SYNTHESIS OF (+)-1-DEOXYGALACTONOJIRIMYCIN[#]

Sung-Jae Pyun, Kee-Young Lee, Chang-Young Oh, and Won-Hun Ham*

College of Pharmacy, SungKyunKwan University, Suwon, 440-746, Korea

whham@skku.edu

([#] Dedicated to Professor Leo A. Paquette on the occasion of his 70th birthday.)

Abstract - A total synthesis of (+)-1-deoxygalactonojirimycin has been accomplished using stereoselective dihydroxylation and piperidine formation by catalytic hydrogenation.

A number of hydroxylated piperidines and pyrrolines occur in nature in plants and microorganisms.¹ These natural products, which have been called "sugar-shaped" alkaloids from plants,² are reversible, competitive inhibitors of glycosidases. The purpose of these natural products is possibly to inhibit the carbohydrate metabolism and consequently the growth of plant consuming pests. Since selective glycosidase inhibitors have a large number of interesting potential applications including treatment of AIDS,³ diabets,⁴ and anticancer agents,⁵ they have a considerable attention.

The "sugar-shaped" alkaloids (aza-sugars) closely resemble monosaccharides by being analogues of these where the ring oxygen has been exchanged with a nitrogen atom. Thus nojirimycin $(1)^6$ and 1-deoxy-nojirimycin $(2)^7$ are analogues of D-glucose and they are glucosidase inhibitors. Similarly natural (+)-galactonojirimycin (3) and its reduction product (+)-1-deoxygalactonojirimycin (4) have been shown to display strong inhibitory activity toward several β -galactosidases (Figure 1).⁸

In this report, we describe the stereoselective synthesis of (+)-1- deoxygalactonojirimycin by utilizing a *trans*-oxazoline (8) as a chiral building block. Our retrosynthetic analysis is shown in Scheme 1. Deoxy-azasugar (4) may be synthesized by hydrogenation and reduction of diol compound (11) which would be prepared by dihydroxylation of α , β -unsaturated ester (10). And α , β -unsaturated ester (10) may

Figure 1



be synthesized by ring cleavage of *trans*-oxazoline (8) in Schotten-Baumann conditions, ozonolysis and Horner-Wadsworth-Emmons reaction.





In our previous report, ⁹ we showed that the palladium(0)-catalyzed oxazoline formation of homoallyl benzamide coming from protected D-serinol might proceed with high stereoselectivity.

The synthesis of (+)-1-deoxygalactonojirimycin (4) began with D-*N*-benzoyl serinol (5) as shown in Scheme 2. Oxidation of alcohol (5) with Dess-Martin periodinane gave the corresponding aldehyde without racemization,¹⁰ which was reacted with vinylmagnesium bromide in THF at 0 °C to afford the corresponding allyl alcohol (6) as the *ca*. 1.1 : 1 mixture of *syn/anti* isomer (¹H NMR analysis) in 75% yield.¹¹ Acetylation of the hydoxyl group yielded the secondary allylic acetate (7). The standard oxazoline ring formation reaction (Pd(PPh₃)₄, K₂CO₃, in MeCN) of secondary allylic acetate (7) gave the desired *trans*-oxazoline (8) in good yield (75%) as a single isomer.

Scheme 2



Reagents and conditions; a) Dess-Martin reagent, CH₂Cl₂, rt.; b) Vinylmagnesium bromide, THF, 0 °C, 75% for two steps; c) Ac₂O, Pyr, DMAP, CH₂Cl₂, rt, 99%; d) Pd(Ph₃P)₄, K₂CO₃, MeCN, 55 °C, 75%

The *trans*-oxazoline (**8**) was treated with benzyl chloroformate in the presence of aqueous sodium bicarbonate (Schotten-Baumann conditions),¹² affording the carbamate (**9**) in 86% yield which was converted into the α,β -unsaturated methyl ester (**10**) in 87% yield in two steps *via* ozonolysis and subsequent Horner-Wadsworth-Emmons reaction. The dihydroxylation¹³ of **10** using a catalytic amount of osmium tetroxide with 1.5 equiv. of *N*-methylmorpholine *N*-oxide as reoxidant (OsO₄/NMO) in CH₂Cl₂ afforded *anti*-diastereomer (**11**) in 80 % yield (> 20 : 1 ¹H NMR analysis). Hydrogenolysis of **11** with 10% Pd(OH)₂ in AcOH/MeOH (1 : 10) was performed under 70 psi H₂ at ambient temperature. Under this condition, we achieved the cyclization of **11** in 62% yield. Reduction of **12** using borane-methyl sulfide complex and deprotection with 6N hydrochlroric acid led to the formation of (+)-1-deoxygalactonojirimycin (**4**) in 68% yield. The optical rotation and spectral data (¹H NMR, ¹³C NMR and HRMS spectrometry) of synthetic **4** were consistent with the reported values (Scheme 3).¹⁴ In summary, we report a new asymmetric synthetic method for the (+)-1-deoxygalactonojirimycin from *trans*-oxazoline (**8**), which was achieved in 25% yield over 8 steps. The key features in this strategy are the diastereoselective oxazoline formation to catalyzed by palladium(0), stereoselective dihydroxylation and piperidine formation by catalytic hydrogenation.

