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SYNTHESIS OF THE JK RING FRAGMENTS OF YESSOTOXIN AND 42,43,44,45,46,47,55-HEPTANOR-41-OXOYESSOTOXIN

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Abstract – The JK ring fragment (7) of 42,43,44,45,46,47,55-heptanor-41-oxoyessotoxin was synthesized via enyne metathesis and 6-*exo* cyclization of a hydroxy epoxide. Conversion of 7 into the JK ring fragment (6) of yessotoxin was achieved in a single step by treatment with an alkenyllithium.

Yessotoxin (YTX, 1)¹ is a disulfated polyether toxin produced by the dinoflagellate *Protoceratium* and by *Lingulodinium* species (Figure 1).² Although YTX was first isolated in association with diarrhetic shellfish poisoning (DSP), it has been removed from the category of DSP toxins due to a lack of diarrhoegenicity.³ A number of YTX congeners, for instance 42,43,44,45,46,47,55-heptanor-41-oxoyessotoxin (2) and 3-5 (Figure 1),⁴ have been identified, whose structural diversity is focused on the K ring and side chains. YTX has recently been shown to exhibit intriguing biological activities, e.g., (i) cytotoxicity against human tumor cell lines,⁵ (ii) activation of caspases via mitochondrial signal transduction pathways,⁶ and (iii) activation of phosphodiesterases.⁷ Meanwhile, structure-activity relationship studies using YTX congeners have been hampered by their scarce availability from natural sources. During the course of our synthetic studies of YTX and its congeners,⁸ we developed an efficient method for convergent synthesis of the CDEF and FGHI ring systems via α -cyano ethers.⁹ Herein, we describe a synthesis of the JK ring fragments of YTX and 2.¹⁰

As shown in Scheme 1, the JK ring fragment (6) of YTX could be retrosynthetically disconnected into the JK ring fragment (7) of 2 and alkenylstannane (8). The methylketone (7) would be derived from allylic alcohol (9) via Sharpless asymmetric epoxidation and 6-*exo* cyclization of a hydroxy epoxide, and

[§] Dedicated to Professor Yoshito Kishi on the occasion of his 70th birthday.



Figure 1. Structures of yessotoxin (1) and its congeners (2-5).

incidentally, construction of the K ring system based on a similar method has recently been reported by Kadota *et al.*¹⁰ For expeditious construction of the conjugated diene system of **9**, we envisaged envie metathesis¹¹ of alkyne (**10**) and 2-methyl-2-propen-1-ol (**11**).



Scheme 1. Synthesis plan for the JK ring fragments.

Synthesis of the JK ring fragment began with a one-pot preparation of triflate (13) from diol $(12)^{9,12}$ by Mori's protocol¹³ (Scheme 2). Alkylation of 13 with lithium trimethylsilylacetylide, followed by removal of the TMS group afforded alkyne (10) in 68% yield for three steps. One of the key steps of the present synthesis, enyne metathesis of 10 and 2-methyl-2-propen-1-ol (11) using Grubbs' second generation catalyst,¹⁴ proceeded smoothly to afford *E*-diene (9) as the major product (*E* : *Z* = 7 : 1) in 57% yield with concomitant formation of dienal (14) (10%) as a byproduct, which was readily converted to 9 by Luche reduction.¹⁵ Portionwise addition of the Grubbs catalyst (0.5 eq) in one portion reduced the ratio of the *E*-diene (*E* : *Z* = 3 : 1). Sharpless epoxidation of 9, followed by removal of the TBS group with TBAF

furnished hydroxy epoxide (15). Acid catalyzed 6-*exo* cyclization¹⁶ of 15 by treatment with CSA in dichloromethane afforded *vic*-diol (16). Oxidative cleavage of the glycol (16) with NaIO₄ resulted in the formation of the JK ring fragment (7) of 2.



Scheme 2. *Reagents and conditions:* (a) Tf₂O, 2,6-lutidine, CH₂Cl₂, -78 °C to -60 °C, 40 min, then TBSOTf, -78 °C to -55 °C, 3 h; (b) trimethylsilylacetylene, *n*-BuLi, HMPA, THF, -78 °C to -55 °C, 30 min; (c) K₂CO₃, MeOH, rt, 15 h, 68% (3 steps); (d) Grubbs' 2nd generation cat. (0.1 eq + 0.1 eq + 0.05 eq), 2-methyl-2-propen-1-ol (10 eq), toluene, reflux, 3 h, **9**: 57%, **14**: 10%; (e) CeCl₃·7H₂O, NaBH₄, EtOH, 0 °C, 30 min, 50%; (f) D-(-)-DET, Ti(O*i*-Pr)₄, TBHP, MS4A, CH₂Cl₂, -20 °C, 1 h; (g) TBAF, THF, rt, 46% (2 steps); (h) CSA, CH₂Cl₂, 0 °C, 3 h; (i) NaIO₄, THF, pH 7 buffer, rt, 6 h, 50% (2 steps).

