

**A NOVEL DIMERIC LACTONE  
BIS-OSMUNDALACTONE FROM THE JAPANESE  
INEDIBLE MUSHROOM *PAXILLUS*  
*ATROMENTOSUS* VAR. *BAMBUSINUS*<sup>§</sup>**

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**Abstract**-As a part of our systematic investigation of biologically active substances of inedible mushrooms, we studied the chemical constituents of a Basidiomycetes fungus, *Paxillus atrotomentosus* var. *bambusinus* belonging to the Paxillaceae family, and isolated a novel dimeric lactone named bis-osumundalactone (**1**). The absolute structure of compound (**1**) was determined by a combination of FAB-MS and high resolution NMR spectra with chemical reactions.

## 1. INTRODUCTION

We have been interested in the biologically active substances present in inedible mushroom without toxicity. Recently, we found that an inedible mushroom, *Cryptoporus volvatus* (Polyporaceae) contained a large amount of novel bitter drimane sesquiterpenoids, cryptoporic acids (CPAs) A ~ G which showed strong inhibition of superoxide anion radical release.<sup>1</sup>

*Paxillus atrotomentosus* belonging to the Paxillaceae family is a lignicolous inedible mushroom with a large cap and frequently appears on decayed pine trees. Previously, Steglich *et al.*<sup>2-4</sup> reported the isolation and structural characterization of novel leucomentin, flavomentin and

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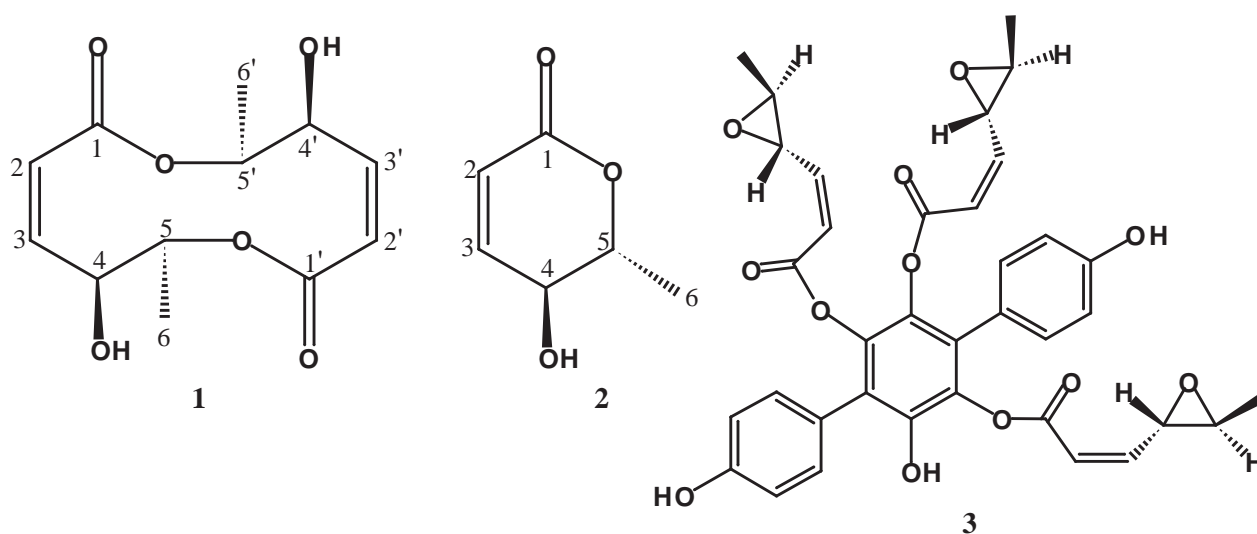
<sup>§</sup>Dedicated to the celebration of 70th birthday of Professor James P. Kutney

spiromentin derivatives from the European *P. atrotomentosus*.

Recently, we have also isolated a new  $\delta$ -lactone, (+)-osumundalactone (**2**) and six new spiromentins E-J from the ethyl acetate extract of the Japanese *P. atrotomentosus*.<sup>5</sup> An inedible mushroom *Paxillus atrotomentosus* var. *bambusinus* is a variety of *P. atrotomentosus*, and rarely appears on decayed bamboo. In this paper, we report the isolation and characterization of a new dimeric lactone, bis-osumundalactone (**1**) by FAB-MS, high resolution NMR spectra and chemical reactions.

