

HETEROCYCLES, Vol. 79, 2009, pp. 427 - 432. © The Japan Institute of Heterocyclic Chemistry  
Received, 10th October, 2008, Accepted, 5th December, 2008, Published online, 11th December, 2008.  
DOI: 10.3987/COM-08-S(D)70

## SYNTHETIC STUDIES ON THE FLUORINATED ANALOGS FOR THE PUTATIVE OXINDOLE-TYPE METABOLITES OF 5-HALOTRYPTAMINES

Tomoya Fujiwara, Takayuki Seki, Masaru Miura, and Yoshio Takeuchi\*

Graduate School of Medicine and Pharmaceutical Sciences for Research,  
University of Toyama, Sugitani 2630, Toyama 930-0194, Japan  
takeuchi@pha.u-toyama.ac.jp

**Abstract** – The suitably protected precursors for direct fluorination, *N*<sup>b</sup>-Boc di-protected 5-fluorotryptamine (**13**), *N*<sup>b</sup>-acetyl-*N*<sup>b</sup>-Boc protected 5-halotryptamines (**15a–c**), were treated with Selectfluor<sup>TM</sup> in MeCN/water in the presence of NaHCO<sub>3</sub> to give the corresponding 3-fluorooxindoles **14** and **16a–c** in good yields. Removal of the protecting groups of **14** and **16a–c** produced (3,5-difluorooxindol-3-yl)ethylamine (**8**) and *N*-acetyl-(3-fluoro-5-halooxindol-3-yl)ethylamines (**9a–c**) in excellent yields, respectively. These compounds are potentially non-epimerizable analogs for the putative metabolites of 5-fluorotryptamine (**6**) and *N*<sup>b</sup>-acetyl-5-halotryptamines (**7a–c**).

## INTRODUCTION

The 3-fluorooxindole derivatives **1** have received much attention as a synthetic target for development of novel medicinal agents since BMS-204532 (Maxipost, **2**)<sup>1</sup> was discovered as an effective calcium-dependent potassium channel opener. Fluorinated biomolecules and medicinals can be recognized by enzymes and receptors as well as the nonfluorinated molecules because the replacement of a hydrogen with a fluorine brings about the minimal steric alterations of the molecules.<sup>2</sup> It should be noted that the fluorinated derivatives can be non-epimerizable analogs when the labile proton at the stereogenic center is replaced by a fluorine. Indeed we previously reported the synthesis of 3'-fluorothalidomide as a non-epimerizable analog of thalidomide.<sup>3</sup> Since the stereoelectronic properties of fluorine are also similar to those of hydroxy group, isosteric analogs of prototype molecules can be

† Dedicated to the memory of Dr. John Daly who made many lasting contributions to the wide area of chemistry.

obtained by substituting the hydroxy group with fluorine.<sup>2</sup> Moreover, such substitution will prevent further metabolism which would be subjected otherwise.<sup>2</sup> Thus 3-fluorooxindole derivatives **1** can be employed as mimics for both oxindoles **3** and 3-hydroxyoxindoles **4**, which are often found in natural products and as metabolites of some indole-containing biomolecules. As a part of our studies on the design, synthesis, and biological evaluation of chiral fluorinated bioorganic molecules,<sup>3,4</sup> we synthesized 3-(3-fluorooxindol-3-yl)-L-alanine (**5**).<sup>5</sup> Here we report the synthesis of (3,5-difluorooxindol-3-yl)ethylamine (**8**) and *N*-acetyl-(3,5-difluorooxindol-3-yl)ethylamine (**9a–c**) as potential analogs for the putative oxindole-type metabolites of 5-fluorotryptamine (**6**) and *N*<sup>b</sup>-acetyl-5-halotryptamines (**7a–c**), which have agonistic activities toward serotonin 5-HT<sub>3</sub> receptor and melatonin MT<sub>1</sub> and MT<sub>2</sub> receptors, respectively.<sup>6</sup> It should be noted that oxindoles **3** may epimerize under physiological conditions.<sup>7</sup> Therefore, compounds **8** and **9a–c** can be the non-epimerizable analogs, which are effective tools for investigating the relationship between the biological behavior and the stereochemistry of the metabolites.

