

A STEREOCONTROLLED SYNTHESIS OF HAPALOSIN

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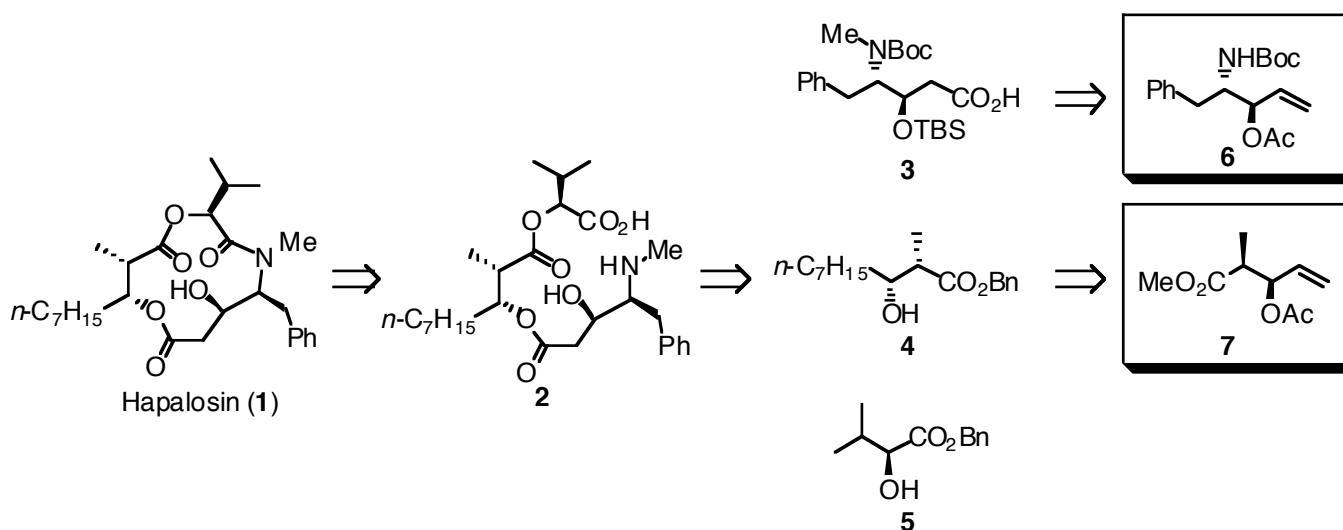
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Abstract – A facile synthetic method for two components of hapalosin, that is, α -hydroxy- β -amino acid and β -hydroxy acid, has been established by utilizing chiral building blocks efficiently resolved in a lipase-catalyzed transesterification. Furthermore, the synthesis of hapalosin through macrolactamization of the seco acid derived from these two components and (*S*)-2-hydroxy-3-methylbutyric acid has thus been demonstrated.

A 12-membered cyclic depsipeptide hapalosin (**1**), isolated by Moore and co-workers, has shown remarkable reversing activity against P-glycoprotein-mediated multidrug resistance (MDR) of cancer cells.¹ Due to the important biological activity and the unique structural features, many research groups have pursued syntheses of **1** and its analogues.²⁻¹¹ Most of them reasonably adopted the macrolactamization of seco acid (**2**) which consists of three components, that is, α -silyloxy- β -amino acid (**3**), α -hydroxy ester (**4**), and β -hydroxy ester (**5**) as depicted in Scheme 1.

