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INDOLE ALKALOIDS FROM THE LEAVES OF *ALSTONIA SCHOLARIS*

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Abstract – A new indole alkaloid, akuammidine-*N*-oxide (**1**) was isolated from the leaves of *Alstonia scholaris* (Apocynaceae) together with akuammidine (**2**), and the structure was elucidated by NMR spectral analysis and chemical correlation. Akuammidine (**2**) showed a moderate antiplasmodial activity.

INTRODUCTION

The genus *Alstonia* and *Kopsia* (Apocynaceae), which are widely distributed throughout tropical Asia, are noted for producing variety of indole alkaloids with useful biological activities.¹ Recent investigation of extracts from the leaves of *Alstonia angustiloba*,² *Kopsia flavida*,³ and *Kopsia arborea*³ resulted in the isolation of some new indole alkaloids with antiplasmodial and vasorelaxant activities. In this paper, we report the isolation and structure elucidation of a new indole alkaloid, akuammidine-*N*-oxide (**1**) from the leaves of *Alstonia scholaris*, and antiplasmodial activity of isolated indole alkaloids.

The leaves of *A. scholaris* were extracted with MeOH, and the MeOH extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which were adjusted to pH 10 with saturated $Na₂CO₃$, were extracted with CHCl₃. Water-soluble materials were extracted with BuOH. BuOH-soluble materials were subjected to an ODS column $(H_2O/MeOH 1:1 \rightarrow 0:1)$ and the fractions eluted by H₂O/MeOH (1:1) were subjected to silica gel columns (CHCl₃/MeOH 1:0 \rightarrow 7:3 and CHCl₃/MeOH 1:0 \rightarrow 5:1) to afford compound **1** (0.001%). CHCl₃-soluble materials were subjected to

an LH-20 column (CHCl₃/MeOH = 1:1) and an amino silica gel column (hexane/EtOAc 8:1 \rightarrow 1:1 and then CHCl₃/MeOH 1:0 \rightarrow 0:1) to give akuammidine⁴ (2, 0.04%).

RESULTS AND DISCUSSION

Compound $1{(\alpha|_{D}^{23} - 39 (c \ 1.0, \text{MeOH})\}$ showed the pseudomolecular ion peak at m/z 369 (M+H)⁺ in ESIMS, and the molecular formula, $C_{21}H_{24}N_{2}O_{4}$, was established by HRESIMS [m/z 369.1828, (M+H)⁺ Δ +1.4 mDa]. IR spectrum suggested the presence of NH (3400 cm^{-1}) and ester carbonyl (1720 cm^{-1}) groups. The 13C NMR (Table 1) spectrum of **1** disclosed twenty-one carbon signals due to one ester carbonyl (δ_c 172.5), five *sp*² quaternary carbons (δ_c 138.7, 134.0, 131.8, 127.2, and 104.5), one *sp*³ quaternary carbon (δ_c 54.4), five sp^2 methines (δ_c 122.7, 120.9, 120.3, 119.0, and 112.3), three sp^3 methines (δ_c 72.8, 67.9, and 29.6), four *sp*³ methylenes (δ_c 71.6, 68.0, 31.7, and 20.9), one methyl (δ_c 13.1), and one methoxy group (δ_c 52.2). ¹H and ¹³C signals for **1** were assigned by detailed analysis of the HSQC spectrum. The ${}^{1}H$ - ${}^{1}H$ COSY spectrum revealed connectivities of C-5 to C-6, C-9 to C-12, C-3 to C-14, C-14 to C-15, and C-18 to C-19 (Figure 1). HMBC correlations of H-3 (δ_H 4.59) to C-2 (δ_C 134.0), H-5 (δ_H 3.17) to C-3 (δ_C 67.9) and C-7 (δ_C 104.5), H₂-6 (δ_H 3.51) to C-2 and C-7, H₂-14 (δ_H 2.41 and 2.96) to C-20 (δ_c 131.8), and H-15 (δ_H 3.35) to C-21 (δ_c 71.6) revealed the presence of a dehydroquinolizidine ring (C-2 to C-7, C-14 to C-15, C-20 to C-21, and N-4). Low field chemical shifts at C-3, C-5, and C-21 around N-4 atom $[\delta_C 67.9, 72.8,$ and 71.6, respectively] suggested that 1 was *N*-oxide form at N-4. The presence of the piperidine ring (C-5, C-3, C-14 to C-16, and N-4) was deduced from the HMBC correlations of $H₂$ -14 to C-16, and $H₂$ -6 to C-16. Connection between indole ring (C-2, C-7 to C-13 and N-1) and dehydroquinolizidine ring was deduced from the HMBC correlations of H-9 (δ_H 7.44) to C-7. Structure of methyl 3-hydroxypropanoate moiety (C-16 to C-17, C-22, and C-23) was elucidated from HMBC correlations of H-5 to C-17 and C-22 (δ_H 172.5), H-15 to C-17, and H_3 -23 (δ_H 2.98) to C-22. The HMBC cross-peaks of H_3 -18 (δ_H 1.73) to C-20 indicate the ethylidene side chain at C-20. Thus, the gross structure of **1** was assigned as sarpagine-type skeletal system with an N-4 oxide.