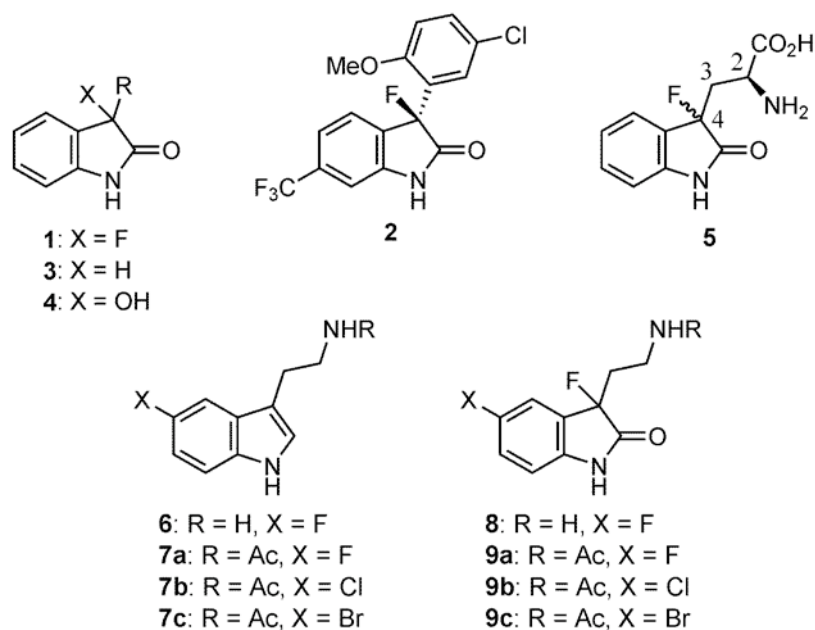


Figure 1. Structures of compounds **1–9**.

## RESULTS AND DISCUSSION

We previously reported the direct fluorination using Selectfluor<sup>TM</sup> for preparation of the tryptophan isostere **5**. In this procedure we found that use of the precursor with di-protected  $\alpha$ -amino moiety is essential for successful fluorination. For synthesis of the fluorooxindole-containing ethylamine **8**, we employed *N*<sup>b</sup>,*N*<sup>b</sup>-di-(*tert*-butoxycarbonyl)tryptamine (**10**) as a model compound in order to examine the optimum reaction conditions for the fluorination. The suitably protected precursor **10**, prepared from