Scheme 1

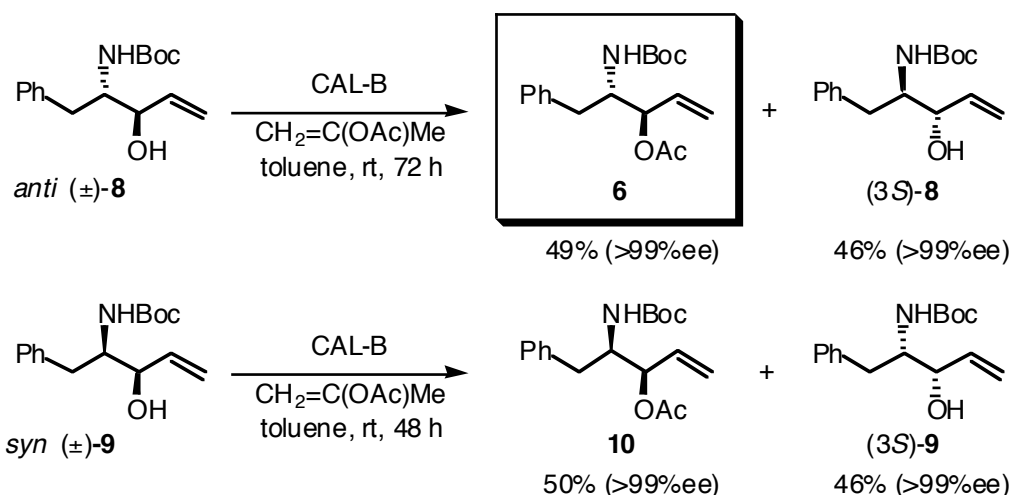


The compound (**3**) was prepared in several ways, some of which started from stereodefined compounds such as L-phenylalanine,²⁻⁶ L-serine,⁷ or the chiral epoxides accessible by the Sharpless' protocols.^{12,13} Others depend on asymmetric reactions using the chiral auxiliaries such as Williams' oxazine¹⁴ and Evans' oxazolidinone.¹⁵ On the other hand, **4** was elaborated by means of either the Cram-selective addition of Grignard reagent to (*R*)-2-phenylpropanal⁷ or the asymmetric reactions involving Evans-type

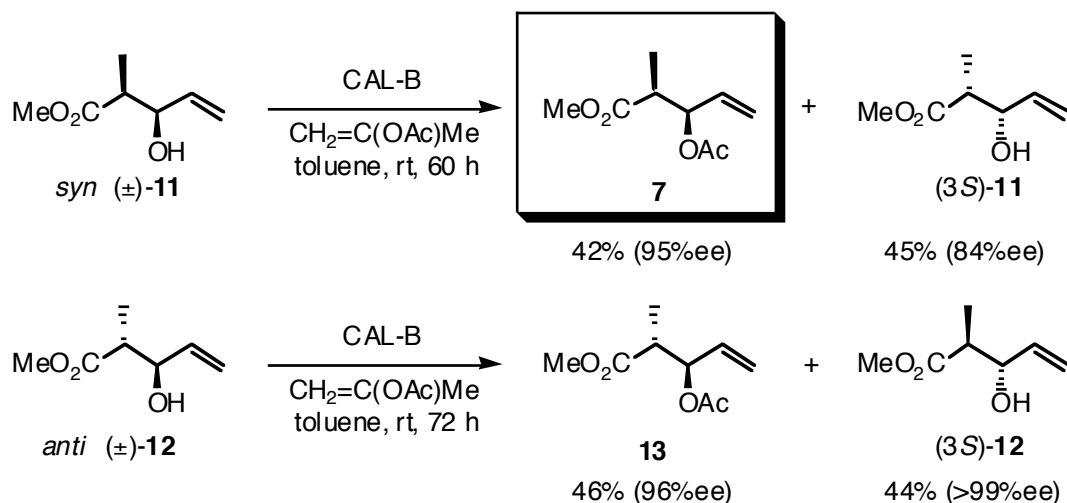
syn-aldol protocol^{3-5,8} and Brown's allylboration of 1-octanal.² These preparative methods for **3** and **4**, however, require careful handling of moisture-sensitive reagents under low temperatures and/or manipulation of easily racemizing compounds. In the light of the important biological activity of **1**, a more intensive structure-activity relationship (SAR) study would be indispensable for further biological information. Thus, it is highly desired to develop a more flexible and concise approach to a wide range of diastereomers, enantiomers, and congeners of **3**, **4** and **5** leading to a variety of analogues of **1**.

The reasonable retrosynthetic analysis of **1** is illustrated in Scheme 1, in which compounds (**6**) and (**7**) seem to be the superb precursors for **3** and **4**, respectively, because **3** can be synthesized through hydroboration/oxidation and **4** through cross-metathesis/hydrogenation. In our efforts to expand the usefulness of lipase-catalyzed kinetic resolutions, we have recently disclosed that methyl 2-substituted 3-hydroxy-4-pentenoates¹⁶ and 4-amino-1-alken-3-ols¹⁷ can efficiently be resolved by use of CAL-B (*Candida antarctica*, fraction B), which seemingly allows access to **6** and **7** with high optical purity together with all the possible stereoisomers. In the event, racemic amino alcohols (**8**),^{18,19} (**9**),^{18,19} racemic \square -hydroxy esters (**11**),^{20,21} and (**12**)^{20,21} were efficiently resolved with high enantiomeric purity as outlined in Schemes 2 and 3.

