HETEROCYCLES, Vol. 63, No. 10, 2004, pp. 2211 - 2215 Received, 23rd July, 2004, Accepted, 19th August, 2004, Published online, 23rd August, 2004

ABSOLUTE STEREOSTRUCTURES OF ACYLATED KHELLACTONE-TYPE COUMARINS FROM ANGELICA FURCIJUGA

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Abstract — Two new acylated khellactone-type coumarin hyuganoside I (1) and hyuganin F (2) were isolated from the leaves of Angelica furcijuga. Their absolute stereostructures were determined on the basis of chemical and physicochemical evidence. Furthermore, the absolute stereostructures of the acyl moieties in hyuganins A (3) and C (4) and isoepoxypteryxin (5) were elucidated.

The Umbelliferae plant *Angelica furcijuga* is indigenous to Japan (Japanese name, hyugatouki) and the dried whole plant has been used for the treatment of hepatopathy, allergosis, inflammation, diabetes, and hypertension as a Japanese folk medicine. During the course of our characterization studies on *A. furcijuga*,¹⁻⁴ we have reported the structure elucidation of four acylated khellactone-type coumarins called hyuganins A—D and three glycosides, hyuganosides II, IIIa, and IIIb from the roots and hyuganosides IV and V from the flowers of *A. furcijuga*,²⁻⁴ The principal constituents were found to show nitric oxide (NO) production inhibitory,^{1,3} vasorelaxant,² and hepatoprotective activities.¹ and also the presence of the 3⁻ and 4⁺-acyl groups in acylated khellactone-type coumarins were essential for their strong activities. As a continuing study, we found that two new acylated khellactone-type coumarins, hyuganoside I (1, 0.013%) and hyuganin F (2, 0.0017%), were isolated from the dried leaves of this plant together with isoepoxypteryxin^{1,2,4} (5, 0.11%), isopteryxin^{1,2,4} (6, 0.13%), hyuganosides II^{1,3} (7, 0.023%) and IV⁴ (8, 0.023%), chlorogenic acid⁴ (9, 0.0013%), kaempferol 3-*O*-*β*-D-glucopyranoside⁴ (10, 0.0063%), quercetin 3-*O*-*β*-D-glucopyranoside⁴ (11, 0.0049%), quercetin 3-*O*-*β*-D-glucopyranoside⁴ (12, 0.012%), and (*Z*)-3-hexenyl *β*-D-glucopyranoside⁶ (13, 0.0079%). In this paper, we describe the isolation and absolute stereostructure elucidation of hyuganoside I (1) and hyuganin F (2). In addition, the absolute stereostructures of the acyl moieties in hyuganoside I (1),⁷ a white powder, [*α*]_D²⁵-15.0° (*c*=0.38, EtOH), C₂₇H₃₄O₁₄, showed absorption bands ascribable to hydroxyl, carbonyl, and ether functions and aromatic ring IIR (KBr): 3432, 1750–1720, 1608, 1560, 1148, 1078, 837 cm⁻¹.

hydroxyl, carbonyl, and ether functions and aromatic ring [IR (KBr): 3432, 1750–1720, 1608, 1560, 1148, 1078, 837 cm⁻¹]. Acid hydrolysis of **1** with 1.0 M hydrochloric acid (HCl) liberated D-glucose, which was identified by HPLC analysis using an optical rotation detector.^{3,8} The ¹H-NMR (500 MHz, DMSO- d_6) and ¹³C-NMR (Table 1) spectra⁹ of **1** showed signals assignable to four methyls [δ 1.10 (3H, d, J = 6.1 Hz, 4"-H₃), 1.20 (3H, s, 5"-H₃), 1.40, 1.46 (3H each, both s, 2'-gem-CH₃)], an acetyl group [δ 2.02 (3H, s, 2"-H₃)], a coumarin moiety [δ 6.29 (1H, d, J = 9.6 Hz, 3-H), 6.87 (1H, d, J = 8.5 Hz, 6-H), 7.63 (1H, d, J = 8.5 Hz, 5-H), 7.97 (1H, d, J = 9.6 Hz, 4-H)], and three methines bearing an oxygen function [δ 4.10 (1H, q, J = 6.1 Hz, 3"-H), 5.23 (1H, d, J = 5.0 Hz, 3'-H), 6.45 (1H, d, J = 5.0 Hz, 4'-H)] together with a β -D-glucopyranosyl part { δ [3.38 (1H, d, J = 6.1, 10.8 Hz), 3.64 (1H, br d, J = ca. 11 Hz), Glc-6-H₂] 4.21 (1H, d, J = 7.6 Hz, Glc-1-H)}. The aglycon of **1** named hyuganin E (**1a**)¹⁰ was obtained by enzymatic hydrolysis with naringinase, whose proton and carbon signals in the ¹H- and ¹³C-NMR spectra were superimposable on those of isoepoxypteryxin (**5**), except for the signals due to the 3"-acyl group. Alkaline treatment of **1a** with 5.0% aqueous potassium hydroxide (KOH) liberated (+)-*cis*-khellactone (**14**) and (-)-*trans*-



