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LYCOPLADINE E, A NEW C₁₆N₁-TYPE ALKALOID FROM LYCOPODIUM COMPLANATUM

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Abstract - A new $C_{16}N_1$ -type alkaloid, lycopladine E (1), has been isolated from the club moss *Lycopodium complanatum*, and the structure and absolute stereochemistry were elucidated on the basis of spectroscopic data and chemical correlation.

INTRODUCTION

Club moss (Lycopodiaceae) are known to be a rich source of *Lycopodium* alkaloids possessing unique heterocyclic ring systems such as $C_{16}N_1$, $C_{16}N_2$, and $C_{27}N_3$, which have attracted great interest from biogenetic, synthetic, and biological points of view.¹ In our continuing efforts to find new *Lycopodium* alkaloids,² a new $C_{16}N_1$ -type alkaloid, lycopladine E (1), was isolated from the club moss *Lycopodium complanatum*. In this paper, we describe the isolation and structure elucidation of 1.



RESULTS AND DISCUSSION

The club moss *L. complanatum* collected in Nayoro, Hokkaido, were extracted with MeOH, and the MeOH extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which were adjusted at pH 10 with saturated Na₂CO₃, were partitioned with CHCl₃. CHCl₃-soluble materials were subjected to an amino silica gel column (hexane/EtOAc, then CHCl₃/MeOH), in which a fraction eluted with hexane/EtOAc (10:1) was purified by LH-20 (MeOH) and silica gel (CHCl₃/MeOH) columns to afford lycopladine E (1, 0.000008% yield), lycopodine (2),³ lycodine,⁴ lycopladines A~D,⁵ complanadines A, C, and D,⁶ and lyconadines A and B.^{5b, 7}

Lycopladine E {1, $[\alpha]_D^{23}$ -28.8 (*c* 0.1, MeOH)} was revealed to have the molecular formula, C₁₈H₃₀N₁O₃, by HRESIMS data [*m*/*z* 308.2204, (M+H)⁺, Δ -2.1 mmu]. IR absorptions implied the presence of a

Table 1. ¹H and ¹³C NMR Data of Lycopladine E (1) in CD₃OD.

Position	$\delta_{\rm H}$	$\delta_{\rm C}$		Position	δ_{H}	$\delta_{\rm C}$	
1a	3.64 (1H, m)	64.6	t	9a	4.27 (1H, dt, 12.8, 3.2)	60.6	t
1b	2.99 (1H, dd, 13.2, 4.8)			9b	2.85 (1H, brd, 12.7)		
2a	2.00 (1H, m)	22.9	t	10a	2.37 (1H, m)	20.9	t
2b	1.83 (1H, m)			10b	1.25 (1H, m)		
3a	1.83 (1H, m)	21.9	t	11a	1.83 (1H, m)	23.1	t
3b	1.45 (1H, m)			11b	1.38 (1H, brd, 12.0)		
4	2.92 (1H, m)	36.7	d	12	2.23 (1H, m)	39.2	d
5	5.22 (1H, t, 6.8)	70.4	d	13		75.0	S
6a	2.25 (1H, m)	30.9	t	14	2.15 (2H, m)	33.5	t
6b	1.49 (1H, d, 16.4)			15	2.80 (1H, m)	24.9	d
7	1.88 (1H, m)	36.7	d	16	0.99 (3H, d, 6.3)	24.4	q
8a	1.68 (1H, m)	41.9	t	17		170.6	S
8b	1.25 (1H, m)			18	2.04 (3H, s)	21.1	q



Figure 1. Selected 2D NMR correlations for lycopladine E (1).

carbonyl group (1730 cm⁻¹). The ¹H and ¹³C NMR (Table 1) spectra of **1** showed signals due to one sp² quaternary carbon, one sp³ quaternary carbon, five sp³ methines, nine sp³ methylenes, and two methyls. Among them, one methine (δ_C 75.0) and two methylenes (δ_C 64.6, and 60.6) were ascribed to those bearing a nitrogen atom.

The ¹H-¹H COSY and TOCSY spectra revealed two structural units **a** (C-1 to C-8, C-14 to C-16, and C-8 to C-15) and **b** (C-9 to C-12). HMBC correlations of H-5 (δ_{H} 5.22) to C-13 (δ_{C} 75.0), H-6b (δ_{H} 1.49) to C-12 (δ_{C} 39.2), H-12 (δ_{H} 2.23) to C-4 (δ_{C} 36.7), and H₂-14 (δ_{H} 2.15) to C-4 (δ_{C} 36.7) and C-12 (δ_{C} 39.2) suggested connectivities among C-4, C-12, and C-14 via C-13. HMBC cross-peaks of an oxymethine proton H-5 (δ_{H} 5.22) and a singlet methyl proton H₃-18 (δ_{H} 2.04) to an ester carbonyl carbon C-17 (δ_{C} 170.6) revealed that an acetoxy group was attached to C-5. ¹H and ¹³C NMR data suggested connections of C-1, C-9, and C-13 via the remaining oxygenated nitrogen atom. Thus, the gross structure of lycopladine E was elucidated to be **1**.