Scheme 2

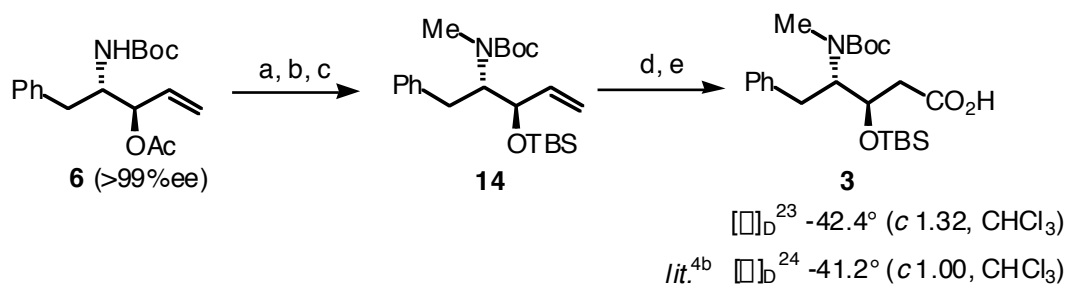


Scheme 3



Encouraged by these successful results, we then examined the transformation of **6** and **7** into **3** and **4**, respectively. As shown in Scheme 4, compound (**6**) was transformed into silyl ether (**14**) through alkaline-hydrolysis followed by silyl protection and *N*-methylation. Hydroboration of **14** followed by TEMPO-mediated oxidation²² gave rise to **3** uneventfully, whose ¹H and ¹³C NMR spectra and $[\alpha]_D$ were in good accordance with those in the literature.^{4b}

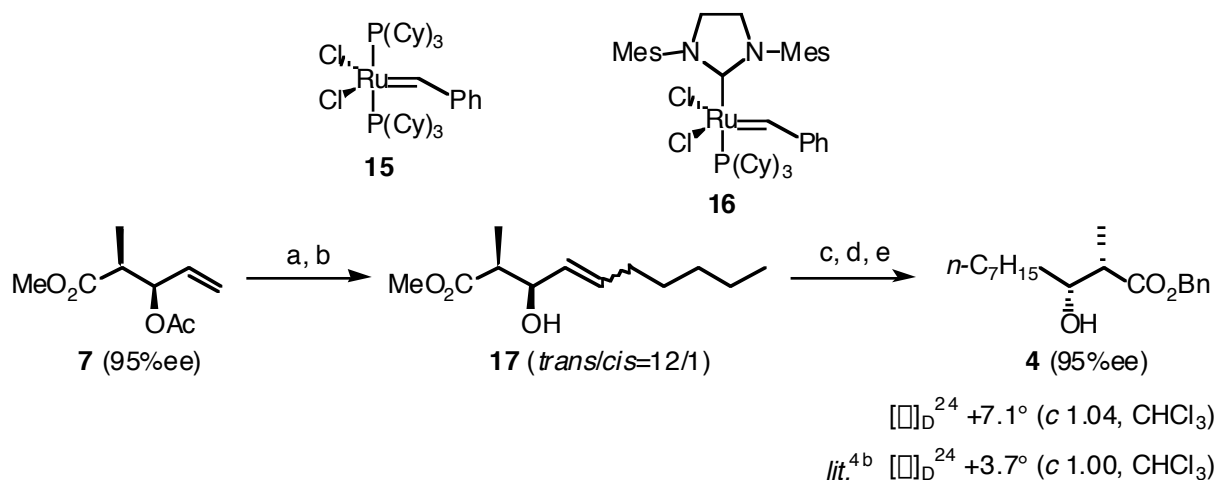
Scheme 4



- (a) 1 M NaOH/MeOH, rt, 1 h, 98%; (b) TBSCl/imidazole/DMF, rt, 10 h, 97%;
 (c) NaH/Mel/DMF, rt, 12 h, 98%; (d) 9-BBN/THF, rt, 12 h; NaOOH, 50 °C, 3 h, 82%;
 (e) cat. TEMPO/cat. NaOCl/NaClO₂/MeCN/pH 6.8 phosphate buffer, rt, 4.5 h, 66%.