Figure 1

khellactone (15), which were known to be an epimerization product at the 4'-position of $14.^{2,11-13}$ The positions of the acyl groups and glucoside linkage in 1 were clarified by heteronuclear multiple-bond correlation (HMBC) experiment on 1 (Figure 1). Furthermore, in nuclear Overhauser enhancement spectroscopy (NOESY) experiment on 1, a nuclear Overhauser effect

(NOE) correlation was observed between the 3'- and 4'-proton pair, so that the absolute stereostructure of the (+)-*cis*-khellactone moiety in **1** was confirmed. In order to clarify the relative stereostructure of the diol part in 2",3"-dihydroxy-2"- methylbutyryl group, the acetonide derivative $(1b)^{14}$ was prepared from **1a**. In the NOESY experiment on **1b**, a NOE correlation was observed between the 4"-methyl and 5"-methyl protons. Furthermore, the absolute stereostructure of the 3"- position was confirmed by the application of modified Mosher's method¹⁵ to the (*R*)-MTPA (**1c**)¹⁶ and (*S*)-MTPA esters (**1d**),¹⁷ which were prepared by the treatment of **1a** with (*S*)- or (*R*)-MTPA chloride in pyridine (Figure 1). Consequently, the absolute stereostructure of **1** was elucidated as shown.

Hyuganin F (2),¹⁸ a white powder, $C_{24}H_{28}O_8$, $([\alpha]_D^{27}-34.8^{\circ} \text{ in MeOH})$. The IR spectrum of **2** showed absorption bands at 1744, 1609, 1148, and 1115 cm⁻¹ ascribable to an ester carbonyl function and aromatic ring, while the UV spectrum showed absorption maxima at 245 (log ε 3.70) and 322 (4.20) nm suggestive of a characteristic coumarin skeleton.² The ¹H-NMR (CDCl₃) and ¹³C-NMR (Table 1) spectra⁹ of **2** showed signals assignable to six methyls [δ 0.95 (t, J = 7.3 Hz, 4'''-H₃), 1.21 (d, J = 7.0 Hz, 5'''-H₃), 1.43 (d, J = 5.5 Hz, 4''-H₃), 1.46, 1.50 (both s, 2'-*gem*-CH₃), 1.59 (s, 5''-H₃)], a methylene [δ 1.72 (m, 3'''-H₂)], a methine [δ 2.40 (dd, J = 6.6, 7.0 Hz, 2'''-H)], three methines bearing an oxygen function [δ 3.08 (br d, J = *ca*. 6 Hz, 3''-H), 5.38 (d, J = 5.0 Hz, 3'-H), 6.59 (d, J = 5.0 Hz, 4''-H)], an *cis*-



olefinic proton pairs [δ 6.24 (d, J = 9.5 Hz, 3-H), 7.60 (d, J = 9.5 Hz, 4-H)], and two aromatic protons [δ 6.81 (d, J = 8.6 Hz, 6-H), 7.38 (d, J = 8.6 Hz, 5-H)]. Alkaline hydrolysis of **2** with 5.0% aqueous KOH provided **14** and **15**.^{2,11-13} On the basis of above-mentioned evidence and the 2D-NMR experiments on **2** as shown in Figure 2, the stereostructure of **2** was clarified except for the acyl moieties.

					(===) =====	(=)		
	1 ^{<i>a</i>)}	$1a^{b)}$	$2^{b)}$		1 ^{<i>a</i>)}	1a ^{b)}		$2^{b)}$
C-2	159.3	159.7	159.6	C-1"	174.1	174.9	C-1"	168.8
C-3	112.4	113.4	113.4	C-2"	77.3	77.6	C-2"	59.5
C-4	144.3	143.2	143.1	C-3"	76.0	71.0	C-3"	60.8
C-5	129.8	129.4	129.5	C-4"	11.7	15.9	C-4"	13.7
C-6	114.0	114.4	114.4	C-5"	21.8	21.4	C-5"	19.2
C-7	156.1	156.5	156.3	C-1""	169.2	171.0	C-1"	175.4
C-8	106.4	106.4	107.4	C-2"'	20.2	20.7	C-2"	41.2
C-9	153.4	153.9	153.9	Glc-1	98.7		C-3"	26.6
C-10	112.4	112.7	112.5	-2	73.4		C-4""	11.6
C-2'	77.6	77.8	77.0	-3	76.2		C-5"	16.7
C-3'	69.9	71.5	72.2	-4	70.3			
C-4'	60.2	61.4	59.8	-5	76.7			
2'-gem-CH2	22.3	22.8	21.4	-6	61.2			
	24.5	24.8	26.3					