We then turned our attention to the synthesis of the JK ring fragment (6) of YTX. The alkenylstannane (8) was prepared as shown in Scheme 3. Treatment of 2,3-dibromo-1-propene (17) with vinylmagnesium bromide in THF at 60 °C gave 2-bromo-1,4-pentadiene (18),¹⁷ which was purified by distillation. Stille coupling of 18 using 2.0 equivalent of *trans*-1,2-bis(tri-*n*-butylstannyl)ethylene with Pd(PPh₃)₄ gave the stannane (8).¹⁸ Treatment of the methylketone (7) with alkenyllithium (19) generated from 8 resulted in the formation of desired 6 and its 41-epimer (20) (6:20 = 1.2 : 1) in 66% yield, which were separated by silica gel chromatography (Scheme 4).¹⁹ Judging from the ¹H NMR spectra,²⁰ chemical shifts of Me-41 and H-42 of 6 (δ 1.39 and 5.82, respectively) more resembled those of YTX (δ 1.43 and 5.86, respectively) than those of 20 (δ 1.29 and 5.99), and therefore, we assigned the structures as depicted in Scheme 4. Although improvement of the stereoselectivity of the final alkylation step is required, the present method would be a straightforward way for the diverse synthesis of YTX congeners, not only naturally occurring but also 41-epimers useful for structure-activity relationship studies.



Scheme 3. *Reagents and conditions:* (a) vinylmagnesium bromide (2.0 eq), THF, 60 °C, 15 h, 7% (isolated yield, ca. 50% conversion yield); (b) *trans*-1,2-bis(tri-*n*-butylstannyl)ethylene (2.0 eq), Pd(PPh₃)₄ (0.15 eq), benzene, reflux, 12 h, 37%.



Scheme 4. *Reagents and conditions:* (a) *n*-BuLi, 8, THF, -78 °C, 3 min, then 7, -78 °C to -55 °C, 1 h, 66%.

In conclusion, the JK ring fragment (7) of 42,43,44,45,46,47,55-heptanor-41-oxoyessotoxin (2) was synthesized via enyne metathesis and 6-*exo* cyclization of a hydroxy epoxide. Single step conversion of 7 into the JK ring fragment (6) of YTX has been achieved by treatment with the alkenyllithium (19) prepared from stannane (8). Further studies directed towards the total synthesis of YTX and its congeners are currently in progress in our laboratory.

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19. Spectral data of 6: ¹H NMR (500 MHz, CDCl₃) & 7.47-7.45 (2H, m, Ph), 7.37-7.34 (3H, m, Ph), 6.30 (1H, d, J = 16.0 Hz, H43), 5.88 (1H, dddd, J = 17.0, 10.5, 6.5, 6.5 Hz, H46), 5.82 (1H, d, J = 16.0 Hz, H42), 5.08 (1H, dddd, J = 17.0, 1.5, 1.5 Hz, H47), 5.05 (1H, d, J = 2.0 Hz, 44=CH₂), 5.04 (1H, dddd, J = 10.5, 1.5, 1.5, 1.5 Hz, H47), 5.02 (1H, d, J = 2.0 Hz, 44=CH₂), 4.98 (1H, s, 39=CH₂), 4.79 (1H, s, 39=CH₂), 4.18 (1H, ddd, J = 11.5, 10.5, 4.5 Hz, H36), 3.88 (1H, s, H40), 3.86 (1H, d, J = 10.0 Hz, H32), 3.73 (1H, dd, J = 11.5, 4.5 Hz, H34), 3.66 (1H, d, J = 10.0 Hz, H32), 3.45 (1H, ddd, J = 11.5, 4.5 Hz, H34), 3.66 (1H, d, J = 10.0 Hz, H32), 3.45 (1H, ddd, J = 12.0, 10.5, 4.5 Hz, H37), 2.97 (2H, dd, J = 6.5, 1.5 Hz, H45), 2.65 (1H, dd, J = 12.0, 12.0 Hz, H38), 2.43 (1H, dd, J = 12.0, 4.5 Hz, H38), 2.17 (1H, ddd, J = 11.5, 4.5, 4.5 Hz, H35eq), 1.67 (1H, ddd, J = 11.5, 11.5, 11.5Hz, H35ax), 1.48 (3H, s, 33-Me), 1.39 (3H, s, 41-Me); ESI-MS 461 (M+Na⁺).

Spectral data of **20**: ¹H NMR (500 MHz, CDCl₃) δ 7.46-7.44 (2H, m, Ph), 7.36-7.34 (3H, m, Ph), 6.41 (1H, d, J = 16.0 Hz, H43), 5.99 (1H, d, J = 16.0 Hz, H42), 5.92 (1H, ddd, J = 17.0, 10.5, 6.5, 6.5 Hz, H46), 5.11 (1H, d, J = 2.0 Hz, 44=CH₂), 5.10 (1H, dd, J = 17.0, 1.5 Hz, H47), 5.06 (1H, d, J = 2.0 Hz, 44=CH₂), 5.04 (1H, dd, J = 10.5, 1.5 Hz, H47), 4.98 (1H, s, 39=CH₂), 4.93 (1H, s, 39=CH₂), 4.08 (1H, ddd, J = 11.5, 9.5, 4.5 Hz, H36), 3.90 (1H, s, H40), 3.85 (1H, d, J = 10.0 Hz, H32), 3.71 (1H, dd, J = 11.5, 4.5 Hz, H34), 3.66 (1H, d, J = 10.0 Hz, H32), 3.44 (1H, ddd, J = 12.5, 9.5, 5.0 Hz, H37), 3.01 (2H, d, J = 6.5 Hz, H45), 2.70 (1H, dd, J = 12.5, 12.5 Hz, H38), 2.44 (1H, dd, J = 11.5, 11.5, 15. Hz, H35ax), 1.46 (3H, s, 33-Me), 1.29 (3H, s, 41-Me); ESI-MS 461 (M+Na⁺).

20. The carbon numbering of compounds in this article corresponds to that of yessotoxin.