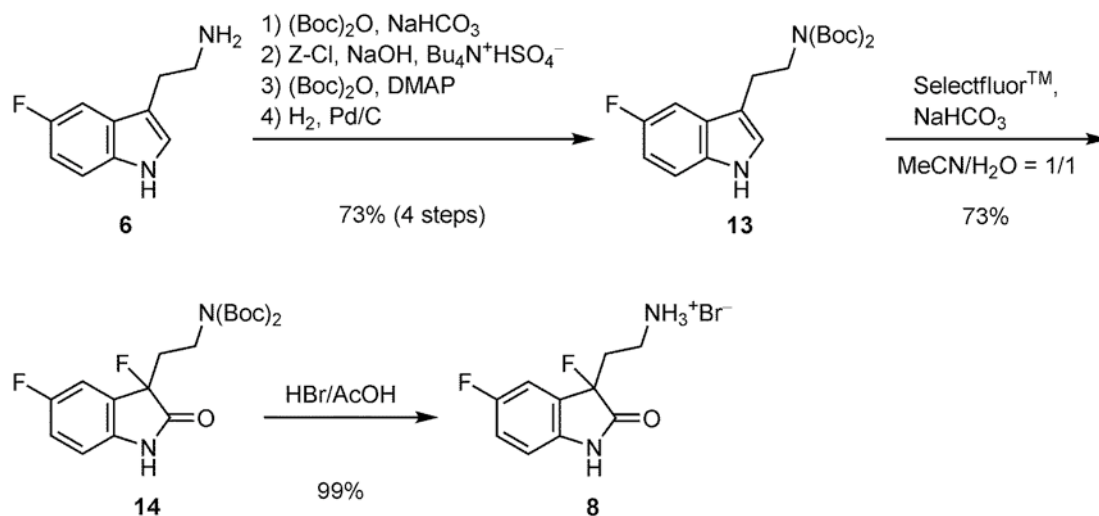
tryptamine according to the literature,<sup>5,8</sup> was treated with 3 equiv of Selectfluor<sup>TM</sup> in MeCN/water (1/1) at rt for 16–24 h<sup>9</sup> to produce the corresponding 3-fluorooxindole **11**. However, the yield of **11** was quite low owing to the formation of mono-deprotected product **12** (Table 1, entry 1). This was presumably due to the rather acidic conditions employed for the reaction. We then attempted the fluorination in the presence of a base.<sup>10</sup> Fluorination of **10** in the presence of 3 equiv of NaHCO<sub>3</sub> gave 3-fluorooxindole **11** in 58% yield without formation of **12** (entry 2). Use of increased or decreased amount of NaHCO<sub>3</sub> did not give any better results (entries 3–6). Excess amount of Selectfluor<sup>TM</sup> led to increase of the yield of **11** (entry 7). The best result was obtained when the fluorination was carried out using 4 equiv of Selectfluor<sup>TM</sup> and 4 equiv of NaHCO<sub>3</sub> to afford **11** in 73% yield (entry 7).

Table 1. Fluorination of *N*<sup>b</sup>,*N*<sup>b</sup>-di-(*tert*-butoxycarbonyl)tryptamine (**10**)

Entry	Selectfluor <sup>TM</sup> (equiv)	NaHCO <sub>3</sub> (equiv)	Yield of <b>11</b> (%) <sup>a</sup>	Yield of <b>12</b> (%) <sup>a</sup>
1	3	0	20	51
2	3	3	58	0
3	3	0.5	43	29
4	3	1.5	56	11
5	3	4.5	47	0
6	3	6	38	0
7	4	4	73	0

<sup>a</sup>Isolated yield.

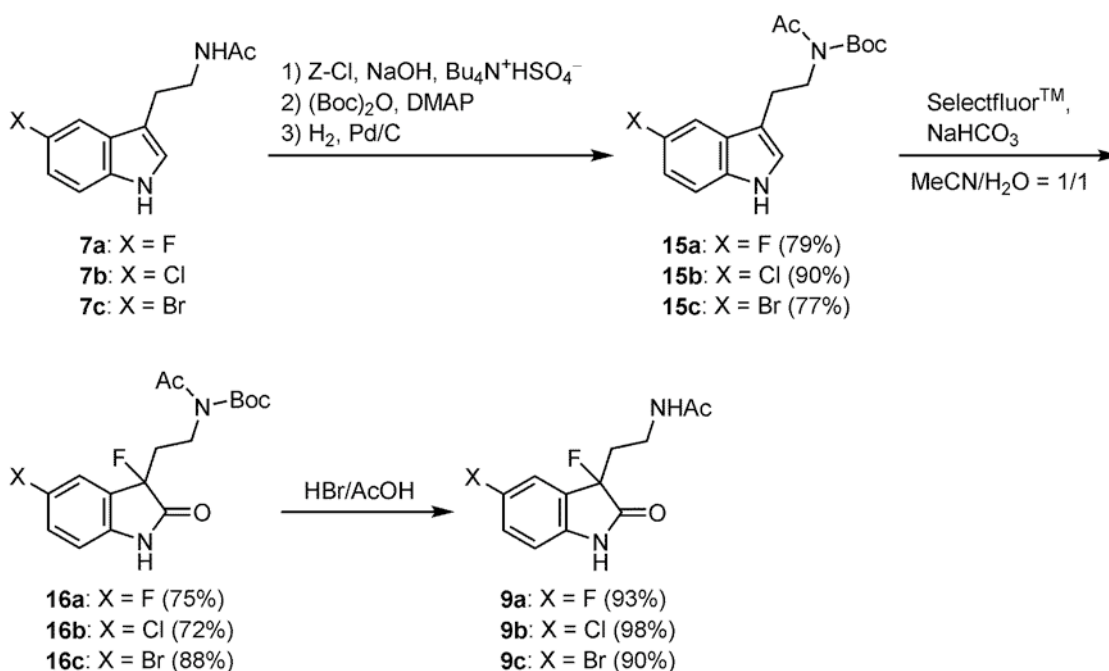
The 5-fluoro analog **13** was prepared from **6** according to the literature.<sup>5,8</sup> We employed above reaction conditions for direct fluorination of **13** and the corresponding 3-fluorooxindole **14** was successfully obtained in 73% yield (Scheme 1).<sup>11</sup> Treatment of **14** with HBr/AcOH smoothly produced the deprotected ethylamine **8** as HBr salt in excellent yield.<sup>12</sup>



