## A NEW AURONE GLUCOSIDE AND A NEW CHALCONE GLUCOSIDE FROM *BIDENS BIPINNATA* LINNE

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Abstract—A new aurone glucoside, bidenoside A, (*Z*)-6-*O*-(3, 6-di-*O*-acetyl- $\beta$ -D-glucopyranosyl)-6, 7, 3', 4'-tetrahydroxyaurone (1) and a new chalcone glucoside, bidenoside B, 2', 4', 6'-trimethoxy-4-*O*- $\beta$ -D-glucopyranosyl - dihydrochalcone (4), together with five known constituents have been isolated from the aerial parts of *Bidens bipinnata*. These structures have been elucidated on the basis of spectroscopic methods.

*Bidens bipinnata*, a weed of the Asterceae family, is widely distributed in China. It has been used as a folk medicine for various diseases, such as inflammations, rheumatism, sore throat, hypertension and diabetes.<sup>1</sup> In this paper we report the isolation and structural elucidation of a new aurone glucoside, (*Z*)-6-*O*-(3, 6-di-*O*-acetyl-  $\beta$  -D-glucopyranosyl)-6, 7, 3', 4'-tetrahydroxyaurone (1), two known auron glucosides, (*Z*)-6-*O*- $\beta$ -D-glucopyranosyl-6, 7, 3', 4'-tetrahydroxyaurone (2)<sup>2</sup> and (*Z*)-6-*O*-(6-*O*-acetyl- $\beta$ -D-glucopyranosyl)-6, 7, 3', 4'-tetrahydroxyaurone (2)<sup>2</sup> and (*Z*)-6-*O*-(6-*O*-acetyl- $\beta$ -D-glucopyranosyl)-6, 7, 3', 4'-tetrahydroxyaurone (3),<sup>2</sup> a new chalcone glucoside, 2', 4', 6'-trimethoxy-4-*O*- $\beta$ -D-glucopyranosyldihydrochalcone (4), together with three known flavonoid glycosides, quercetin 3-*O*- $\beta$ -D-glucopyranoside (5),<sup>3</sup> quercetin 3-*O*- $\alpha$ -L-rhamnoside (6)<sup>4</sup> and iso-okanin 7-*O*- $\beta$ -D-glucopyranoside (7)<sup>5</sup> from this plant.

The EtOAc soluble part obtained from EtOH extract was separated on silica gel and octadecyl silica gel (ODS) column chromatography, successively, followed by HPLC to isolate compounds (1, 4).

The *n*-butanol fraction was separated on silica gel and octadecyl silica gel (ODS) column chromatography, successively, followed by HPLC to isolate compounds (2, 3, 5, 6, 7).

Bidenoside A (1), an orange powder, showed positive to the Molish reaction and give an  $[M+H]^+$  ion peak at m/z 533 in positive ion FAB-MS. The molecular formula was determined as  $C_{25}H_{24}O_{13}$ 

on the basis of HR-FAB-MS (M+1; Calcd for 533.1294; Found: 533.1271). The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra showed a presence of glucopyranosyl unit. The anomeric proton signal at 5.01 ppm (d, J= 8.0Hz) in the <sup>1</sup>H-NMR spectrum indicated  $\beta$ -glucose. The <sup>1</sup>H-NMR signals at  $\delta$  7.46 (d, J= 1.8 Hz) , 6.76 (d, J= 8.3 Hz) and 7.26 (dd, J= 8.3, 1.8 Hz) indicated the presence of 2, 5, 6 related aromatic protons (aurone with a 3', 4'-disubstituted B-ring) and at  $\delta$  7.13 (d, J= 8.5 Hz) and 6.97 (d, J= 8.5 Hz) showed ortho related protons in the A-ring. All data closely related to those of the co-occurring aurone glucoside (2, 3). On comparison of its <sup>1</sup>H- and <sup>13</sup>C-NMR spectral data with those of 3, 1 was very similar to 3 except for the sugar moiety (Table 1). The two singlet signals of 1 at  $\delta$  1.97 and 2.05 in the <sup>1</sup>H-NMR were

consistent with two acetyl groups attached to the sugar moiety. Therefore, compound **1** has another acetyl group more than **3**. On comparing with **3**, C-2", C-3" and C-4" of **1** appeared with up- and/or down- field shifts, -1.8 ppm, +0.5 ppm and -1.9 ppm, respectively. These data indicated that another acetyl group was linked to the C-3"-hydroxyl position of the glucose moiety. All carbon and proton signals were assigned by the aids of <sup>1</sup>H-<sup>1</sup>H COSY and <sup>1</sup>H-<sup>13</sup>C COSY.