Figure 1. Selected 2D NMR correlations of **1**.

Figure 2. Selected NOESY correlations and relative stereochemistry for **1**

The relative stereochemistry of **1** was elucidated by NOESY correlations as shown in computer-generated 3D drawing (Figure 2). Configuration of C-16 was elucidated by NOESY correlations among H_3 -23, H-6, and H-9, and between H_2 -17 and H-5. NOESY correlations of H-19 to H-21 indicated the geometry of C-19 - C-20 double bond was *E*. Thus, the relative stereochemistry of **1** was assigned as shown in Figure 2.

Treatment of compound 1 with $Na₂SO₃$ in aqueous MeOH afforded the reductive derivative, whose spectroscopic data and specific rotation were identical with akuammidine (2) .⁴ Thus, the absolute configuration of **1** was assigned as the same as **2**.

Malaria caused by parasites of the genus *Plasmodium* is one of the leading infectious diseases in many tropical and some temperate regions.⁵ The emergence of widespread chloroquine-resistant and multiple-drug-resistant strains of malaria parasites leads to the need for the development of new therapeutic agents against malaria.⁶ Since some indole alkaloids from *Alstonia* species have already been reported on inhibitory activity against some parasites. 7 Akuammidine (**2**) showed a moderate *in vitro* antiplasmodial activity against *Plasmodium falciparum* (**2**: 32.6% inhibition at 10 µg/mL), whereas akuammidine-*N*-oxide (1) did not show $(>10 \mu g/mL)$.⁸

Position	$\delta_{\rm H}$	$\delta_{\rm c}$
2		134.0
\mathfrak{Z}	4.59 (d, 10.7)	67.9
5	3.17 (t, 29.6)	72.8
6a	3.51 (brd, 13.7)	20.9
6b	3.51 (brd, 13.7)	
τ		104.5
8		127.2
9	7.44 (d, 7.9)	119.0
10	7.03 (t, 7.2)	120.3
11	7.10 (t, 7.2)	122.7
12	7.32 (d, 7.9)	112.3
13		138.7
14a	2.41 (d, 10.0)	31.7
14 _b	2.96 (d, 10.0)	
15	3.35 (d, 11.5)	29.6
16		54.4
17a	3.70 (d, 9.6)	68.0
17 _b	3.82 (d, 9.6)	
18	1.73 (d, 6.9)	13.1
19	5.55 (q, 6.9)	120.9
20		131.8
21a	4.00 (d, 15.8)	71.6
21 _b	4.45 (d, 15.8)	
22		172.5
23	2.98(s)	52.2

Table 1. ¹H and ¹³C NMR Data [δ_H (J, Hz) and δ_C] of akuammidine-*N*-oxide (1) in CD₃OD at 300K

EXPERIMENTAL

General Experimental Procedures. ¹H and 2D NMR spectra were recorded on a JEOL ECA600 spectrometer and chemical shifts were reported using residual CD₃OD (δ_H 3.31 and δ_C 49.0) as internal standards. HSQC experiments were optimized for J_{CH} =145 Hz and HMBC experiments for J_{CH} =8Hz. Mass spectra were recorded on a Micromass LCT spectrometer.

Plant Material. The leaves of *Alstonia scholaris* were collected in Purwodadi Botanical Garden, East Java, Indonesia in 2006. A voucher specimen is deposited at the Purwodadi Botanical Garden, Indonesia.

Extraction and Isolation. The leaves of *A. scholaris* (500 g) were extracted with MeOH, and the MeOH extract was partitioned between EtOAc and 3% tartaric acid. Water-soluble materials, which were adjusted to pH 10 with saturated aqueous $Na₂CO₃$, were extracted with CHCl₃. Water-soluble materials were extracted with BuOH. BuOH-soluble materials were subjected to an ODS column

 $(H₂O/MeOH 1:1 \rightarrow 0:1)$ and the fractions eluted by $H₂O/MeOH (1:1)$ were subjected to silica gel columns (CHCl₃ /MeOH, 1:0 \rightarrow 7:3 and CHCl₃/MeOH, 1:0 \rightarrow 5:1) to afford compound **1** (0.001%). $CHCl₃$ -soluble materials were subjected to an LH-20 column (CHCl₃/MeOH, 1:1) and an amino silica gel column (hexane/EtOAc, $8:1 \rightarrow 1:1$ and then CHCl₃/MeOH, $1:0 \rightarrow 0:1$) to give akuammidine⁴ (2, 0.04%).