Scheme 1. Synthesis of (3,5-difluorooxindol-3-yl)ethylamine (**8**)

We next attempted synthesis of *N*<sup>b</sup>-acetyl protected fluorooxindole-containing ethylamine **9a–c**. Although direct fluorination of **7b** with Selectfluor<sup>TM</sup> gave **9b**, the yield was unsatisfactory owing to concomitant formation of side products.<sup>13</sup> We then focused on the *N*<sup>b</sup>-di-protected structure. The precursors **15a–c**, prepared in a similar manner to the synthesis of **13**, were treated with Selectfluor<sup>TM</sup> in the presence of NaHCO<sub>3</sub> to give the corresponding 3-fluorooxindoles **16a–c** in good yields (Scheme 2). Removal of the Boc groups of **16a–c** with HBr/AcOH successfully produced the *N*<sup>b</sup>-acetyl protected fluorooxindole-containing ethylamine **9a–c** in excellent yields.<sup>14</sup>

Preparation of the individual enantiomers of **8** and **9a–c** is in progress for investigation of the relationship between the biological behavior and the stereochemistry.



Scheme 2. Synthesis of *N*-acetyl-(3-fluoro-5-halo)oxindol-3-ylethylamines (**9a–c**)

In summary, we achieved the synthesis of fluorooxindole-containing ethylamine **8** as a metabolic analog of **6** by fluorination of the  $N^b$ -di-protected tryptamine derivative **13** with Selectfluor<sup>TM</sup> in the presence of NaHCO<sub>3</sub> followed by deprotection. Similarly,  $N^b$ -acetyl protected fluorooxindole-containing ethylamine **9a–c** were successfully synthesized as metabolic analogs of **7a–c**.

