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## ESTROGEN RECEPTOR LIGANDS. PART 15: SYNTHESIS OF BENZOTHIOPYRAN-BASED SELECTIVE ESTROGEN RECEPTOR ALPHA MODULATORS (SERAM)

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**Abstract** – Benzothiopyran (2) was prepared and the bioactive (+)-2 was found to exhibit a reduced affinity toward the estrogen receptors (ER $\alpha/\beta$ ) when compared to the corresponding dihydrobenzoxathiin (+)-1.

In a previous communication,<sup>1</sup> we identified ER $\alpha$  subtype selective ligands or Selective Estrogen Receptor Alpha Modulators (SERAMs) that centered on the dihydrobenzoxathiin core structure.<sup>2</sup> This compound, as exemplified by 1, exhibited low nanomolar binding affinity and sub-nanomolar functional activity, as well as *in vivo* efficacy for the suppression of estradiol-driven uterine proliferation, with minimal uterotropic activity. Subsequent expanded structure-activity relationship<sup>3</sup> eventually led to a potential developmental candidate.<sup>4</sup>



Figure 1

Although dihydrobenzoxathiin (1) had excellent potency and selectivity for ER $\alpha$ , it was judged to be unacceptably prone to oxidative metabolism, with subsequent formation of covalent protein adducts.<sup>5</sup> We initially identified that an iminium ion resulting from oxidation of the piperidine residue present in the side chain of 1 was a significant contributor to the formation of covalent adducts.<sup>6</sup> Meanwhile,



Figure 2: Possible Metabolic Pathway leading to Protein Binding Adducts

formation of a quinone intermediate (1-I), generated from a phenolic-radical fragmentation process embedded in the dihydrobenzoxathiin core, had also been postulated as being responsible for the covalent adducts with biological proteins.<sup>5</sup> Intuitively, the isolation of hydroquinone (3) from the *in vitro* incubation of 1 with human liver microsomes supported the plausibility of the above metabolic pathway (Figure 2).<sup>7</sup> Therefore, it seemed prudent to replace the oxygen in the core with a methylene unit, thus eliminating the possible formation of the quinone intermediate and thereby ameliorating any potential for cytochrome P-450 mediated activation. Herein, we report the synthesis, estrogen/anti-estrogen activity, and metabolic stability of target (2).



**Scheme 1** *Reagents and Conditions*: (a) 1.5 equiv. *t*-Butylthiol, 1.8 equiv. NaH, Diglyme, 95°C, 15 h, 72% ; (b) Piperidine, AcOH, 110°C, 2 days, 50%; (c) i. LiAlH<sub>4</sub>, 0°C ii. 5% TFA, CH<sub>2</sub>Cl<sub>2</sub>, -5°C, 10 min, 90% for two steps; (d) 1 equiv. Pd black, H<sub>2</sub>, EtOH, rt, 2 days, 75%

The synthesis began with the known compound (4), which was easily synthesized from *m*-chlorophenol by a Reimer-Tiemann reaction in 60-70% yield.<sup>8</sup> Replacement of the chlorine atom with a *t*-butylthio group was initially problematic due, in part, to the lability of the benzyl group under the basic conditions.<sup>9</sup> However, the best conversion was realized by using a minimum amount of the reagents at lower temperature. The resulting *t*-butylthiobenzaldehyde (5) was condensed with a properly protected ketone (12) in the presence of piperidine/HOAc to give a mixture of (E/Z)- $\alpha$ , $\beta$ -unsaturated ketones (6).<sup>10</sup> This



Knoevenagel reaction was very sluggish and reaction times of 2 days were necessary to ensure total consumption of the starting material. Selective 1,2-reduction of  $\alpha$ , $\beta$ -unsaturated ketones (6) with 1 equiv. LiAlH<sub>4</sub> in THF gave allylic alcohols (7), thus setting the stage for subsequent cyclization to the benzothiopyran.



**Scheme 2** *Reagents and Conditions*: (a) i. 3 equiv. NaH, MOMCl, DMF, rt ii. TBAF, HOAc, THF, 80% for two steps; (b) 2-Chloroethylpiperidine, Cs<sub>2</sub>CO<sub>3</sub>, acetone, reflux; (c) 10% HCl, MeOH, reflux, 1 h, 81%

