03 (1H, m, 11β-H), 2.13 (1H, dd, J=10.3 and 2.4 Hz, 14β-H), 2.16 (1H, br d, J=16.6 Hz, 15β-H), 2.24 (1H, ddd, J=16.6, 2.9 and 2.9 Hz, 15α-H), 2.61 (1H, dd, J=16.1 and 6.3 Hz, 11α-H), 3.39 (3H, s, 3-OCH₃), 3.59 (1H, dd, J=8.8 and 4.4 Hz, 5-H), 3.69 (3H, s, -CO₂CH₃), 3.74 (1H, d, J=8.8 Hz, 6-H), 3.91 (1H, d, J=5.4 Hz, 3-H), 4.90 (1H, br s, 17-H syn to C-15), 5.12 (1H, br s, 17-H syn to C-13), 6.06 (1H, dd, J=9.8 and 5.4 Hz, 2-H), 6.40 (1H, d, J=9.8 Hz, 1-H); ¹³C NMR (150 MHz, acetone- d_6) δ 20.31 (C-18), 21.15 (C-11), 40.01 (C-15), 40.68 (C-12), 49.27 (C-5), 49.68 (C-4), 49.82 (C-6), 51.72 (-CO₂CH₃), 52.98 (C-14), 56.72 (C-8), 57.31 (3-OCH₃), 78.63 (C-3), 79.02 (C-13), 105.85 (C-17), 125.62 (C-1), 127.98 (C-2), 128.08 (C-10), 140.45 (C-9), 155.98 (C-16), 175.43 (C-7), 175.78 (C-19); EI-MS m/z 374 (M+, 14%), 297 (100); HR-EI-MS m/z [M]+: Found, 374.1734. Calcd for C₂₁H₂₆O₆, 374.1729.

iso-GA₃ methyl ester (**16**): a colorless amorphous powder [lit.,² mp 172-173 °C]; ¹H NMR (600 MHz, CDCl₃ with a few drops of D₂O) δ 1.20 (3H, s, 18-H₃), 1.39 (1H, dd, J=11.2 and 2.0 Hz, 14β-H), 1.55 (1H, dd, J=11.2 and 2.9 Hz, 14α-H), 1.56 and 1.76 (each 1H, each m, 12-H₂), 1.76 and 1.95 (each 1H, each m, 11-H₂), 2.24 (1H, br d, J=16.7 Hz, 15β-H), 2.57 (1H, d, J=5.9 Hz, 6-H), 2.59 (1H, ddd, J=16.7, 2.9 and 2.9 Hz, 15α-H), 2.66 (1H, br d, J=5.9 Hz, 9-H), 3.26 (1H, dd, J=5.9 and 2.9 Hz, 5-H), 3.74 (3H, s, -CO₂CH₃), 4.25 (1H, d, J=5.4 Hz, 3-H), 4.75 (1H, dd, J=5.4 and 5.4 Hz, 2-H), 4.98 (1H, dd, J=2.9 and 2.0 Hz, 17-H syn to C-13), 5.79 (1H, ddd, J=5.4, 2.9 and 1.5 Hz, 1-H); ¹³C NMR (150 MHz, CDCl₃ with a few drops of D₂O) δ 16.82 (C-18), 18.62 (C-11), 36.88 (C-12), 38.68 (C-15), 45.37 (C-9), 45.59 (C-5), 48.17 (C-4), 48.54 (C-14), 48.81 (C-6), 49.37 (C-8), 52.04 (-CO₂CH₃), 73.63 (C-2), 75.17 (C-3), 78.76 (C-13), 106.86 (C-17), 113.52 (C-1), 152.13 (C-10), 153.34 (C-16), 175.09 (C-7), 176.69 (C-19); EI-MS m/z 360 (M⁺, 8%), 397 (100).