Thus **1** was determined as (*Z*)-6-*O*-(3, 6-di-*O*-acetyl- $\beta$ -D-glucopyranosyl)-6, 7, 3 ', 4'-tetrahydroxyaurone. Bidenoside B (**4**), a light yellow powder, showed positive to the Molish reaction and gave an [M+H]<sup>+</sup> ion peak at *m*/*z* 479 in postive ion FAB-MS. The molecular formula was determined as C<sub>24</sub>H<sub>30</sub>O<sub>10</sub> on the basis of HR-EI-MS (Calcd for 478.1838; Found: 478.1824). The <sup>1</sup>H-NMR signals of **4** at  $\delta$  6.98 ( 2H, d, *J*= 8.7 Hz) and  $\delta$  7.08 (2H, d, *J*= 8.7 Hz) indicated a chalcone with a 4-substituted B-ring and two same meta-H atoms at  $\delta$  6.19 (2H, s) in the A-ring. The H-  $\alpha$  ( $\delta$  2.99 2H, t, *J*= 7.3 Hz) and H-  $\beta$  ( $\delta$  2.85 2H, t, *J*= 7.3 Hz) of this compound indicated that **4** was a dihydrochalcone. The coupling constant of the doublet for H-1" of **4** in the <sup>1</sup>H-NMR spectrum (*J*= 7.5 Hz) indicated that it has one  $\beta$ -D-glucose.



Chart 1 The structures of 1 and 3

Figure 1 Partial Coherence of 4 in HMBC

	1	3
Aglycone		
2	147.3	147.6
3	185.2	185.5
4	115.2 7.13 d J=8.5	115.7 7.24 d <i>J</i> =8.5
5	113.5 6.97 d J=8.5	113.8 7.07 d J=8.5
6	153.2	153.6
7	134.8	134.7
8	156.2	156.4
9	119.2	119.3
10	115.5 6.67 s	115.7 6.76 s
1'	125.3	125.5
2'	119.2 7.46 d <i>J</i> =1.8	119.4 7.55 d <i>J</i> =1.8
3'	146.5	146.8
4'	149.5	149.7
5'	116.6 6.76 d <i>J</i> =8.3	116.8 6.86 d J=8.2
6'	126.6 7.26 dd <i>J</i> =8.3, 1.8	126.8 7.35 dd <i>J</i> =8.2, 1.8
Glucose		
1"	102.7 5.01 d J=8.0	103.2 4.99 d <i>J</i> =7.6
2"	<b>73.0</b> 3.64 dd <i>J</i> =9.3, 8.0	74.8
3"	<b>78.0</b> 5.00 dd <i>J</i> =9.6, 9.3	77.5
4"	<b>69.7</b> 3.48 dd <i>J</i> =9.6, 9.6	71.6
5"	75.4 3.70-3.73 m	75.8
C"	64.2 4 19 dd L 11 0 5 2	646 426 dd L110 67
0	$\begin{array}{c} 4.16 \text{ uu } J = 11.9,  3.3 \\ 4.24 \text{ hr} \text{ d } L = 11.0 \end{array}$	4.20  du  J = 11.9, 0.7
Acetyl	4.54 Dr.u $J=11.9$	4.45 du <i>J</i> =11.9, 2.5
recetyr		
	172.2	172.6
	172.1	
	20.8 2.05 s	20.7 2.06 s
	21.1 1.97 s	

 Table 1.
 <sup>1</sup>H- and
 <sup>13</sup>C-NMR Spectral Data of 1 and 3 a)

a) Chemical shifts are in  $\delta$  -values from TMS and are followed by multiplicities and *J*-values (in Hz), 25°C, in CD<sub>3</sub>OD (500 MHz)

The singals at  $\delta$  3.81 and 3.71 ( $\times$  2) were consistent with three methoxyl groups attached to the A-ring. The <sup>1</sup>H-<sup>13</sup>C heteronuclear multiple bond coherence (HMBC) (between Glc-1"-H and 4-C) spectra revealed the glucose was connected at the 4-OH of **2** (Figure 1). The three methoxyl groups were connected at 2', 4', 6' on A-ring of **2**. The <sup>1</sup>H-<sup>1</sup>H and <sup>1</sup>H-<sup>13</sup>C COSY spectra were utilized to assign all carbon and proton signals. Thus, **2** was established as 2', 4', 6'-trimethoxy-4-O- $\beta$ -D-glucopyranosyldihydrochalcone. **EXPERIMENTAL** 

**General procedures** All melting points were determined on a Yanagimoto melting point apparatus and were uncorrected. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were measured with a JEOL JNM-LA 500, BM 400, and JNM-EX 270 spectrometer. FAB-MS spectra were measured on a JEOL JMS-DX 302 mass spectrometer. Optical rotations were determined in MeOH on a JASCO DIP-140 polarimeter. Preparative HPLC was performed on a Hitachi (L-6000 pump) instrument with a Waters 5 C<sub>18</sub>-AR-II column (10×250 mm) and Waters 5 SL (10×250 mm) using RI ERC-7520 and UV SSC-5200.