## REFERENCES AND NOTES

1. P. Hewawasam, V. K. Gribkoff, Y. Pendri, S. I. Dworetzky, N. A. Meanwell, E. Martinez, C. G. Boissard, D. J. Post-Munson, J. T. Trojnacki, K. Yeleswaram, L. M. Pajor, J. Knipe, Q. Gao, R. Perrone, and J. E. Starrett, Jr., *Bioorg. Med. Chem. Lett.*, 2002, **12**, 1023.
2. a) J. Bégué and D. Bonnet-Delpon, 'Bioorganic and Medicinal Chemistry of Fluorine', John Wiley & Sons, New York, 2008; b) I. Ojima, J. R. McCarthy, and J. T. Welch, 'Biomedical Frontiers of Fluorine Chemistry', ACS Symposium Series 639, American Chemical Society, Washington, DC, 1996; c) R. Filler and Y. Kobayashi, 'Biomedical Aspects of Fluorine Chemistry', Kodansha/Elsevier Biomedical, Tokyo, 1982.
3. Y. Takeuchi, T. Shiragami, K. Kimura, E. Suzuki, and N. Shibata, *Org. Lett.*, 1999, **1**, 1571.
4. a) Y. Takeuchi, N. Shibata, E. Suzuki, Y. Imura, T. Kosasa, T. Yamanishi, and H. Sugimoto, PCT Int. Appl. WO 2002020482; b) Chem. Abstr. 2002, **136**, 247496v; c) H. Fujisawa, T. Fujiwara, Y. Takeuchi, and K. Omata, *Chem. Pharm. Bull.*, 2005, **53**, 524; d) Y. Takeuchi, H. Fujisawa, T. Fujiwara, M. Matsuura, H. Komatsu, S. Ueno, and T. Matsuzaki, *Chem. Pharm. Bull.*, 2005, **53**, 1062.
5. T. Fujiwara, B. Yin, M. Jin, K. L. Kirk, and Y. Takeuchi, *J. Fluorine Chem.*, 2008, **129**, 829.
6. a) K. S. Bower, K. L. Price, L. E. C. Sturdee, M. Dayrell, D. A. Dougherty, and S. C. R. Lummis, *Eur. J. Pharmacol.*, 2008, **580**, 291; b) G. Spadoni, C. Balsamini, A. Bedini, A. Carey, G. Diamantini, B. D. Giacomo, A. Tontini, G. Tarzia, R. Nonno, V. Lucini, M. Pannacci, B. M. Stankov, and F. Frascini, *Med. Chem. Res.*, 1998, **8-9**, 487.
7. a) R. S. Phillips, E. W. Miles, and L. A. Cohen, *Biochemistry*, 1984, **23**, 6228; b) Idem, *J. Biol. Chem.*, 1985, **260**, 14665; c) D. M. Kiick and R. S. Phillips, *Biochemistry*, 1988, **27**, 7339; d) M. Roy, E. W. Miles, R. S. Phillips, and M. F. Dunn, *ibid.*, 1988, **27**, 8661; e) R. S. Phillips, S. L. Bender, P. Brzovic, and M. F. Dunn, *ibid.*, 1990, **29**, 8608; f) R. B. Labroo and L. A. Cohen, *J. Org. Chem.*, 1990, **55**, 4901.
8. Y. Kiso, M. Inai, K. Kitagawa, and T. Akita, *Chem. Lett.*, 1983, 739.
9. Y. Takeuchi, T. Tarui, and N. Shibata, *Org. Lett.*, 2000, **2**, 639.
10. T. Umemoto, M. Nagayoshi, K. Adachi, and G. Tomizawa, *J. Org. Chem.*, 1998, **63**, 3379.
11. General procedure for the fluorination of the protected tryptamines: Selectfluor<sup>TM</sup> (283.4 mg, 0.800 mmol) was added to a solution of  $N^b, N^b$ -di-(*tert*-butoxycarbonyl)-5-fluorotryptamine (**13**) (75.7 mg,