Diene carboxylic acid (17): a colorless amorphous powder; 1 H NMR (600 MHz, CDCl₃ with a few drops of D₂O) δ 1.36 (3H, s, 18-H₃), 1.51 (1H, br dd, J=10.7 and 2.0 Hz, 14 β -H), 1.57 and 1.78 (each 1H,

each m, 12-H₂), 1.72 and 1.94 (each 1H, each m, 11-H₂), 1.92 (1H, br d, J=10.7 Hz, 14α-H), 2.21 (1H, br d, J=16.1 Hz, 15β-H), 2.52 (1H, m, 9-H), 2.55 (1H, ddd, J=16.1, 2.9 and 2.9 Hz, 15α-H), 2.88 (1H, d, J=6.8 Hz, 6-H), 3.05 (1H, br d, J=6.8 Hz, 5-H), 3.46 (3H, s, 2-OCH₃), 3.71 (3H, s, -CO₂CH₃), 3.80 (1H, dddd, J=4.4, 3.4, 2.5 and 1.5 Hz, 2-H), 4.07 (1H, d, J=4.4 Hz, 3-H), 4.97 (1H, br s, 17-H syn to C-15), 5.12 (1H, dd, J=2.9 and 2.0 Hz, 17-H syn to C-13), 5.42 (1H, ddd, J=3.4, 2.5 and 2.0 Hz, 1-H); ¹³C NMR (150 MHz, CDCl₃ with a few drops of D₂O) δ 21.11 (C-18), 18.77 (C-11), 37.49 (C-12), 39.33 (C-15), 46.21 (C-9), 46.33 (C-4), 47.28 (C-5), 48.80 (C-14), 49.76 (C-8), 50.26 (C-6), 51.78 (-CO₂CH₃), 57.13 (2-OCH₃), 71.74 (C-3), 78.42 (C-2), 78.93 (C-13), 106.53 (C-17), 111.34 (C-1), 146.16 (C-10), 154.26 (C-16), 175.79 (C-7), 176.40 (C-19); EI-MS m/z 374 (M+-H₂O, 7%), 297 (100); HR-FAB-MS m/z ([M+1]+: positive ion, glycerol): Found, 393.1909. Calcd for C₂₁H₂₉O₇, 393.1913.

ACKNOWLEDGMENTS

We gratefully acknowledge generous gifts of GA₁ from Professor T. Sassa at Yamagata University, Japan, and GA₄ from Kyowa Hakko Kogyo Co., Ltd. This work was supported in part by Grant-in-Aid for Scientific Research (C) to H. Seto (No. 09672183) from the Ministry of Education, Science, Sports and Culture of Japan.

REFERENCES AND NOTES

- 1. For reviews, see L. N. Mander, *Natl. Prod. Rep.*, 1986, **5**, 541; J. R. Hanson, *Natl. Prod. Rep.*, 1990, **7**, 41; L. N. Mander, *Chem. Rev.*, 1992, **92**, 573.
- 2. B. E. Cross, J. Chem. Soc., 1960, 3022.
- 3. D. C. Aldridge, J. R. Hanson and T. P. C. Mulholland, J. Chem. Soc., 1965, 3539.
- 4. J. S. Moffatt, J. Chem. Soc., 1960, 3045.
- 5. R. J. Pryce, *Phytochemistry*, 1973, **12**, 507; A. G. Avent, J. R. Hanson and B. H. d'Oliveira, *Mag. Reson. Chem.*, 1989, **27**, 237, and references cited therein.
- 6. H. Seto and L.N. Mander, Synth. Commun., 1992, 22, 2823.
- 7. Compounds (6, 9 and 13) are synthetic intermediates in conversion of natural abundant GA₃ (3) to GA₁ (1) and GA₄ (2) recently developed by us: H. Seto, K. Yoshida, M. Hoshino, T. Shimizu and S. Yoshida, in Abstracts of the 117th Annual Meeting of the Pharmaceutical Society of Japan, Tokyo, March 1997, p. 87, which will be fully reported elsewhere.
- 8. For the mechanisms of acidic degradation of GA₃ (3), see R. J. Pryce, J. Chem. Soc., Perkin Trans. 1, 1974, 1179; P. S. Kirkwood, J. MacMillan and M. Hutchison, *ibid.*, 1982, 707, and references cited therein.
- 9. P. Deslongchamps, 'Stereoelectronic Effects in Organic Chemistry,' Pergamon Press Inc., New York, 1983, pp. 163-208, and references cited therein.
- 10. P. S. Kirkwood, J. MacMillan and M. L. Sinnott, J. Chem. Soc., Perkin Trans. 1, 1980, 2117.
- 11. N. Takahashi, I. Yamaguchi and H. Yamane, in Chemistry of Plant Hormones, ed. By N. Takahashi, CRC Press, Inc., Boca Raton, Florida, 1986, pp. 57-151.