Table 1. ¹³C-NMR Data for Hyuganoside I (1) and Hyuganins E (1a) and F (2)

Measured in ^{*a*})CD₃OD and ^{*b*})CDCl₃ at 125 MHz. Glc: β -D-glucopyranosyl

Many acylated khellactone-type coumarins were isolated from several Umbelliferae plants (e.g. *Peucedanum praeruptorum*,¹⁹ *Angelica keiskei*,²⁰ *etc.*) and they were known to show various biological activities such as platelet-aggregation inhibitory¹⁹ and anti-AIDS activities *etc.*²¹ However, the total absolute stereostructures including the acyl group were characterized only rarely. We previously reported the structures of hyuganins A (**3**) and C (**4**),² but the absolute configuration of the 2methylbutyryl moiety in **3** and **4** was remained uncharacterized. To clarify the absolute configuration of **3**, we carried out X-Ray crystallographic analysis.²² As shown in Figure 3, the 2¹¹¹-methylbutyryl moiety in **3** was determined to be a *R*orientation.²³ On the other hand, alkaline hydrolysis of isoepoxypteryxin (**5**) with 10% aqueous KOH followed by purification using reversed-phase silica gel (ODS) column chromatography and finally HPLC gave (2*R*,3*R*)-(+)-2,3-epoxy-2-methylbutyric acid²⁴ { $[\alpha]_D^{30}$ +27.9° (*c* 0.11, CHCl₃)}, so that the absolute configurations of the 2"- and 3"-positions in **5** were clarified to be in the 2"*R*, 3"*R* orientations. Finally, alkaline hydrolysis of **2** and **4** with 5% aqueous KOH and then subjection to reversed-phase silica gel column chromatography gave the organic acids fractions, which were analyzed with HPLC (optical rotation detector). As the result, (2R,3R)-(+)-2,3-epoxy-2-methylbutyric acid (**i**, 10.7 min, positive optical rotation) from **2** and (-)-(*R*)-2-methylbutyric acid (**ii**, 15.0 min, negative optical rotation) from **2** and **4** were identified by comparison of their retention times and optical rotation with those of authentic samples from **3** and **5**. On the basis of above-mentioned evidence, the absolute stereostructures of **2**–**5** were determined to be as shown.



Figure 3. Perspective View of Hyuganin A (3)

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- a) 1: High-resolution positive-ion FAB-MS: Calcd for $C_{27}H_{34}O_{14}Na$ (M+Na)⁺: 605.1846. Found: 605.1859. UV [MeOH, nm (log ε)]: 218 (sh, 4.09), 242 (sh, 3.61), 251 (sh, 3.50), 298 (sh, 3.79), 322 (3.96). Positive-ion FAB-MS: *m/z* 605 (M+Na)⁺. Negative-ion FAB-MS: *m/z* 581 (M–H)⁻. b) Hyuganoside I (1) was also isolated from the fresh roots of this plant (0.001% from the fresh roots).
- 8 M. Yoshikawa, T. Morikawa, Y. Kashima, K. Ninomiya, and H. Matsuda, J. Nat. Prod., 2003, 66, 922.
- 9 The ¹H- and ¹³C-NMR spectra of new compounds were assigned on the basis of homo- and hetero-correlation spectroscopy (¹H-¹H, ¹³C-¹H COSY) and heteronuclear multiple bond correlation (HMBC) experiments.
- 1a: A white powder, [α]_D²³ +9.7° (*c*=0.33, CHCl₃). High-resolution EI-MS: Calcd for C₂₅H₃₄O₉ (M⁺): 420.1420. Found: 420.1400. UV [MeOH, nm, (log ε)]: 217 (sh, 4.08), 244 (sh, 3.56), 252 (sh, 3.44), 298 (sh, 3.74), 323 (3.91). IR (KBr): 3440, 1750–1718, 1608, 1560, 1490, 1236, 1118, 836 cm⁻¹. ¹H-NMR (500 MHz, CDCl₃) δ: 1.19 (3H, d, *J* = 6.4 Hz, 4"-H₃), 1.36 (3H, s, 5"-H₃), 1.42, 1.46 (3H each, both s, 2'-*gem*-CH₃), 2.17 (3H, s, 2"'-H₃), 3.83 (1H, q, *J* = 6.4 Hz, 3"-H), 5.43 (1H, d, *J* = 4.9 Hz, 3'-H), 6.26 (1H, d, *J* = 9.6 Hz, 3-H), 6.59 (1H, d, *J* = 4.9 Hz, 4'-H), 6.81 (1H, d, *J* = 8.8 Hz, 6-H), 7.38 (1H, d, *J* = 8.8 Hz, 5-H), 7.61 (1H, d, *J* = 9.6 Hz, 4-H). EI-MS *m/z* (%): 420 (M⁺, 15), 286 (27), 244 (61), 229 (100), 213 (76).
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- 14 1b: ¹H-NMR (500 MHz, CDCl₃) δ: 1.17 (3H, d, J = 6.3 Hz, 4"-H₃), 1.33 (3H, s, 5"-H₃), 1.38, 1.45 (3H each, both s), 1.42, 1.46 (3H each, both s, 2'-gem-CH₃), 2.14 (3H, s, 2"-H₃), 4.31 (1H, q, J = 6.3 Hz, 3"-H), 5.34 (1H, d, J = 4.7 Hz, 3'-H), 6.25 (1H, d, J = 9.5 Hz, 3-H), 6.60 (1H, d, J = 4.7 Hz, 4'-H), 6.80 (1H, d, J = 8.7 Hz, 6-H), 7.36 (1H, d, J = 8.7 Hz, 5-H),

