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AMPHIDINOLACTONE A, A NEW 13-MEMBERED MACROLIDE FROM DINOFLAGELLATE *AMPHIDINIUM* SP.

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Abstract - A new 13-membered macrolide, amphidinolactone A (1), has been isolated from a marine dinoflagellate *Amphidinium* sp., and the structure and relative stereochemistry were elucidated by spectroscopic data. Amphidinolactone A (1) is the first macrolide without a branched methyl or an exomethylene among all the macrolides isolated from the dinoflagellates so far.

INTRODUCTION

Marine dinoflagellates of the genus *Amphidinium* have been recognized as a source of novel secondary metabolites with interesting structures and bioactivities.¹⁻⁷ In our continuing search for bioactive metabolites from Okinawan marine organisms, we have investigated extracts of laboratory cultured dinoflagellates *Amphidinium* sp., which were symbionts of the Okinawan marine acoel flatworms *Amphiscolops* sp., and isolated a series of cytotoxic macrolides, amphidinolides,¹ as well as long chain polyhydroxyl polyketides.^{2,6,7} Here we describe the isolation and structure elucidation of a new 13-membered macrolide, amphidinolactone A (1), which was isolated from extracts of a strain (Y-25) of the dinoflagellate *Amphidinium* sp.



RESULTS AND DISCUSSION

The dinoflagellate *Amphidinium* sp. (strain number Y-25) was isolated from inside cells of a marine acoel flatworm *Amphiscolops breviviridis* collected off Sunabe, Okinawa. The harvested cells of the cultured dinoflagellate were extracted with MeOH/toluene (3:1). After addition of 1 M NaCl, the mixture was extracted with toluene. The toluene-soluble fraction was evaporated under reduced pressure to give a residue, which was subjected to a silica gel column and a Sep-Pak C_{18} cartridge followed by C_{18} HPLC to afford amphidinolactone A (1, 0.1 mg, 0.000014%, wet weight).

Position	δ_{H}		$\delta_{\rm C}$	
1	-		α	S
2a	2.19	(1H, m)	32.3	t
2b	2.10	(1H, m)		
3a	1.86	(1H, m)	22.2	t
3b	1.26	(1H, m)		
4a	2.32	(1H, m)	25.5	t
4b	1.86	(1H, m)		
5	5.30	(1H, ddd, 11.0, 9.6, 4.6 Hz)	130.2	d
6	5.66	(1H, m)	124.4	d
7a	2.32	(1H, m)	35.6	t
7b	2.19	(1H, m)		
8	4.00	(1H, ddd,7.6, 6.7, 2.9 Hz)	71.8	d
9	5.53	(1H, dd, 15.8, 7.6 Hz)	135.9	d
10	5.23	(1H, dd, 15.8, 7.6 Hz)	130.1	d
11	3.83	(1H, dd, 8.1, 7.6 Hz)	73.5	d
12	5.02	(1H, ddd, 8.8, 7.3, 3.7 Hz)	73.5	d
13a	2.68	(1H, ddd, 14.5, 3.7, 3.7 Hz)	29.0	t
13b	2.50	(1H, ddd, 14.5, 7.3, 7.3 Hz)		
14	5.58	(1H, m)	131.5	d
15	5.60	(1H, m)	124.8	d
16	2.88	(2H, m)	25.6	t
17	5.46	(1H, m)	126.7	d
18	5.46	(1H, m)	132.4	d
19	2.06	(2H, m)	20.0	t
20	0.96	(3H, t, 7.5 Hz)	14.3	q

Table 1. ¹H and ¹³C NMR Data of Amphidinolactone A (1) in C_6D_6 .

² not observed.

