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NEW XANTHONES FROM THE BARKS OF CRATOXYLUM

SUMATRANUM SSP. NERIIFOLIUM

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Abstract – Three new prenylated xanthones, neriifolone A-C (1-3), and eight known xanthones (4-11) were isolated from the barks of *Cratoxylum sumatranum* ssp. *neriifolium*. All the new compounds were characterized by intensive spectroscopic methods (1D and 2D NMR, UV and IR spectroscopy and mass spectrometry).

Cratoxylum is a small genus belonging to the Clusiaceae, which is distributed in several Asian countries. Some species of this genus have been used in folk medicine as a treatment for diarrhea, flatulence¹ as well as diuretic, stomachic and tonic complaints.² Xanthone is the major chemical substance present in this genus³⁻¹¹ and some of these chemical substances showed interesting biological activities.^{4,6-8,10,11} In this paper, we describe the isolation and structural elucidation of three new xanthones (**1-3**) together with eight known xanthones (**4-11**) from the barks of *C. sumatranum* ssp. *neriifolium*.

The dichloromethane extract from the barks of *C. sumatranum* spp. *neriifolium* was separated by chromatographic techniques to afford eleven compounds: three new xanthones, neriifolone A (1), neriifolone B (2) and neriifolone C (3), and eight known xanthones (4-11). The structures of all of the new compounds were then elucidated using extensive 1D and 2D NMR spectroscopic techniques.



Figure 1. Compounds isolated from C. sumatranum spp. neriifolium

Neriifolone A (1) was obtained as a yellow solid. The molecular formula, $C_{19}H_{19}O_6$, was determined by HRMS (ESI-TOF) ($[M+H]^+$ m/z 343.1194). The UV spectrum showed absorption bands of the xanthone skeleton⁸⁻¹¹ at 246, 256, 320 and 354 nm whereas the IR spectrum showed the stretches for the conjugated carbonyl and hydroxyl functionalities at 1647 and 3392 cm⁻¹, respectively. The ¹H NMR spectrum of **1**

(Table 1) exhibited a signal for a hydrogen bonded hydroxyl group (1-OH) at δ 13.44 and three aromatic protons at δ 7.81 (1H, *d*, *J* = 8.7 Hz), 7.03 (1H, *d*, *J* = 8.7 Hz) and 6.32 (1H, *s*) which were assigned to H-8, H-7 and H-2, respectively. Moreover, a methoxy group at δ 3.95 (3H, *s*) and a 1,1-dimethylallyl unit at δ 6.45 (1H, *dd*, *J* = 17.4, 10.5 Hz, H-4'), 4.96 (1H, *dd*, *J* = 17.4, 0.9 Hz, Ha-5') 4.89 (1H, *dd*, *J* = 10.5, 0.9 Hz, Hb-5') and 1.77 (6H, *s*, Me-2' and Me-3') were also observed in the ¹H NMR spectrum. The presence of a 1,1-dimethylallyl unit on C-4 was confirmed by the HMBC correlations (Figure 2) between Me-2' (δ 1.77), Me-3' (δ 1.77) and H-4' (δ 6.45) to C-4 (δ 112.8). The assignment of the methoxy group on C-5 was based on the cross peak of the methoxy protons (δ 3.95) and H-7 (δ 7.03) with C-5 (δ 136.0) in the HMBC experiment.



Figure 2. NOE cross peaks of 2 and HMBC correlations of 1, 2 and 3

Neriifolone B (2) was isolated as a yellow solid. The HRMS (ESI-TOF) gave a molecular ion peak at m/z 379.0789 ([M+Na]⁺) which was consistent with the molecular formula C₁₉H₁₆O₇. The UV and IR spectra of 2 exhibited the same pattern as that of 1. However, the IR spectrum of 2 also showed an additional carbonyl absorption peak at 1780 cm⁻¹. The ¹H and ¹³C NMR spectral data of 2 (Table 1) were similar to those of 1 except for the disappearance of the 1,1-dimethylallyl unit. Compound 2 exhibited signals for the

3,3-dimethyl- δ -lactone ring which showed ¹H and ¹³C NMR signals at δ 1.69/28.0 (Me-2' and Me-3'), 2.85/45.1 (H/C-4'), 34.8 (C-1') and 158.1 (C-3), 110.9 (C-4) and 167.0 (C-5'). Based on the HMBC correlations between Me-2' and Me-3' (δ 1.69) and H-4' (δ 2.85) to C-4 (δ 110.9), this unit was placed at C-3 and C-4 of the xanthone skeleton. The cross peak between Me-2' and Me-3' (δ 1.69) with 5-OMe (δ 4.01) in the NOE experiment also supported this placement.