On the other hand, compound (**4**) was prepared by the procedure depicted in Scheme 5. The initially attempted cross-metathesis of **7** with 1-heptene (3 eq.) resulted in poor yield (20-40%) even at the refluxing temperature of dichloromethane in the presence of 5 mol% of cata;yst (**15**).²³ However, the reaction with catalyst (**16**)²⁴ smoothly afforded α -hydroxy ester (**17**) even at room temperature. Hydrogenation of **17** followed by alkaline-hydrolysis gave a carboxylic acid which was alkylatively esterified to afford **4** (95%ee),²⁵ whose ¹H and ¹³C NMR spectra were identical to those in the literature.^{4b} This procedure implies the high synthetic potential of **7** leading to a wide range of stereodefined α -hydroxy esters with high enantiomeric purity in place of Evans' asymmetric aldol protocol.

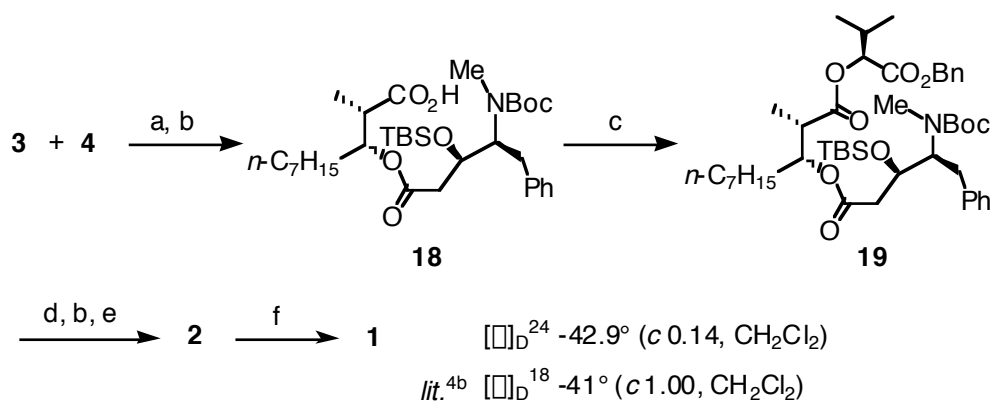
Scheme 5



- (a) K₂CO₃/MeOH, rt, 1 h, 85 %; (b) **16** (5 mol%)/1-heptene (3 eq.)/CH₂Cl₂, rt, 12 h, 71%;
 (c) H₂ (1 atm)/10%Pd-C /EtOH, 2 h, 82%; (d) 1 M NaOH/MeOH, rt, 5 h; (e) Cs₂CO₃/BnBr/DMF, rt, 12 h, 86% in two steps.

With components (**3**) and (**4**) in hand, the stage was set for macrolactamization. As depicted in Scheme 6, **1** was synthesized through the slightly modified procedure of the Nishiyama's protocol.⁴ Condensation of **3** with **4** followed by reductive cleavage of the benzyl ester afforded carboxylic acid (**18**), which was condensed with **5**²⁶ to give fully-protected seco acid (**19**). Sequential treatment with TBAF, H₂/Pd(OH)₂, and TFA gave seco acid (**2**) as a trifluoroacetic acid salt. Finally, the macrolactamization of **2** was accomplished by means of DPPA²⁷ in DMF under high dilution condition to afford **1** in 32% yield, whose ¹H and ¹³C NMR spectra and [α]_D were fully identical to those reported.^{1,2,4,7,8}

Scheme 6



(a) EDCI/DMAP/CH₂Cl₂, 40 °C, 8 h, 89% (b) H₂ (1 atm)/cat. Pd(OH)₂/EtOH, rt, 1 h;
 (c) **5**/EDCI/DMAP/CH₂Cl₂, rt, 12 h, 97%; (d) TBAF/THF, rt, 1.5 h, 77%; (e) TFA (10 eq.)
 /CH₂Cl₂, rt, 2 h, 90%; (f) DPPA (2 eq.)/*i*-Pr₂NEt (6 eq.)/DMF (1 mM soln of **2**), rt, 72 h, 32%.

In summary, we have achieved an efficient synthesis of hapalosin utilizing chiral building blocks obtained by lipase-catalyzed kinetic resolutions of methyl 2-substituted 3-hydroxy-4-pentenoates and 4-amino-1-alken-3-ols. We believe that our strategy constitutes an advantageous route to various congeners of hapalosin required for SAR studies, which will be reported in due course.