- 0.200 mmol) in a mixture of MeCN and water (1:1, 4 mL) at rt. After stirring for 20 h, the mixture was concentrated and extracted with EtOAc. The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica gel column chromatography (eluent; hexane/EtOAc = 3/1) to give the corresponding 3-fluorooxindole **14** (60.5 mg, 0.147 mmol, 73%) as a colorless glass, together with the corresponding oxindole derivative (4.1 mg, 0.010 mmol, 5%) as a colorless oil: IR (KBr)  $\nu$  3430, 3315, 2979, 2933, 1759, 1734, 1701, 1676, 1631, 1487 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.48 (18H, s), 2.35–2.50 (2H, m), 3.71–3.78 (2H, m), 6.88 (1H, ddd,  $J$  = 8.5, 4.1, 0.9 Hz), 7.05 (1H, tt,  $J$  = 9.2, 2.3 Hz), 7.16 (1H, dt,  $J$  = 7.3, 2.3 Hz), 8.62 (1H, br s); <sup>19</sup>F NMR (376 MHz, CDCl<sub>3</sub>)  $\delta$  -119.46 (1F, dddd,  $J$  = 9.2, 7.3, 4.1, 1.8 Hz), -159.14 (1F, t,  $J$  = 16.2 Hz); MS (EI)  $m/z$ : 412 (M<sup>+</sup>), 356 (M<sup>+</sup>-C<sub>4</sub>H<sub>8</sub>), 300 (M<sup>+</sup>-C<sub>8</sub>H<sub>16</sub>); HRMS (EI) calcd for C<sub>20</sub>H<sub>26</sub>F<sub>2</sub>N<sub>2</sub>O<sub>5</sub> (M<sup>+</sup>): 412.1810; found 412.1794.
12. Synthesis of **8**: To a solution of **14** (23.7 mg, 0.058 mmol) in AcOH (0.5 mL) was added 25% HBr/AcOH (0.5 mL) at 0 °C. After stirring at rt for 10 min, the mixture was concentrated in vacuo to give HBr salt of **8** (16.7 mg, 0.057 mmol, 99%) as a pale yellow solid: mp 136 °C (decomp.); IR (KBr)  $\nu$  3700–3100 (br), 3215, 3033, 1739, 1634, 1490 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  2.31 (1H, dddd,  $J$  = 27.0, 14.9, 7.8, 6.9 Hz), 2.58 (1H, dddd,  $J$  = 14.9, 13.5, 7.8, 6.9 Hz), 3.29 (2H, br m), 6.94 (1H, ddd,  $J$  = 8.5, 4.1, 1.4 Hz), 7.17 (1H, dddd,  $J$  = 9.2, 8.7, 2.7, 1.8 Hz), 7.31 (1H, dt,  $J$  = 7.8, 2.7 Hz); <sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>OD)  $\delta$  -119.67 (1F, dddd,  $J$  = 9.2, 7.8, 4.1, 1.8 Hz), -160.00 (1F, m); MS (FAB<sup>+</sup>)  $m/z$ : 213 (M<sup>+</sup>-Br); HRMS (FAB<sup>+</sup>) calcd for C<sub>10</sub>H<sub>11</sub>F<sub>2</sub>N<sub>2</sub>O (M<sup>+</sup>-Br): 213.0840; found 213.0854, (FAB<sup>-</sup>) calcd for C<sub>10</sub>H<sub>10</sub>BrF<sub>2</sub>N<sub>2</sub>O (M<sup>+</sup>-H): 290.9945; found 290.9942.
13. The N<sup>b</sup>-acetyl moiety will be subjected to electrophilic fluorination thus giving unidentified oxidized products.
14. General procedure for the acid deprotection of the Boc group of **16a–c**: To a solution of **16a** (19.9 mg, 0.056 mmol) in AcOH (1 mL) was added 25% HBr/AcOH (1 mL) at 0 °C. After stirring for 10 min at rt, the mixture was poured into saturated aqueous NaHCO<sub>3</sub> and then extracted with EtOAc. The organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica gel column chromatography (eluent; EtOAc/MeOH = 40/1) to give **9a** (13.2 mg, 0.052 mmol, 93%) as a colorless solid: mp 170–172 °C; IR (KBr)  $\nu$  3441, 3353, 1732, 1722, 1653, 1636, 1554, 1489 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD)  $\delta$  1.82 (3H, s), 2.34 (2H, dt,  $J$  = 15.1, 7.3 Hz), 3.20 (1H, dt,  $J$  = 13.7, 7.3 Hz), 3.25 (1H, dt,  $J$  = 13.7, 7.3 Hz), 6.90 (1H, dd,  $J$  = 7.8, 3.7 Hz), 7.11 (1H, tt,  $J$  = 8.7, 1.8 Hz), 7.27 (1H, dt,  $J$  = 7.8, 1.8 Hz); <sup>19</sup>F NMR (376 MHz, CD<sub>3</sub>OD)  $\delta$  -120.28 (1F, m), -155.90 (1F, t,  $J$  = 15.3 Hz); MS (EI)  $m/z$ : 254 (M<sup>+</sup>), 192 (M<sup>+</sup>-F-CH<sub>3</sub>CO); HRMS (EI) calcd for C<sub>12</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>O<sub>2</sub> (M<sup>+</sup>): 254.0867; found 254.0869.