Akuammidine-*N***-oxide (1):** colorless amorphous solid; $[\alpha]_D^2$ ³ -39 (*c* 1.0, MeOH); IR (film) v_{max} 3400, 2940, 2360, 1720, and 1630 cm⁻¹; UV (MeOH) λ_{max} 270 (ε 1000) and 205 (ε 5000) nm; ¹H and ¹³C NMR data (Table 1); ESIMS m/z 369 (M+H)⁺; HRESITOFMS m/z 369.1828 [(M+H)⁺, calcd for $C_{21}H_{25}N_{2}O_{4}$, 369.1814].

Chemical conversion of akuammidine-*N***-oxide (1) into akuammidine (2).** To a solution of akuammidine-*N*-oxide $(1, 0.2 \text{ mg})$ in aqueous MeOH (0.2 mL) was added Na₂SO₃ (1.0 mg) and the mixture was kept at rt for 30 min. After evaporation, the residue was applied to a silica gel column (CHCl₃/MeOH, 4:1) to give a compound (0.15 mg), whose spectroscopic data and $[\alpha]_D$ value were identical with those of natural akuammidine (**2**).

Antiplasmodial Assay. Human malaria parasites were cultured according to the method by W. Trager et al.⁸ The antimalarial activity of the isolated compounds was determined by the procedure described by Budimulja et al.⁸ In brief, Stock solution of the samples were prepared in DMSO (final DMSO concentrations of $\langle 0.5\% \rangle$ and were diluted to the required concentration with complete medium (RPMI) 1640 supplemented with 10% human plasma, 25 mM HEPES and 25 mM NaHCO₃) until the final concentration of samples at well culture plate were: 10; 1; 0.1; 0.01; 0.001 µg/mL. The malarial parasite *P. falciparum* 3D7 clone was propagated in a 24-well culture plate in the presence of a wide range of concentrations of each compound. The growth of the parasite was monitored by making a blood smear fixed with MeOH and stained with Geimsa solution. The antimalarial activity of each compound was expressed as an IC₅₀ value, defined as the concentration of the compound causing 50% inhibition of parasite growth relative to an untreated control.

The percentage of growth inhibition was expressed according to following equation : Growth inhibition % = 100 - [(test parasitaemia/control parasitemia) \times 100. Chloroqine: IC₅₀= 0.0061 µg/mL.

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REFERENCES

- 1. (a) P. Kamarajan, N. Sekar, V. Mathuram, and S. Govindasamy, *Biochem. Int*., 1991, **25**, 491; (b) V. saraswathi, S. Subramanian, N. Ramamoorth, V. Mathuram, and S. Govindasamy, *Med. Sci. Res*., 1997, **25**, 167; (c) N. Keawpradub, G. C. Kirby, J. C. P. Steele, and P. J. Houghton, *Planta Med*., 1999, **65**, 690; (d) K. Husain, I. Jantan, N. Kamaruddin, I. M. Said, N. Aimi, and H. Takayama, *Phytochemistry*, 2001, **57**, 603; (e) K. Husain, I. Jantan, I. M. Said, N. Aimi, and H. Takayama, *J. Asian Nat. Prod. Res*., 2003, **5**, 63; (f) G. C. Jagetia and M. S. Baliga, *Phytother Res*., 2006, **20**, 103; (g) M. R. Khan, A. D. Omoloso, and M. Kihara, *Fitoterapia*, 2003, **74**, 736; (h) R. S. Gupta, A. K. Bhatnager, Y. C. Joshi, M. C. Sharma, V. Khushalani, and J. B. Kachhawa, *Pharmacology*, 2005, **75**, 57.
- 2. K. Koyama, Y. Hirasawa, K. Zaima, T. C. Hoe, K. L, Chan, and H. Morita, *Bioorg. Med. Chem*., 2007, **16**, 6483.
- 3. (a) M. Sekiguchi, Y. Hirasawa, K. Zaima, T. C. Hoe, K.-L. Chan, and H. Morita, *Heterocycles*, 2008, **75**, 2283; (b) M. Sekiguchi, Y. Hirasawa, K. Zaima, T. C. Hoe, K.-L. Chan, and H. Morita, *Heterocycles*, 2008, **76**, 867; (c) K. Zaima, Y. Matsuno, Y. Hirasawa, A. Rahman, G. Indrayanto, N. C. Zaini, and H. Morita, *Heterocycles*, 2008, **75**, 2535.
- 4. W. Boonchuay and W. E. Court, *Planta Med*., 1976, **29**, 380.
- 5. J. Wiesner, R. Ortmann, H. Jomaa, and M. Schlitzer, *Angew. Chem. Int. Ed*., 2003, **42**, 5274.
- 6. M. H. Gelb and W. G. Hol, *Science*, 2002, **297**, 343.
- 7. T. Yamauchi and F. Abe, International Congress Series 1157 (Towards Natural Medicine Research in the 21^{st} Century): 51-58.
- 8. (a) W. Trager and J. B. Jensen, *Science*, 1976, **193**, 673; (b) A. S. Budimulja, Syafruddin, P. Tapchaisri, P. Wilairat, and S. Marzuki, *Mol. Biochem. Parasitol*., 1997, **84**, 137.