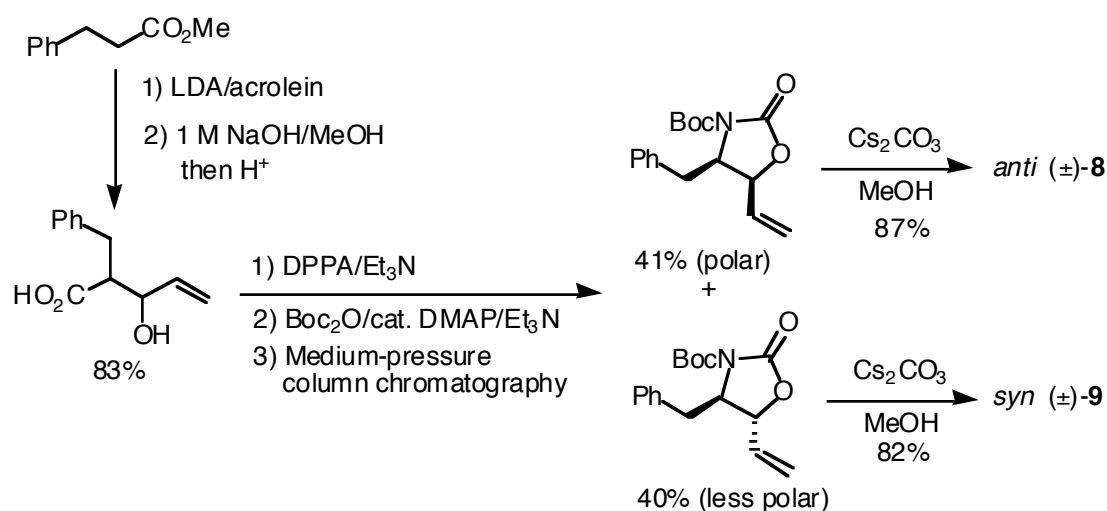
ACKNOWLEDGMENTS

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REFERENCES AND NOTES

1. K. Stratmann, D. L. Burgoyne, R. E. Moore, G. M. L. Patterson, and C. D. Smith, *J. Org. Chem.*, 1994, **59**, 7219.
2. (a) T. Q. Dinh and R. W. Armstrong, *J. Org. Chem.*, 1995, **60**, 8118.
 (b) T. Q. Dinh, X. Du, and R. W. Armstrong, *J. Org. Chem.*, 1996, **61**, 6606.
3. A. K. Ghosh, W. Liu, Y. Xu, and Z. Chen, *Angew. Chem., Int. Ed. Engl.*, 1996, **35**, 74; *Angew. Chem.*, 1996, **108**, 73.
4. (a) K. Ohmori, T. Okuno, S. Nishiyama, and S. Yamamura, *Tetrahedron Lett.*, 1996, **37**, 3467.
 (b) T. Okuno, K. Ohmori, S. Nishiyama, S. Yamamura, K. Nakamura, K. N. Houk, and K. Okamoto, *Tetrahedron*, 1996, **52**, 14723.
5. B. Wagner, R. Beugelmans, and J. Zhu, *Tetrahedron Lett.*, 1996, **37**, 6557.

6. C. E. O'Connell, K. A. Salvato, Z. Meng, B. A. Littlefield, and C. E. Schwartz, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 1541.
7. M. Haddad, C. Botuha, and M. Larchevêque, *Synlett*, 1999, 1118.
8. C. Hermann, G. C. G. Pais, A. Geyer, A. M. Kühnert, and M. E. Maier, *Tetrahedron*, 2000, **56**, 8461.
9. T. Q. Dinh, X. Du, C. D. Smith, and R. W. Armstrong, *J. Org. Chem.*, 1997, **62**, 6773.
10. N. Kashihara, S. To-e, K. Nakamura, K. Umezawa, S. Yamamura, and S. Nishiyama, *Bioorg. Med. Chem. Lett.*, 2000, **10**, 101.
11. For a review, see: S. Nishiyama, *J. Syn. Org. Chem., Japan*, 2001, **59**, 938.
12. M. Catasús, A. Moyano, M. A. Pericás, and A. Riera, *Tetrahedron Lett.*, 1999, **40**, 9309.
13. M. E. Maier and C. Hermann, *Tetrahedron*, 2000, **56**, 557.
14. Y. Aoyagi and R. M. Williams, *Tetrahedron*, 1998, **54**, 10419.
15. G. C. G. Pais and M. E. Maier, *J. Org. Chem.*, 1999, **64**, 4551.
16. T. Mandai, T. Oshitari, and M. Susowake, *Synlett*, 2002, 1665.
17. T. Mandai and T. Oshitari, submitted for publication.
18. Prepared as follows.