### Isolation of Bis-osumundalactone

The EtOAc extract of fresh material of *Paxillus atrotomentosus* var. *bambusinu* was subjected repeatedly to column chromatography on silica gel using  $\text{CHCl}_3$ -MeOH gradient and HPLC (Diol; 20%EtOAc- $\text{CHCl}_3$ ) to give bis-osmundalactone (**1**) with a known phenolic compound, leucomentin-3 (**3**).<sup>3</sup>



### Structural elucidation of Bis-osumundalactone

The FT-IR and UV spectral data of bis-osumundalactone (**1**) showed a hydroxy group [ $3408\text{ cm}^{-1}$ ] and an  $\alpha,\beta$ -unsaturated ester [ $1720\text{ cm}^{-1}$ ;  $\lambda_{\text{max}} 204\text{ nm}$ ]. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of **1** in  $\text{CDCl}_3$  indicated the presence of a secondary methyl [ $\delta_{\text{H}} 1.49$  (*d*,  $J=6.2\text{ Hz}$ ),  $\delta_{\text{C}} 18.1$  (*q*)], an  $\alpha,\beta$ -unsaturated ester [ $\delta_{\text{H}} 5.98$  (1H, *dd*,  $J= 10.0, 1.8\text{ Hz}$ ),  $6.86$  (1H, *dd*,  $J= 10.0, 2.4\text{ Hz}$ );  $\delta_{\text{C}} 120.5$  (*d*),  $149.0$  (*d*)] and a secondary hydroxyl group [ $\delta_{\text{H}} 4.25$  (1H, *m*);  $\delta_{\text{C}} 67.6$  (*d*)]. The COSY spectrum of **1** suggested the presence of the partial structure as shown in Figure 1. The  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR

and COSY spectra of **1** were quite similar to those of (+)-osmundalactone (**2**) as shown in Tables 1 and 2.

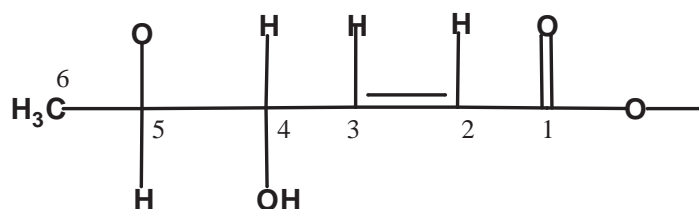


Figure 1. Partial structure of bis-osmundalactone (**1**).

Table 1. <sup>1</sup>H NMR data of bis-osmundalactone (**1**), osmundalactone (**2**) and the diacetate (**4**) of **1**<sup>a</sup>

H	bisosmundalactone ( <b>1</b> )	osmundalactone ( <b>2</b> )	diacetate ( <b>4</b> )
2	5.98 ( <i>dd</i> , 10.0, 1.8)	5.88 ( <i>dd</i> , 10.0, 2.0)	6.11 ( <i>dd</i> , 9.9, 1.6)
3	6.86 ( <i>dd</i> , 10.0, 2.4)	6.85 ( <i>dd</i> , 10.0, 2.3)	6.77 ( <i>dd</i> , 9.9, 3.2)
4	4.25 ( <i>m</i> )	4.19 ( <i>m</i> )	5.28 ( <i>ddd</i> , 6.6, 3.2, 1.6)
5	4.39 ( <i>dq</i> , 8.8, 6.2)	4.33 ( <i>dq</i> , 8.8, 6.6)	4.60 ( <i>quint</i> , 6.6)
6	1.49 ( <i>d</i> , 6.2)	1.42 ( <i>d</i> , 6.6)	1.44 ( <i>d</i> , 6.6)
2'	5.98 ( <i>dd</i> , 10.0, 1.8)		6.11 ( <i>dd</i> , 9.9, 1.6)
3'	6.86 ( <i>dd</i> , 10.0, 2.4)		6.77 ( <i>dd</i> , 9.9, 3.2)
4'	4.25 ( <i>m</i> )		5.28 ( <i>ddd</i> , 6.6, 3.2, 1.6)
5'	4.39 ( <i>dq</i> , 8.8, 6.2)		4.60 ( <i>quint</i> , 6.6)
6'	1.49 ( <i>d</i> , 6.2)		1.44 ( <i>d</i> , 6.6)
OH	2.83 ( <i>br. d</i> , <i>J</i> =6.4)	4.23 ( <i>br. d</i> , <i>J</i> =6.2)	
OAc			2.14 ( <i>s</i> )