EXPERIMENTAL

(-)-N-[(R)-2-Hydroxy-1-(*tert*-butyldimethylsilanyloxymethyl)but-3-enyl]benzamide (6).

Scheme 3



Reagents and conditions; a) CbzCl,NaHCO₃, CH₂Cl₂/H₂O, rt, 85%; b) O₃, MeOH, -78°C; c) (MeO)₂POCH₂CO₂Me, LiCl, *i*Pr₂NEt, MeCN, 86% for two steps; d) OsO₄, NMO, MC, 80% (> 20 : 1); e) Pd(OH)₂, H₂, MeOH/AcOH (10 : 1), 62%; f) BMS then NaOH, H₂O₂, THF, reflux; g) 6N HCl; h) DOWEX-50, 68% for three steps

To a stirred solution of 5 (7.75 g, 25.05 mmol) in 50 mL of CH₂Cl₂ at 0 °C under argon was added Dess-Martin periodinane (15.8 g, 37.58 mmol), and stirring was allowed to continue for 2 h. The reaction mixture was diluted with Et₂O (50 mL) followed by washing with saturated aqueous NaHCO₃ solution (50 mL \times 2), brine (50 mL \times 2), dried with MgSO₄, and evaporated in *vacuo*. The crude aldehyde was immediately employed in the next step without further purification. To a stirred solution of crude aldehyde in THF (70 mL) at 0 °C, added with a double-tipped needle to a rt solution (125 mL, 1.0 M, 125 mmol) of vinylmagnesium bromide in THF. After being stirred for 1 h, the reaction mixture was washed with saturated aqueous NH₄Cl (100 mL \times 2), brine (100 mL \times 2), dried with MgSO₄, and evaporated in *vacuo*. Purification by silica gel chromatography (ethyl acetate/hexane = 1/10) gave 6 (6.54 g, 75%); colorless oil; [α]²⁴_D -8.20 ° (c 1.0, CHCl₃); IR (neat) 3427, 2930, 2857, 1643 cm⁻¹; ¹H NMR (500 MHz, $CDCl_3$) δ 0.058 (s, 3H), 0.096 (s, 3H), 0.91 (s, 9H), 3.62 (s, 1H), 3.99 - 4.01 (dd, J = 2.5 and 4.0 Hz, 2H), 4.16 (m, J = 2.5, 4.0 and 7.5 Hz, 1H), 4.64 (m, 1H), 5.21 (dt, J = 1.5 and 11.0 Hz, 1H), 5.39 (dt, J = 1.5 and 15.5 Hz, 1H), 5.88 (ddd, J = 6.0, 11.0 and 15.5 Hz, 1H), 6.79 (d, J = 7.5 Hz, 1H), 7.43 - 7.52 (m, 3H), 7.76 - 7.78 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ -5.33, 18.37, 26.04, 53.74, 65.36, 73.83, 116.47, 127.15, 128.86, 131.84, 134.60, 137.54, 167.91; HRMS (EI, 70eV) calcd for C₁₈H₂₉NO₃Si 335.1917, found 335.1917.

(-)-N-((1R, 2R)-2-Acetoxy-1-(tert-butyldimethylsilanyloxymethyl)-3-butenyl)benzamide (7).