The allylic alchohols (7), without further purification, were then cyclized with TFA in dichloromethane to yield the 2,3-disubstituted 2*H*-1-benzothiopyran (8) in 90% yield.<sup>11</sup> An acid-mediated cyclization of the allylic carbocation 7-I to 7-II followed by loss of isobutylene was postulated as the reaction mechanism (Figure 3).<sup>11</sup> Hydrogenation of compound (8) with Pd black provided exclusively the racemic, *syn*-compound (9).<sup>12</sup> The *syn* relationship of the two phenyl groups was unambiguously established by the observed small coupling constant (J = 3.5 Hz,  $\delta = 4.2$  ppm, see ref. 2), between H<sub>2</sub>-H<sub>3</sub>. Subsequent chiral resolution of the racemic benzothiopyran (9) was realized by HPLC, using a Chiracel AD column and 30% IPA/hexane as the eluant, to provide the dextrorotatory enantiomer ((+)-9).<sup>13</sup>

With (+)-9 now readily available, protection of the hydroxyl group in the A-ring, followed by deprotection of TIPS group produced (+)-10 in quantitative yield (Scheme 2). The phenol ((+)-10) was next converted to the respective ether (11) by reaction with 2-piperidinoethyl chloride in the presence of cesium carbonate in acetone. Removal of the two MOM groups with dilute acid resulted in the final

compound ((+)-2) in 81% yield.<sup>14</sup> Similarly, in the racemic series, (9) was also converted to (2) by application of the same procedure.

It is clear from the data in Table 1 that the benzothiopyran (2) retained the ER $\alpha$  potency and the magnitude of receptor subtype selectivity (ER $\beta$ /ER $\alpha$  ratio) exhibited by the dihydrobenzoxathiin (1) in an *in vitro* ER binding assay. However, the replacement of the oxygen atom with a methylene unit altered the size of the ring, which presumably contributed to the reduction of the binding affinity (ER $\alpha$  for (+)-1= 0.8 nM and (+)-2= 3.1 nM).<sup>15</sup> This weaker potency paralleled the weaker estradiol antagonism observed in the immature rat uterine model (92% inhibition vs 76% inhibition).

Table 1. Binding affinities<sup>a</sup> and *in vivo* data

Binding affinity				Uterine Assay (sc) <sup>c</sup>
Compd	ERα	ER $\beta$ ( $\alpha$ selectivity)		% inhib @ 1mpk / % control @ 1 mpk
(+)-1 <sup>b</sup>	0.8	45	(56)	92/0.4
$(\underline{+})$ -1 <sup>b</sup>	3.0	143	(48)	77/5.0
$(+)-2^{d}$	3.1	130	(43)	76/4.7
( <u>+</u> )-2	5.8	590	(101)	N.A.
0				

<sup>a</sup>Utilizing full length recombinant human ER $\alpha/\beta$  proteins; IC<sub>50</sub> (nM), n=>48-56, see ref 1. <sup>b</sup>Absolute stereochemistry was determined by X-Ray crystallography, see ref 1 <sup>c</sup>See ref 1 & 3; <sup>d</sup>The absolute stereochemistry of (+)-**2** was assigned based on analogy with (+)-**1**, see ref 13 and 14

As with the dihydrobenzoxathiins, only the single enantiomer ((+)-2) reproduced the activity exhibited by the racemate (2). In addition, the metabolism of the tritium-labelled ((+)-2) was investigated and, in spite of the modification, was found to have a similar level of covalent protein adducts as (+)-1 in incubations with rat liver microsomes. Further results in this area will be reported in future publications from this laboratory.

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- 12. Ca. 3% yield of trans isomer was also formed. See also ref. 2.
- 13. The absolute stereochemistry of (+)-9 was assigned based on analogy with (+)-1, and confirmed later by biological data, see also ref. 1 and Table 1. (+)-10:  $[\alpha]_D$  +227.6° (*c* 0.69, MeOH)
- 14. (+)-2: [α]<sub>D</sub> +193.2° (*c* 0.49, MeOH); <sup>1</sup>H NMR (500 MHz, Acetone-d<sub>6</sub>) δ (ppm) 7.00 (d, *J*=8.3 Hz, 1H), 6.82 (d, *J*=8.4 Hz, 2H), 6.76 (d, *J*=8.5 Hz, 2H), 6.68 (d, *J*=8.7 Hz, 2H), 6.68 (d, hidden, 1H), 6.60 (d, *J*=8.5 Hz, 2H), 6.58 (dd, *J*=2.5 Hz, 1H), 4.49 (d, *J*=3.7 Hz, 1H), 4.10 (t, 2H), 3.60 (m, 1H), 3.05 (dd, *J*=10.5 and 16.25 Hz, 1H), 2.90 (dd, *J*=3.6 and 16.00 Hz, 1H), 2.80 (br t, 2H), 2.60 (br s, 4H), 1.60 (m, 4H), 1.50 (br m, 2H); MS *m/z* 462.0 (M<sup>+</sup>)
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