<sup>a</sup>Chemical shifts from TMS (multiplicity, *J* in Hz) in CDCl<sub>3</sub> and assignments from COSY spectrum

Table 2.  $^{13}\text{C}$  NMR data of bis-osmundalactone (**1**), osmundalactone (**2**) and the diacetate (**4**) of **1**<sup>a</sup>

C	bisosmundalactone ( <b>1</b> )	osmundalactone ( <b>2</b> )	diacetate ( <b>4</b> )
1	163.4 ( <i>s</i> )	164.1 ( <i>s</i> )	162.1 ( <i>s</i> )
2	120.5 ( <i>d</i> )	119.6 ( <i>d</i> )	122.8 ( <i>d</i> )
3	149.0 ( <i>d</i> )	150.2 ( <i>d</i> )	142.8 ( <i>d</i> )
4	67.6 ( <i>d</i> )	67.0 ( <i>d</i> )	67.7 ( <i>d</i> )
5	79.1 ( <i>d</i> )	79.2 ( <i>d</i> )	76.4 ( <i>d</i> )
6	18.1 ( <i>q</i> )	17.9 ( <i>q</i> )	18.3 ( <i>q</i> )
1'	163.4 ( <i>s</i> )		162.1 ( <i>s</i> )
2'	120.5 ( <i>d</i> )		122.8 ( <i>d</i> )
3'	149.0 ( <i>d</i> )		142.8 ( <i>d</i> )
4'	67.6 ( <i>d</i> )		67.7 ( <i>d</i> )
5'	79.1 ( <i>d</i> )		76.4 ( <i>d</i> )
6'	18.1 ( <i>q</i> )		18.3 ( <i>q</i> )
OAc			20.8 ( <i>q</i> )
			170.0 ( <i>s</i> )

<sup>a</sup>Chemical shifts from TMS in  $\text{CDCl}_3$  and assignments from HMQC and HMBC spectra

The fast atom bombardment (FAB)-mass spectrometry (MS) of bis-osmundalactone (**1**) gave peaks at  $m/z$  257  $[\text{M}+\text{H}]^+$  and 289  $[\text{M}+\text{Na}]^+$ . The high resolution FAB-MS showed a peak at  $m/z$  257.1034 corresponding to a molecular formula of  $\text{C}_{12}\text{H}_{16}\text{O}_6$  which was two times that of osmundalactone (**2**). In the  $^{13}\text{C}$  NMR spectrum of **1**, only 6 carbon signal were observed, suggesting that **1** might be a symmetrical dimer of **2**. Acetylation of **1** with  $\text{Ac}_2\text{O}$  and pyridine afforded diacetate (**4**) [CI-MS ( $\text{CH}_4$ ) and FAB-MS:  $m/z$  341  $[\text{M}+\text{H}]^+$ ;  $\delta_{\text{H}}$  2.14 (3H, *s*);  $\delta_{\text{C}}$  20.8 (*q*), 170.0 (*s*)] indicating the presence of two secondary hydroxyl groups in **1**. The relative structure

of **1** was inferred by the careful analysis of HMQC ( $^1\text{H}$  detected multiple quantum coherence), HMBC and NOESY spectra of **4**. In HMBC spectrum (Figure 2) of **4**, H-5 was correlated with C-3, C-4 and C-1'. Bis-osmundalactone (**1**) might be a  $C_2$ -symmetrical dimer, since **1** showed an optical activity  $\{[\alpha]_D^{20} + 27.0^\circ (\text{CHCl}_3)\}$ .