To a stirred solution of **6** (3.5 g, 10.4 mmol) in CH₂Cl₂ (40 mL) were added acetic anhydride (1.1 mL, 14.2 mmol), pyridine (0.93 mL, 14.2 mmol), and stirring was allowed to continue for 12 h. The reaction mixture was washed with 1N HCl (20 mL × 2), saturated aqueous NaHCO₃ solution (20 mL × 2), brine (20 mL × 2), dried with MgSO₄, and evaporated in *vacuo*. Purification by silica gel chromatography (ethyl acetate/hexane = 1/15) gave **7** (3.9 g, 99%); colorless oil; $[\alpha]^{25}_{D}$ -8.4 ° (c 1.0, CHCl₃); IR (neat) 3322, 2931, 2857, 1745, 1646 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.030 (s, 3H), 0.054 (s, 3H), 0.91 (s, 9H), 1.58 (s, 3H), 3.69 (dd, *J* = 3.0, 10.0 Hz, 1H), 3.82 (dd, *J* = 5.5, 10.0 Hz, 1H), 4.39 (m, 1H), 5.31 (dt, *J* = 1.0, 11.0 Hz, 1H), 5.37 (dt, *J* = 1.0, 17.5 Hz, 1H), 5.67 (m, 1H), 5.88 (ddd, *J* = 6.5, 11.0, 17.5 Hz, 1H), 6.49 (d, *J* = 9.5 Hz, 1H), 7.43–7.52 (m, 3H), 7.74–7.75 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ -5.35, 18.42, 21.34, 26.03, 53.55, 62.00, 73.86, 119.41, 127.07, 128.93, 131.85, 133.71, 134.61, 167.25, 170.63; HRMS (EI, 70 eV) calcd for C₂₀H₃₁NO₄Si 377.2022, found 377.2027.

(4R, trans)-4,5-Dihydro-4-(tert-butyldimethylsilanyloxymethyl)-2-phenyloxazoline (8).

To a stirred solution of secondary allylic acetate (**7**) (4.0 g, 10.6 mmol) and K₂CO₃ (4.4 g, 31.78 mmol) in MeCN (50 mL) was added Pd(PPh₃)₄ (61.2 mg, 0.53 mmol) under N₂. The resulting mixture was heated under reflux for 24 h, whereupon it was allowed to cool rt and was filtered through a pad of silica, which was then evaporated under reduced pressure to give crude product. Purification by silica gel chromatography (ethyl acetate/hexane = 1/15) gave **8** (2.52 g, 75%); colorless oil; $[\alpha]^{25}_{D}$ +3.66 ° (c 1.0, CHCl₃); IR (neat) 2930, 2857, 1650 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.038 (s. 3H), 0.075 (s, 3H), 0.86 (s, 9H), 3.69 (dd, *J* = 7.0, 10.0 Hz, 1H), 3.94 (dd, *J* = 4.0, 10.0 Hz, 1H), 4.06 (ddd, *J* = 4.0, 5.5, 7.0 Hz, 1H), 5.02 (dd, *J* = 5.5, 6.5 Hz, 1H), 5.21 (dt, *J* = 1.5, 10.5 Hz, 1H), 5.36–5.40 (dt, *J* = 1.5, 17.0 Hz, 1H), 5.96 (ddd, *J* = 6.5, 10.5, 17.0 Hz, 1H), 7.39–7.48 (m, 3H), 7.95–7.97 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ -5.08, 26.04, 64.95, 74.33, 83.31, 116.62, 128.02, 128.54, 131.59, 137.00, 164.11; HRMS m/e calcd for C₁₈H₂₇NO₂Si (M+1) 318.1889, found 318.1902.

1(*R*)-[2-(*tert*-Butyldimethylsiloxy)-1(*R*)-[[*N*-(phenylmethoxy)carbonyl]amino]ethyl]prop-2-enyl benzoate (9).

To a solution of oxazoline (8) (3.32 g, 10.47 mmol) in CH_2Cl_2 (20 mL) was added a solution of NaHCO₃ (3.52g, 41.90 mmol) in water (20 mL), and the mixture was cooled in an ice bath. To this solution was added dropwise a solution of benzyl chloroformate (3.0 mL, 21.01 mmol). The mixture was stirred at rt for 4 h. The organic phase was separated, and the aqueous phase was extracted with CH_2Cl_2 (2 × 20 mL). The combined organic phase was washed with water, dried (MgSO₄), and concentrated in *vacuo*. Purification by silica gel chromatography (ethyl acetate/hexane = 1/4) gave **9** (4.18g, 85 %) as a colorless