For the ring opening with $Cs_2CO_3/MeOH$, see:
T. Ishizuka and T. Kunieda, *Tetrahedron Lett.*, 1987, **28**, 4185.

19. The typical procedure is as follows. A mixture of (\pm)-**8** (3.17 g, 11.4 mmol), 2-propenyl acetate (3.78 mL, 34.2 mmol), and CAL-B [1.14 g, 0.1 g per 1 mmol of (\pm)-**8**] in toluene (34 mL) was stirred at rt for 72 h. The lipase was filtered off and the filtrate was concentrated to give solids which were chromatographed (SiO_2) to afford acetate (**6**) (1.78 g, 49%) and alcohol (3*S*)-(**8**) (1.45 g, 46%). Treatment of **6** with $K_2CO_3/MeOH$ (rt, 40 min) gave rise to alcohol (3*R*)-(**8**) in a quantitative yield. The %ee of (3*R*)-**8** and (3*S*)-**8** was determined by HPLC using Chiralcel OD-H (hexane/2-propanol=20/1, 230 nm). The reaction of (\pm)-**9** was performed in an almost similar manner. Compound **10** was converted to alcohol (3*R*)-(**9**) by the treatment with $K_2CO_3/MeOH$ (rt, 40 min). The %ee of (3*R*)-**9** and (3*S*)-**9** was determined by HPLC using Chiralcel OD-H (hexane/2-propanol=50/1, 254 nm) after conversion to benzoates.
20. Prepared by the aldol condensation of methyl propionate with acrolein (LDA/THF, $-78^\circ C$). The aldols thus obtained revealed to be a *ca.* 1:1 mixture of diastereomers, which were isolated by

medium-pressure column chromatography (SiO₂, hexane/EtOAc=6/1-4/1).

21. The typical procedure is as follows. A mixture of (±)-**11** (4.89 g, 31.8 mmol), 2-propenyl acetate (7.01 mL, 63.6 mmol), and CAL-B [3.18 g, 0.1 g per 1 mmol of (±)-**11**] in toluene (48 mL) was stirred at rt for 60 h. The lipase was filtered off and the filtrate was concentrated to give an oil which was chromatographed (SiO₂) to afford acetate (**7**) (2.50 g, 42%) and alcohol (3*S*)-(**11**) (2.05 g, 45%). The compound (**7**) was treated with K₂CO₃/MeOH (rt, 40 min) to give (3*R*)-**11** in quantitative yield. The %ee of (3*R*)-**11** and (3*S*)-**11** was determined by HPLC using Chiralcel OD-H (hexane/2-propanol=20/1, 230 nm). The reaction of (±) **12** was performed in an almost similar manner. The %ee of (3*R*)-**12** and (3*S*)-**12** was determined by HPLC using Chiralcel OD-H (hexane/2-propanol=20/1, 230 nm).
22. M. Zhao, J. Li, E. Mano, Z. Song, D. M. Tschaen, E. J. J. Grabowski, and P. J. Reider, *J. Org. Chem.*, 1999, **64**, 2564.
23. P. Schwab, R. H. Grubbs, and J. W. Ziller, *J. Am. Chem. Soc.*, 1996, **118**, 100.
24. (a) M. Scholl, S. Ding, C. W. Lee, and R. H. Grubbs, *Org. Lett.* 1999, **1**, 953. (b) J. P. Morgan and R. H. Grubbs, *Org. Lett.*, 2000, **2**, 3153.
25. Determined by HPLC using Chiralcel OD-H (hexane/2-propanol=9/1, 0.8 mL/min, 220 nm). **4**: *t*_R=6.60 min, the enantiomer of **4**: *t*_R=5.85 min.
26. Commercially available from Aldrich. For the preparation, see: (a) K. Mori, *Tetrahedron*, 1976, **32**, 1101. (b) P. Koch, Y. Nakatani, B. Luu, and G. Ourisson, *Bull. Soc. Chim. Fr.*, 1983, **11**, II-189. (c) W.-R. Li, W. R. Ewing, B. D. Harris, and M. M. Joullié, *J. Am. Chem. Soc.*, 1990, **112**, 7659.
27. T. Shioiri, K. Ninomiya, and S. Yamada, *J. Am. Chem. Soc.*, 1972, **94**, 6203.