7.60 (1H, d, J = 9.5 Hz, 4-H). EI-MS m/z (%): 460 (M⁺, 6), 445(M⁺–CH₃, 18), 286 (15), 244 (34), 229 (68), 213 (23), 129 (100).

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- 16 **1c**: ¹H-NMR (500 MHz, CDCl₃) δ : 1.26 (3H, d, J = 6.1 Hz, 4"-H₃), 1.36 (6H, s, 2'-*gem*-CH₃), 1.40 (3H, s, 5"-H₃), 2.12 (3H, s, 2"-H₃), 5.33 (1H, d, J = 4.8 Hz, 3'-H), 5.33 (1H, q, J = 6.1 Hz, 3"-H), 6.26 (1H, d, J = 9.6 Hz, 3-H), 6.60 (1H, d, J = 4.8 Hz, 4'-H), 6.80 (1H, d, J = 8.6 Hz, 6-H), 7.36 (1H, d, J = 8.6 Hz, 5-H), 7.61 (1H, d, J = 9.6 Hz, 4-H).
- 17 1d: ¹H-NMR (500 MHz, CDCl₃) δ: 1.36 (3H, d, J = 6.4 Hz, 4"-H₃), 1.30 (3H, s, 5"-H₃), 1.32, 1.34 (3H each, both s, 2'-*gem*-CH₃), 2.14 (3H, s, 2"-H₃), 5.22 (1H, d, J = 4.7 Hz, 3'-H), 5.33 (1H, q, J = 6.4 Hz, 3"-H), 6.26 (1H, d, J = 9.5 Hz, 3-H), 6.58 (1H, d, J = 4.7 Hz, 4'-H), 6.80 (1H, d, J = 8.5 Hz, 6-H), 7.36 (1H, d, J = 8.5 Hz, 5-H), 7.60 (1H, d, J = 9.5 Hz, 4-H).
- 18 **2**: High-resolution EI-MS: Calcd for $C_{24}H_{28}O_8$ (M⁺): 444.1784. Found: 444.1796. IR (KBr): 1744, 1609, 1148, 1115 cm⁻¹. EI-MS *m/z* (%): 444 (M⁺, 4), 229 (100).
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- 22 Crystal data for **3**: Colorless prismatic crystals, mp 133–134 °C (from *n*-hexane—EtOAc), $C_{24}H_{28}O_7$, M = 428.48, crystal dimensions: $0.15 \times 0.09 \times 0.23$ mm, crystal system: orthorhombic, lattice type: primitive, lattice parameters: a = 15.051(2), b = 15.679(2), c = 9.704(2) Å, V = 2289.9(6) Å³, space group: $P2_12_12_1$ (#19), Z = 4, μ (CuK α) = 7.54 cm⁻¹, temperature: 23.0 °C, structure solution: direct methods (SHELXS-86), residuals: R = 0.095, Rw = 0.152, RI = 0.048, goodness of fit indicator: 1.16. All measurements were made on a Rigaku AFC7R diffractometer with graphite monochromated Cu-K α (λ =1.54178 Å) radiation and a rotating anode generator.
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