Amphidinolactone A (1) had the molecular formula of $C_{20}H_{30}O_4$ as revealed by HRESIMS [*m/z* 357.2057 (M+Na)⁺, +1.6 mmu]. IR absorptions at 3370 and 1720 cm⁻¹ indicated the presence of hydroxy group and ester carbonyl group. ¹H and ¹³C NMR data (Table 1) (obtained from the HMQC spectrum) of 1 disclosed the presence of eight sp² methines, three oxymethines, seven sp³ methylenes, and one methyl group. Considering the molecular formula, the existence of a carbonyl unit (C=O) was indicated. Since five out of six unsaturations were accounted for, amphidinolactone A (1) was inferred to contain one ring. Detailed analyses of the ¹H-¹H COSY spectrum of 1 established connectivities of a long carbon chain from C-2 to C-20. ¹H and ¹³C chemical shifts of CH₂-2 (δ_H 2.19, 2.10; δ_C 32.3) and CH-12 (δ_H 5.02; δ_C 73.0) suggested that C-12 was involved in an ester linkage with C-1. ¹H-¹H couplings ($J_{5,6}$ = 11.0 Hz, $J_{9,10}$ = 15.8 Hz) of two disubstituted double bonds at C-5 and C-9 indicated the *Z* and *E* geometry, respectively. Geometries of two disubstituted double bonds at C-14 and C-17 were assigned as both *Z* from NOESY correlations of H₂-13/H₂-16 and H₂-16/H₂-19 as well as the carbon chemical shifts of C-16 (δ_C 25.6), which was typical value for a methylene carbon between two *Z* double bonds.⁸ Thus, the gross structure of amphidinolactone A (1) was elucidated as shown in Fig. 1.



Figure 1. Key chemical shifts and 2D NMR correlations for amphidinolactone A (1).

The relative stereochemistry of C-8, C-11, and C-13 in **1** was deduced from NOESY correlations as shown in Figure 2. NOESY correlations for H-6/H-8, H-8/H-10, and H-10/H-12 implied that H-8 and H-12 were both α -oriented. On the other hand, NOESY correlations were observed for H-7b/H-9, H-9/H-11, and H-11/H-13, suggesting that H-11 was β -oriented. Therefore, the relative stereochemistry of C-8, C-11, and C-12 of amphidinolactone A was elucidated to be **1**.



<----> NOESY

Figure 2. Selected NOESY correlations and relative stereochemistry for amphidinolactone A (1).

Amphidinolactone A (1) is a new 13-membered macrolide consisting a C₂₀ carbon-chain possessing four disubstituted double bonds and two hydroxy groups. Amphidinolactone A (1) is the first macrolide without a branched methyl or an exomethylene among all the macrolides isolated from the dinoflagellates *Amphidinium* sp. so far. Amphidinolactone A (1) showed cytotoxicity against L1210 murine leukemia cells (IC₅₀, 8 µg/mL), while 1 did not show such activity against human epidermoid carcinoma KB cells (IC₅₀ > 10 µg/mL) in vitro.

EXPERIMENTAL

General Experimental Procedures.

IR spectrum was recorded on a JASCO FT/IR-230. ¹H and ¹³C NMR spectra were recorded on a Bruker AMX-600 spectrometer using 2.5 mm symmetrical microtubes (DMS-0025, Shigemi Co., Ltd). The 7.20 and 128.0 ppm resonances of residual C_6D_6 were used as internal references for ¹H and ¹³C NMR spectra, respectively. ESI mass spectra were obtained on a JEOL JMS- 700TZ spectrometer.

Material.

The dinoflagellate *Amphidinium* sp. (strain number Y-25) was isolated from inside cells of a marine acoel flatworm *Amphiscolops breviviridis* collected off Sunabe, Okinawa. The dinoflagellate has been deposited in Graduate School of Pharmaceutical Sciences, Hokkaido University.

Extraction and Isolation.

The dinoflagellate was unialgally cultured at 25 °C for 2 weeks in a seawater medium enriched with 1% ES supplement.⁹ The harvested cells of the cultured dinoflagellate (713 g, wet weight, from 3000 L of culture) were extracted with MeOH/toluene (3:1). After addition of 1 M NaCl, the mixture was extracted with toluene. The toluene-soluble fraction was evaporated under reduced pressure to give a residue (1.13 g), which was subjected to a silica gel column (CHCl₃/MeOH, 1:0 \rightarrow 0:1) and a Sep-Pak C₁₈ cartridge (CH₃CN/H₂O, 7:3) followed by C₁₈ HPLC [YMC Pack Pro C₁₈, 5 µm, 10 mm x 250 mm; eluent, MeOH/H₂O, 80:20; flow rate, 2.0 mL/min; UV detection at 210 nm] to afford amphidinolactone A (1, 0.1 mg, 0.000014 %, wet weight).