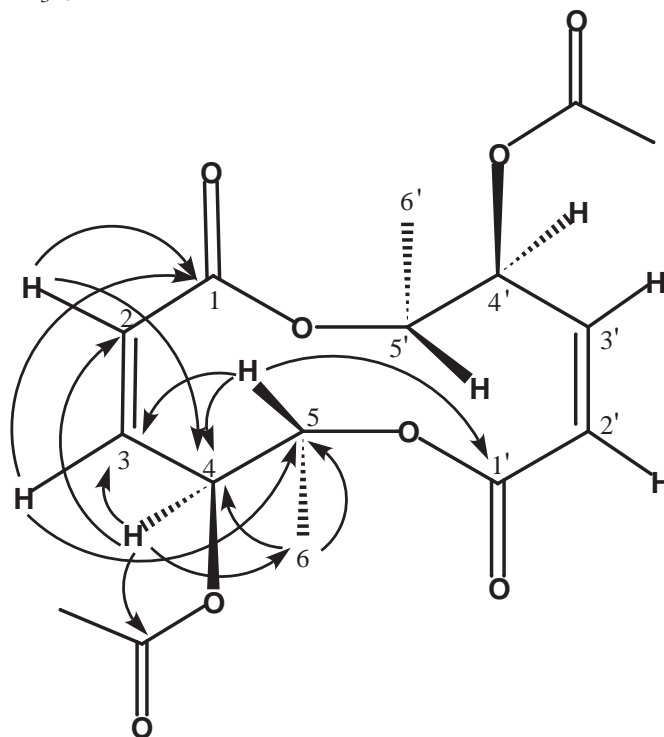


Figure 2. HMBC correlation of the diacetate (**4**)

The absolute configuration of **1** was determined by chemical correlation with (+)-osmundalactone (**2**). Hydrolysis of compound (**1**) with 10 % NaOH, followed by neutralization as usual manner gave **2**, the spectral data ( $^1\text{H}$  and  $^{13}\text{C}$  NMR, IR, and MS) of which were identical with those of (+)-osmundalactone (**2**) isolated from *P. atrotomentosus*.<sup>5</sup> The melting point and specific rotation of the synthetic product were mp  $72\text{--}74^\circ$  and  $[\alpha]_D^{20} -29.8^\circ$  ( $c$  0.92,  $\text{CHCl}_3$ ), while the literature values<sup>5</sup> were mp  $73\text{--}74^\circ$  and  $[\alpha]_D^{20} -28.8^\circ$  ( $c$  2.35,  $\text{CHCl}_3$ ). In conclusion, the absolute configuration of bis-osmundalactone (**1**) was established as (4*S*, 5*R*, 4'*S*, 5'*R*)-configurations.

Table 3 showed the distribution of osmundalactone (**2**) and its related compounds in three *Paxillus* species. Both Japanese and European *P. atrotomentosus* produce leucomentin-3 (**3**).<sup>3</sup> It is noteworthy that the Japanese *Paxillus atrotomentosus* var. *bambusinus* elaborates the characteristic bis-osmundalactone (**1**) without the monomeric osmundalactone (**2**).

Table 3. Distribution of bis-osmundalactone (1), osmundalactone (2) and leucomentin-3 (3) in *Paxillus* species

species	(1)	(2)	(3)
Japanese <i>P. atrotomentosus</i> <sup>5)</sup>		++++*	+++
European <i>P. atrotomentosus</i> <sup>2-4)</sup>			+++
<i>P. atrotomentosus</i> var. <i>bambusinus</i>	++		+++

\*The symbol (++ ~ +++) was shown on the base of a concentration of products

## EXPERIMENTAL

IR spectra were measured on a Jasco FT-IR 500 spectrophotometer. <sup>1</sup>H and <sup>13</sup>C NMR were recorded on a Varian unity 600 (<sup>1</sup>H; 600 MHz, <sup>13</sup>C; 150 MHz) or a Varian Unity 200 (<sup>1</sup>H; 200 MHz, <sup>13</sup>C; 50 MHz) spectrometer. The solvent used for NMR spectra was CDCl<sub>3</sub> unless otherwise stated. MS spectra were measured on a JEOL JMS HX-100 or a JEOL AX-500 spectrometer. The specific rotation was taken on a JASCO DIP-140 polarimeter. Silica gel 60 for column chromatography was purchased from Merk.

**Isolation of Bis-osmundalactone (1)** The fresh materials (1.51 kg) of *Paxillus atrotomentosus* var. *bambusinus* collected in Nishinomiya-city, Hyougo prefecture, Japan, in July 1997, was extracted with EtOAc (3 L) for two times for 1 day at rt. After filtration, the brown solution was evaporated *in vacuo* to afford dark brown residue (55.74 g). A part of residue (4.0 g) was chromatographed on silica gel (100g) with a gradient solvent system of CHCl<sub>3</sub>-EtOAc increasing the amount of 5% portion EtOAc stepwise to give 153 fractions. 50% EtOAc-CHCl<sub>3</sub> eluate (Fr. 64-83) was evaporated *in vacuo* to afford the crude oil (218.3 mg), which was further purified by reversed HPLC (Nucleosil 5C<sub>18</sub>; 50% H<sub>2</sub>O-MeCN) to give leucomentin-3 (3)<sup>3</sup> (174 mg), the spectral data (IR, <sup>1</sup>H and <sup>13</sup>C NMR) of which were identical with those of leucomentin-3.<sup>3</sup> A