**Plant Material** The aerial part of *Bidens bipinnata* was collected at the wild field of Heilongjiang, in China and was identified by Prof. Gui-Jun Zhang and a voucher specimen has been deposited at the Chinese Medicine Museum of Heilongjiang University of Traditional Chinese Medicine, Harbin, China .

**Extraction and Isolation** The air-dried aerial part (4 kg) of *Bidens bipinnata* was extracted with hot EtOH (10L) twice for 2 h. at 60-70 °C and the combined extract was concentrated *in vacuo* to a syrup, followed by suspension in water. The suspension was extracted with *n*-hexane, ethyl acetate and then *n*-butanol, successively. The EtOAc extract (20 g) was chromatographed on silica gel and eluted with *n*-hexane-EtOAc (4 : 2 to 4 : 6, gradient elute), to give 12 fractions (Fr. 1-12). Fraction No.9 (1.5 g) was subjected to silica gel column, eluted with *n*-hexane-EtOAc (4 : 6 to 2 : 8), to give 4 fractions. Fraction No.3 (0.6 g) was subjected to reversed-phase (ODS) column chromatography (eluting with 50 % aqueous MeOH), followed by preparative HPLC on silica gel column (EtOAc-Me<sub>2</sub>CO-H<sub>2</sub>O 6:0.5:0.1) to afford **1** (6.3 mg) and **4** (7.6 mg). The butanol extract (50 g) was chromatographed on silica gel and eluted successively with EtOAc-MeOH gradient elute, to give 10 fractions (Fr. 1-10). Fraction No.4 (2.5 g) was subjected to reversed-phase (ODS) column chromatography (eluting with 40 % aqueous MeOH) followed by preparative HPLC (40% aqueous MeOH) to afford **2** (5 mg), **3** (8 mg), **5** (16 mg), **6** (6 mg) and **7** (9 mg).

Bidenoside A (1): an orange amorphous powder (MeOH), mp 165-167 °C,  $[\alpha]_D^{25}$ -52.5° (*c* 0.21, MeOH).UV  $\lambda^{\text{MeOH}}$ max nm(log  $\varepsilon$ ): 408 (4.12), 336 (4.04), 271 (3.91). FAB-MS (pos) *m/z*: 533 [M+1]<sup>+</sup>. HR-FAB-MS: C<sub>25</sub>H<sub>25</sub>O<sub>13</sub> (M+1; Calcd for 533.1294; Found: 533.1271). <sup>1</sup>H and <sup>13</sup>C-NMR: given in Table 1.

Bidenoside B (4): a light yellow powder (MeOH), mp 196-198°C,  $[\alpha]_D^{25}$  -130.2° (*c* 0.21, MeOH). UV  $\lambda$  <sup>MeOH</sup>max nm(log  $\varepsilon$ ) : 285 (4.12), 230 (4.31). FAB-MS (pos) *m/z*: 479 [M+1]<sup>+</sup>. HR-EI-MS: C<sub>24</sub>H<sub>30</sub>O<sub>10</sub>

(Calcd for 478.1838; Found: 478.1824) <sup>1</sup>H-NMR (CD<sub>3</sub>OD) : 2.99 (2H, t, J= 7.3 Hz, H- $\alpha$ ), 2.85 (2H, t, J= 7.3 Hz, H- $\beta$ ), 6.98 (2H, d, J= 8.7 Hz, H-2, 6), 7.08 (2H, d, J= 8.7 Hz, H-3, 5), 6.19 (2H, brs, H-3', 5'), 3.71 (6H, s, H-2', 6' OCH<sub>3</sub>), 3.81 (3H, s, H-4' OCH<sub>3</sub>), 4.83 (1H, d, J= 7.5 Hz, H-1"), 3.68 (1H, dd, J= 12.0, 5.2 Hz, H-6"a), 3.87 (1H, dd, J= 12.0, 2.0 Hz, H-6"b). <sup>13</sup>C-NMR (CD<sub>3</sub>OD): 136.1 (C-1), 130.3 (C-2), 117.7 (C-3), 157.4 (C-4), 117.7 (C-5), 130.3 (C-6), 206.3 (C=O), 47.5 ( $\alpha$ -C), 30.3 ( $\beta$ -C), 114.0 (C-1'), 159.8 (C-2', 6'), 91.8 (C-3', 5'), 164.4 (C-4'), Glucose 102.6 (C-1"), 75.0 (C-2"), 78.1 (C-3"), 71.4 (C-4"), 78.0 (C-5"), 62.6 (C-6").

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