oil; $[\alpha]^{23}_{D}$ +4.2 ° (c 1.0, CH₂Cl₂); IR (neat) : 3316, 3012, 2941, 1720, 1615 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) : δ 0.03 (3H, s) 0.06 (3H, s), 0.90 (9H, s), 3.67 (1H, dd, *J* = 10, 5.5 Hz), 3.80 (1H, dd, *J* = 10, 3.5 Hz), 4.06 (1H, m), 5.04 (2H, s), 5.14 (1H, d, *J* = 10.5 Hz), 5.31 (1H, d, *J* = 10.5 Hz), 5.42 (1H, d, *J* = 17 Hz), 5.73 (1H, m), 5.92 (1H, m), 7.25 (5H, m), 7.43 (2H, m), 7.56 (1H, m), 8.04 (2H, m). ¹³C NMR (125 MHz, CDCl₃) : δ -4.88, 18.91, 26.53, 55.71, 62.81, 67.54, 74.80, 119.94, 128.76, 128.84, 129.13, 129.18, 130.42, 130.74, 133.80, 134.15, 137.03, 156.90, 166.26; HRMS calcd for C₂₆H₃₅NO₅Si (M+1) 470.2363, Found 470.2354.

Methyl 4(*R*)-benzoxy-6-(*tert*-butyldimethylsiloxy)-5(*R*)-[[*N*-(phenylmethoxy)carbonyl]amino]hex-2(*E*)-enoate (10).

Compound (9) (2.0 g, 4.26 mmol) was dissolved in dry methanol (50 mL) and cooled to -78 °C. Pass ozonied oxygen until the reaction is complete. The reaction mixture was evaporated under reduced pressure. The crude aldehyde was immediately employed in the next step without further purification.

To a stirred solution of LiCl (217 mg, 5.11 mmol) in MeCN (80 mL) were added trimethylphosphonoacetate (0.81 mL, 5.11 mmol) and *i*Pr₂NEt (0.89 mL, 5.11 mmol) and stirring was allowed to continue for 1 h. To this solution was added dropwise a solution of crude aldehyde (4.3 mmol) in MeCN (20 mL) and the reaction mixture was stirred for 3 h. The reaction mixture was poured into H₂O (30 mL), extracted with EtOAc (20 mL × 2). The organic extract was washed with brine (20 mL), dried with MgSO₄, and evaporated in *vacuo*. Purification by silica gel chromatography (ethyl acetate/hexane = 1/2) gave α,β -unsaturated ester (**10**) (1.95 g, 87 % for 2 steps) as a colorless oil; $[\alpha]^{23}_{D}$ +18.5 ° (c 0.7, CH₂Cl₂); IR (neat) : 3348, 3018, 2950, 1721, 1603, 1100 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) : δ 0.03 (6H, s), 0.87 (9H, s), 3.66 (1H, dd, *J* = 10, 5.5 Hz), 3.74 (3H, s), 3.82(1H, dd, *J* = 10, 3 Hz), 5.06 (2H, s), 5.17 (1H, d, *J* = 10 Hz), 5.91 (1H, m), 6.07 (1H, d, *J* = 16 Hz), 7.01 (1H, dd, *J* = 16, 5.5 Hz), 7.28 (5H, m), 7.45 (2H, m), 7.58 (1H, m), 8.04 (2H, m); ¹³H NMR (125 MHz, CDCl₃) : δ -4.95, -4.87, 14.91, 18.87, 21.75, 26.49, 52.44, 55.44, 62.64, 67.74, 72.65, 123.91, 128.64, 128.83, 128.91, 129.23, 129.26, 129.74, 130.06, 130.38, 130.48, 134.16, 136.84, 143.28, 156.67, 165.86, 166.67; HRMS calcd for C₂₈H₃₇NO₇Si (M+1) 528.2418, Found 528.2423.

(2R,3R,4S,6R)-4-Benzoxy-6-(*tert*-butyldimethylsiloxy)-5-[[N-(phenylmethoxy)carbonyl]amino]hexanoic Acid Methyl Ester 2,3,4,6-Tetrol (11)

To a stirred solution of **10** (500 mg, 0.95 mmol) in CH_2Cl_2 (20 mL) were added a *N*-methylmorpholine *N*-oxide (166.5 mg, 1.43 mmol) and the solution of 4 wt. % OsO_4 /water (0.12 mL, 0.019 mmol, 2 mol %) and the reaction was allowed to stir for 12 h, at which time all staring material had been consumed as judged by TLC. The reaction mixture was poured into a solution of 15% Na₂SO₃ (50 mL), extracted with