remaining crude oil (51.74 g) was chromatographed on silica gel (600 g) with a gradient solvent system of CHCl<sub>3</sub>-MeOH increasing the amount of 1% portion MeOH stepwise to give 35 fractions. 5% MeOH-CHCl<sub>3</sub> eluate (Fr. 14-16) was evaporated in *vacuo* to afford the crude oil (4.02 g), a part (200 mg) of which was chromatographed on Diol with CHCl<sub>3</sub>-EtOAc gradient, followed by HPLC (Diol; 20%EtOAc-CHCl<sub>3</sub>) to give bis-osmundalactone (**1**) (23 mg).

**Bis-osmundalactone (1)** colorless oil,  $[\alpha]_D^{20} +27.0^\circ$  (*c* 1.00, CHCl<sub>3</sub>); CI-MS (CH<sub>4</sub>): *m/z* 257 [M+H]<sup>+</sup>; FAB(+)-MS: *m/z* 257 [M+H]<sup>+</sup>, 289 [M+Na]<sup>+</sup>; HRFAB(+)-MS: *m/z* 257.1034 [M+H]<sup>+</sup>, C<sub>12</sub>H<sub>17</sub>O<sub>6</sub> requires 257.1025; FT-IR (KBr)cm<sup>-1</sup>: 3408 (OH), 1720 (COO); UV (EtOH) λ<sub>max</sub>nm (log ε); 204.0 (4.33); <sup>1</sup>H and <sup>13</sup>C NMR(CDCl<sub>3</sub>)(Tables 1 and 2).

**Acetylation of Compound (1)** A solution of compound (**1**) (10 mg) in pyridine (1 mL) was treated with acetic anhydride (1 mL). The mixture was stirred at rt overnight. Water was added and the mixture was extracted with CHCl<sub>3</sub>. The organic phase was washed with 1N HCl, 5% NaHCO<sub>3</sub> solution and brine, dried (MgSO<sub>4</sub>), and evaporated to give a residue which was purified by a silica gel column chromatography with hexane-AcOEt gradient to afford a diacetate (**4**) (7 mg) as a colorless oil;  $[\alpha]_D^{20} +38.9^\circ$  (*c* 0.75, CHCl<sub>3</sub>); CI-MS (CH<sub>4</sub>): *m/z* 341 [M+H]<sup>+</sup>, 193, 171, 126, 111 (100%), 84; FAB-MS: *m/z* 341 [M+H]<sup>+</sup>, 363 [M+Na]<sup>+</sup>; FT-IR (KBr)cm<sup>-1</sup>: 1734 (COO), 1636 (C=C); UV (EtOH) λ<sub>max</sub>nm (log ε); 205.5 (4.33); <sup>1</sup>H and <sup>13</sup>C NMR(CDCl<sub>3</sub>)(Tables 1 and 2).

**Chemical conversion of bis-osmadulactone (1) into osmadulactone (2)** To a solution of compound (**1**)(10 mg) in MeOH (3 mL) was added 10% NaOH solution (0.5 mL). The mixture was stirred at rt for 12 h. The reaction mixture was poured in 10% HCl solution (20 mL) and stirred at rt for 1 h. The mixture was extracted with CHCl<sub>3</sub> (50 mL x 3). The organic layer was washed with brine, dried (MgSO<sub>4</sub>) and evaporated under reduced pressure to give crude product (14 mg) which was recrystallized from EtOAc-Et<sub>2</sub>O to furnish osmadulactone (**2**) (7 mg) as colorless needles; mp 81.0-83.0° (lit.,<sup>5</sup> mp 81.5-82.5°),  $[\alpha]_D^{20} +71.5^\circ$  (*c* 0.70, CHCl<sub>3</sub>) {lit.,<sup>5</sup>  $[\alpha]_D +70.7^\circ$  (*c*1.27, CHCl<sub>3</sub>)}; HRCI-MS; *m/z* 129.0558 [M+H]<sup>+</sup>, C<sub>12</sub>H<sub>17</sub>O<sub>6</sub> requires 257.1025; CI-MS (CH<sub>4</sub>): *m/z* 129 ([M+H]<sup>+</sup>, 100%), 111, 103, 87. The spectral data (IR, UV, <sup>1</sup>H and <sup>13</sup>C NMR) of synthetic product were identical with those of natural osmadulactone (**2**).<sup>5</sup>

## ACKNOWLEDGMENT

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