The relative stereochemistry of **1** was deduced from cross-peaks observed in the NOESY spectrum as shown in computer-generated 3D drawing (Figure 2). These NOESY correlations indicated the relative stereochemistry of C-5 and C-15 as well as chair conformations of the two six-membered rings in the trans quinolizidine ring (N-1, C-1 to C-4, and C9 to C-13) and two cyclohexane rings in bicyclo[3.3.1]nonane moiety (C-4 to C-8 and C-12 to 15).

The absolute configuration of lycopladine E (1) was elucidated by chemical correlation as follows. Lycopodine $(2)^3$ was converted into dihydrolycopodine $(3)^{8,9}$ by the reported procedure⁹, which was



Figure 2. Selected NOESY correlations and relative stereochemistry for lycopladine E (1).



Scheme 1. Chemical conversion of lycopodine (2) to lycopladine E (1).

treated with pyridine and acetic anhydride to afford 5-*O*-acetyldihydrolycopodine (4)¹⁰. Compound 4 was oxidized by *m*-CPBA to give compound 1 as a single product (Scheme 1), whose spectral data and $[\alpha]_D$ value { $[\alpha]_D^{23}$ -29.4 (*c* 0.5, MeOH)} were coincident with those of natural lycopladine E (1). Thus, the absolute configurations at six chiral centers in lycopladine E (1) were established to be *4S*, *5R*, *7S*, *12R*, *13R*, and *15R*.

Lycopladine E (1) is a new C_{16} N-type alkaloid having a lycopodane-skeleton with N-oxide and acetoxy group at C-5. Effects of lycopladine E (1) on neurotrophic factor biosynthesis in 1321N1 human astrocytoma cells were examined by a semiquantitative RT-PCR method^{11,12} to find that the mRNA expressions for NGF were enhanced by 1.

EXPERIMENTAL

General Experimental Procedures

Optical rotation was recorded on a JASCO P-1030 polarimeter. IR spectrum was recorded on a JASCO FT/IR-230 spectrometer. NMR spectra were recorded on a Bruker AMX-600 spectrometer using 2.5 mm micro cells (Shigemi Co., Ltd). The 3.31 and 49.0 ppm resonances of residual CD₃OD were used as internal references for ¹H and ¹³C NMR spectra, respectively. Positive-mode ESIMS was obtained on a JEOL JMS 700-TZ spectrometer using a sample dissolved in MeOH.

Plant Material

The club moss *Lycopodium complanatum* was collected at Nayoro in Hokkaido in 2001. The botanical identification was made by Mr. N. Yoshida, Health Sciences University of Hokkaido. A voucher specimen has been deposited in the herbarium of Hokkaido University.

Extraction and Isolation

The club moss *Lycopodium complanatum* (3 kg) was crushed and extracted with MeOH. The MeOH extract was treated with 3% tartaric acid (pH 2) and then partitioned with EtOAc. The aqueous layer was

treated with saturated Na₂CO₃ (aq) to pH 10 and extracted with CHCl₃ to give a crude alkaloidal fraction. A part of the alkaloidal fraction was purified by an amino silica gel column (hexane/EtOAc, $50:1\rightarrow1:1$ and then CHCl₃/MeOH, $1:0\rightarrow0:1$), in which a fraction eluted with hexane/EtOAc (10:1) was purified by LH-20 (CHCl₃/MeOH, 1:1) and silica gel columns (CHCl₃/MeOH, $1:0\rightarrow1:1$) to afford lycopladine E (**1**, 0.2 mg, 0.000008% yield), lycopodine (**2**),³ lycodine,⁴ lycopladines A~D,⁵ complanadines A, C, and D,⁶ and lyconadines A and B^{5b, 7}

Lycopladine E (1): A colorless amorphous solid; $[\alpha]_D^{23}$ -28.8 (*c* 0.1, MeOH); IR (KBr) ν_{max} 1730 and 1230 cm⁻¹; ¹H and ¹³C NMR data (Table 1); ESIMS *m/z* 308 (M+H)⁺; HRESIMS *m/z* 308.2204 (M+H; calcd for C₁₈H₃₀N₁O₃, 308.2225).

Lycopladine E (1) derived from lycopodine (2): Lycopodine (2, 19.0 mg, 0.077 mmol) was converted to dihydrolycopodine (3, 11.0 mg) by the reported procedure, part of which (8.7 mg) was treated with pyridine (1.0 mL) and acetic anhydride (1.0 mL) at 90 °C for 5 h. The reaction mixture was evaporated in vacuo to give 5-*O*-acetyldihydrolycopodine (4, 14.2 mg), part of which (12.8 mg) was treated with CH_2Cl_2 (1.0 mL) and *m*-CPBA (14.2 mg, 0.082 mmol) at 4 °C for 1 h. The reaction mixture was partitioned between CHCl₃ and saturated Na₂CO₃ (aq), and the CHCl₃ layer was evaporated in vacuo. The residue was purified by a silica gel column (CHCl₃/MeOH, 1:0 \rightarrow 1:1) to afford lycopladine E (1, 3.16 mg, 0.010 mmol).

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