CH₂Cl₂ (20 mL × 2). The organic extract was washed with brine (20 mL), dried with MgSO₄, and evaporated in *vacuo*. Purification by silica gel chromatography (ethyl acetate/hexane = 1/2) gave **11** (426 mg, 80 %, > 20 : 1) as a colorless oil; IR (neat) 3435, 2954, 2857, 1728, 1602, 1509, 113 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) : δ -0.10 (6H, s), 0.86 (9H, s), 3.79 (3H, s), 3.80 (1H, d, *J* = 5 Hz), 4.13 (1H, m), 4.17 (1H, d, *J* = 6 Hz), 4.30 (1H, m), 4.37 (1H, d, *J* = 6 Hz), 5.19(2H, m), 5.38 (1H, d, *J* = 9.5 Hz), 5.55 (1H, dd, *J* = 9 Hz, 1.5 Hz), 7.37 (5H, m), 7.43 (2H, m), 7.56 (1H, m), 8.03 (2H, m); ¹³C NMR (125 MHz, CDCl₃) : δ -4.98, 14.89, 18.84, 21.74, 26.44, 52.75, 54.40, 61.10, 64.04, 68.21, 71.21, 73.14, 128.74, 128.86, 129.06, 129.17, 129.31, 130.15, 130.55, 130.63, 130.74, 134.13, 136.71, 158.43, 165.92, 173.81; HRMS calcd for C₂₈H₃₉NO₉Si (M+1) 562.2472, Found 562.2473.

(3R, 4R, 5S, 6R)-5-Benzoxy-6-(*tert*-butyldimethylsilyloxymethyl)-3,4-dihydroxypiperidin-2-one (12)

To a solution of **11** (300 mg, 0.53 mmol) in 20 mL of 1:9 AcOH/MeOH were added 300 mg 10% Pd(OH)₂ and the reaction mixture was vigorously shaken under 70 psi H₂ for 10 h at rt. The mixture was then filtered and concentrated *in vacuo*. Purification by column chromatography (ethyl acetate/hexane = 4/1) over silica gel gave lactam (**12**) (131 mg, 62%) as a white solid; mp 72-73 °C; $[\alpha]^{23}_{D}$ +69.79 ° (c 0.95, CH₂Cl₂); IR (KBr) : 3392, 2931, 2857, 1726, 1671, 1468, 1268, 1110 cm⁻¹;¹H NMR (500 MHz, CDCl₃) : δ 0.00 (6H, s), 0.85 (9H, s), 3.52 (1H, m), 3.77 (2H, m), 4.17 (1H, dd, *J* = 10, 2 Hz), 4.44 (1H, d, *J* = 10 Hz), 5.83 (1H, s), 6.40 (1H, s), 7.46 (2H, m), 7.59 (1H, m), 8.01 (2H, m); ¹³C NMR (125 MHz, CDCl₃) : δ -4.84, 14.88, 18.80, 21.74, 26.45, 55.61, 63.30, 69.93, 70.84, 72.19, 129.26, 129.78, 130.59, 134.23, 166.57, 172.96; HRMS calcd for C₁₉H₂₉NO₆Si (M+1) 396.1842, Found 396.1842.

(+)-1-Deoxygalactonojirimycin (4).

Borane-methyl sulfide complex (0.175 mL of a 10.1M solution in THF, 1.77 mmol) was added to a cooled (0 °C) solution of the lactam (**12**) (100 mg, 0.25 mmol) in THF (5 mL). After 30 min, the solution was warmed to rt. After another 2 h, the reaction was quenched by the slow addition of 3N NaOH and 30% H₂O₂ and heated at reflux for 1 h. After cooling to rt, to this solution was added excess 6N HCl (5 mL) and stirred for 3 h and the mixture concentrated in *vacuo*, and the residue was purified by using ion-exchange chromatography (Dowex-50, H⁺) eluting with 3% NH₄OH. Subsequent evaporation of water in vacuo below 40 °C afforded a colorless syrup, which was dissolved in a small amount of methanol and then precipitated by addition of acetone to give a hygroscopic, amorphous solid of (+)-1-deoxygalactonojirimycin (**4**) (28 mg, 68%); $[\alpha]^{25}_{D}+52.5$ ° (c 1.0. H₂O) [lit.,¹³ [α]²³_D+52.8 ° (c 1.0. H₂O)]; IR(neat) 3448 cm⁻¹; ¹H NMR (500 MHz, D₂O) : δ 2.41 (1H, dd, *J* = 12.5, 11.0 Hz), 2.77 (1H, dd, *J* = 12.5, 6.5 Hz), 3.16 (1H, dd, *J* = 12.5, 5.5 Hz), 3.51(1H, dd, *J* = 9.7, 3.0 Hz), 3.65 (1H, dd, *J* = 11, 6.5 Hz), 3.77 (1H, dt, *J* = 11, 5.5 Hz), 4.03 (1H, dd, *J* = 3.2, 1.5 Hz); ¹³C NMR