Amphidinolactone A (1): colorless amorphous solid; $[\alpha]_D^{19}$ -62 (*c* 0.065, C₆H₆); IR (film) ν_{max} 3370 and 1720 cm⁻¹; ¹H and ¹³C NMR data (Table 1); ESIMS *m*/*z* 357 (M+Na)⁺; HRESIMS *m*/*z* 357.2057 (M+Na; calcd for C₂₀H₃₀O₄Na, 357.2041).

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REFERENCES

- (a) J. Kobayashi and M. Tsuda, *Nat. Prod. Rep.*, 2004, **21**, 77. (b) T. Kubota, Y. Sakuma, M. Tsuda, and J. Kobayashi, *Mar. Drugs*, 2004, **3**, 83. (c) M. Tsuda, Y. Kariya, R. Iwamoto, E. Fukushi, J. Kawabata, and J. Kobayashi, *Mar. Drugs*, 2005, **3**, 1. (d) T. Kubota, T. Endo, Y. Takahashi, M. Tsuda, and J. Kobayashi, *J. Antibiot.*, 2006, **59**, 512.
- (a) M. Satake, M. Murata, T. Yasumoto, T. Fujita, and H. Naoki, *J. Am. Chem. Soc.*, 1991, **113**, 9859.
 (b) G. K. Paul, N. Matsumori, M. Murata, and K. Tachibana, *Tetrahedron Lett.*, 1995, **36**, 6279. (c) G. K. Paul, N. Matsumori, K. Konoki, M. Murata, and K. Tachibana, *J. Mar Biotechnol.*, 1997, **5**, 124.
 (d) R. Echigoya, L. Rhodes, Y. Oshima, and M. Satake, *Harmful Algae*, 2005, **4**, 383. (e) M. Morsy, S. Matsuoka, T. Houdai, N. Matsumori, S. Adachi, M. Murata, T. Iwashita, and T. Fujita, *Tetrahedron*, 2005, **61**, 8606.
- (a) Y. Doi, M. Ishibashi, H. Nakamichi, T. Kosaka, T. Ishikawa, and J. Kobayashi, J. Org. Chem., 1997, 62, 3820.
 (b) T. Kubota, M. Tsuda, Y. Doi, A. Takahashi, H. Nakamichi, M. Ishibashi, E. Fukushi, J. Kawabata, and J. Kobayashi, *Tetrahedron*, 1998, 54, 14455.
 (c) T. Kubota, A. Takahashi, M. Tsuda, and J. Kobayashi, *Mar. Drugs*, 2005, 3, 113.

- (a) X. Huang, D. Zhao, Y. Guo, H. Wu, L. Lin, Z. Wang, J. Ding, and Y. Lin, *Bioorg. Med. Chem.* Lett., 2004, 14, 3117. (b) X. Huang, D. Zhao, Y. Guo, H. Wu, E. Trivellone, and G. Cimino, *Tetrahedron Lett.*, 2004, 45, 5501.
- 5. K. Washida, T. Koyama, K. Yamada, M. Kita, and D. Uemura, Tetrahedron Lett., 2006, 47, 2521.
- (a) J. Kobayashi, T. Kubota, M. Takahashi, M. Ishibashi, M. Tsuda, and H. Naoki, J. Org. Chem., 1999, 64, 1478. (b) T. Kubota, M. Tsuda, M. Takahashi, M. Ishibashi, H. Naoki, and J. Kobayashi, J. Chem. Soc., Perkin Trans. 1, 1999, 64, 3483. (c) T. Kubota, M. Tsuda, M. Takahashi, M. Ishibashi, S. Oka, and J. Kobayashi, Chem. Pharm. Bull., 2000, 48, 1447.
- 7. T. Kubota, Y. Sakuma, K. Shimbo, M. Tsuda, M. Nakano, Y. Uozumi, and J. Kobayashi, *Tetrahedron Lett.*, 2006, **47**, 4369.
- E. Wenkert, B. L. Buckwalter, I. R. Burfitt, M. J. Gasic, H. E. Gottlieb, E. W. Hagaman, F. M. Schell, and P. M. Wovkulich 'Topics in Carbon-13 NMR Spectroscopy,' Vol. 2, ed. by G. C. Levy, Wiley, New York, 1975, pp. 81.
- 9. L. Provasoli, In *Culture and Collection of Algae*; ed. by A. Watanabe and A. Hattori, Japanese Society of Plant Physiology: Tokyo, 1968; pp. 63-75.