(125 MHz, D₂O with CH₃CN as internal standard) δ 49.70, 59.43, 62.04, 68.83, 69.91, 75.70; HRMS calcd for C₆H₁₃NO₄ (M+1) 164.0923, Found 164.0924

ACKNOWLEDGEMENTS

This work was supported by grant No. R01-2001-000-00214-0 from Korea Science & Engineering Foundation.

REFERENCES

- 1. L. E. Fellows, New Scientist, 1989, 26, 45.
- 2. L. E. Fellows, Chem. Brit., 1987, 842.
- A. Karpas, G. W. J. Fleet, R. A. Dwek, L. E. Fellows, A. S. Tyms, S. Petursson, S. K. Namgoo ng, N. G. Ramsden, G. S. Jacob, and T. W. Rademacher, *Proc. Natl. Acad. Sci. U. S. A.*, 1988, 85, 9229.
- P. S. Johnston, H. E. Lebovitz, R. F. Coniff, D. C. Simonson, P. Raskin, and C. L. Munera, J. Clin. Endocrinol. Metab., 1998, 83, 1515.
- 5. T. Tsuruoka, H. Fukuyasu, M. Ishii, T. Usui, S. Shibahara, and S. Inouye, J. Antibiot., 1996, 49, 155.
- (a) S. Inouye, T. Tsuruoka, and Y. J. Niida, *Antibiot.*, 1966, **19**, 288. (b) S. Inouye, T. Tsuruo ka, T. Ito, and T. Niida, *Tetrahedron*, 1968, **24**, 2125.
- 7. S. Murao and S. Miyata, Agric. Biol. Chem., 1980, 44, 219.
- (a) Y. Miyake and M. Ebata, Agric. Biol. Chem., 1988, 52, 153.
 (b) Y. Miyake and M. Ebata, Agric. Biol. Chem., 1988, 52, 1649.
- (a) K.-Y. Lee, Y.-H. Kim, M.-S. Park, and W.-H. Ham, *Tetrahedron Lett.*, 1998, **39**, 8129. (b) K.-Y. Lee, Y.-H. Kim, M.-S. Park, C.-Y. Oh, and W.-H. Ham, *J. Org. Chem.*, 1999, **64**, 9450. (c) K.-Y. Lee, Y.-H. Kim, C.-Y. Oh, and W.-H. Ham, *Org. Lett.*, 2000, **2**, 4041. (d) K.-Y. Lee, C.-Y. Oh, and W.-H. Ham, *Org. Lett.*, 2002, **4**, 4403. (e) K.-Y. Lee, C.-Y. Oh, Y.-H. Kim, J.-E. Joo, and W.-H. Ham, *Tetrahedron Lett.*, 2002, **43**, 9361. (f) Y.-S. Lee, Y.-H Shin, Y.-H. Kim, K.-Y. Lee, C.-Y. Oh, S.-J. Pyun, H.-J. Park, J.-H. Jeong, and W.-H. Ham, *Tetrahedron:Asymmetry*, 2003, **14**, 87.
- 10. (a) D. B. Dess and J. C. Martin, J. Org. Chem., 1983, 48, 4155. (b) A. G. Meyers, B. Zhong,
 M. Movassaghi, D. W. Kung, B. A. Lanman, and S. Kwon, *Tetrahedron Lett.*, 2000, 41, 1359.
- 11. J.-N. Denis, A. Correa, and A. E. Greene, J. Org. Chem., 1991, 56, 6939.
- 12. C. Kouklovsky, A. Pouilhes, and Y. Langlois, J. Am. Chem. Soc., 1990, 112, 6672.

- 13. (a) J. K. Cha, W. J. Christ, and Y. Kishi, *Tetrahedron Lett.*, 1983, 24, 3943.
 For a recent review, see: (b) J. K. Cha and N.-S. Kim, *Chem. Rev.*, 1995, 95, 1761; (c) V. VanRheenen, D. Y. Cha, and W. M. Hartley, *Organic Syntheses*, Wiley & Sons: New York, 1988; Coll. Vol., VI, p. 342.
- 14. Y. Miyake and M. Ebata, Agric. Biol. Chem., 1988, 52, 661.