Position	1		2		3	
	$\delta_{\rm H}$ (mult, J in Hz)	$\delta_{ m C}$	δ_{H} (mult, J in Hz)	$\delta_{ m C}$	$\delta_{\rm H}$ (mult, J in Hz)	$\delta_{ m C}$
1	-	162.6	-	162.8	-	162.1
2	6.32 (s)	100.1	6.41 (s)	100.2	6.19 (<i>s</i>)	100.0
3	-	164.8	-	158.1	-	161.6
4	-	112.8	-	110.9	-	110.6
4a	-	157.0	-	155.5	-	156.7
5	-	136.0	-	135.9	-	147.4
6	-	157.6	-	158.0	7.41 (<i>dd</i> , 8.1, 0.6)	120.9
7	7.03 (<i>d</i> , 8.7)	114.7	7.09 (<i>d</i> , 9.0)	115.4	7.28 (<i>t</i> , 8.1)	124.8
8	7.81 (<i>d</i> , 8.7)	121.9	7.85 (<i>d</i> , 9.0)	122.3	7.68 (<i>dd</i> , 8.1, 0.6)	116.1
8a	-	114.7	-	114.7	-	122.3
9	-	181.4	-	181.7	-	182.1
9a	-	103.8	-	106.3	-	105.1
10a	-	151.5	-	151.9	-	146.3
1′	-	42.0	-	34.8	2.03 (dd, 13.5, 2.4, trans-H)	46.8
					1.94 (dd, 13.5, 7.8, cis-H)	
2'	1.77 (s)	30.3	1.69 (s)	28.0	5.56 (<i>dd</i> , 7.8, 2.4)	94.0
3'	1.77 (s)	30.3	1.69 (s)	28.0	-	109.2
4′	6.45 (dd, 17.4, 10.5)	151.2	2.85 (s)	45.1	$^{i}1.75(s)$	32.8
5'	4.96 (<i>dd</i> , 17.4, 0.9, Ha)	109.2	-	167.0	i 1.64 (s)	32.8
	4.89 (<i>dd</i> , 10.5, 0.9, Hb)					
1-OH	13.44 (s)	-	13.28 (s)	-	12.98 (s)	-
5-OMe	3.95 (s)	61.9	4.01 (s)	61.9	-	-

Table 1. ¹H- and ¹³C-NMR spectral data of 1, 2 and 3 in acetone- d_6

^{*i*} Interchangeable

Neriifolone C (**3**) was isolated as a yellow solid. The molecular formula was determined as $C_{18}H_{17}O_6$ by HRMS (ESI-TOF) ([M+H]⁺ m/z 329.1018). The UV and IR spectra confirmed that **3** has a xanthone skeleton. Compound **3** exhibited the following ¹H NMR (Table 1) signals for the xanthone nucleus. A chelated hydroxyl proton was located at 12.98 ppm. A singlet aromatic proton signal at δ 6.19 which showed ²J and ³J HMBC correlations (Figure 2) with C-1 (δ 162.1), C-4 (δ 110.6) and C-9a (δ 105.1) was assigned as the isolated proton H-2. The ABM system of spectrum at δ 7.68 (dd, J = 8.1, 0.6 Hz), 7.41 (dd, J = 8.1, 0.6 Hz) and 7.28 (t, J = 8.1 Hz) and the ²J or ³J HMBC correlations of H-6 (δ 7.41) with C-8 (δ 116.1) and C-10a (δ 146.3); H-7 (δ 7.28) with C-5 (δ 147.4) and C-8a (δ 122.3) and H-8 (δ 7.68) with C-6

(δ 120.9), C-9 (δ 182.1) and C-10a (δ 146.3) were assigned as aromatic protons H-8, H-6 and H-7, respectively. Also, a characteristic signal of a 2-(1-hydroxy-1-methylethyl)-2,3-dihydrofuran moiety at δ 1.75 (*s*, H-4'), 1.64 (*s*, H-5'), 5.56 (*J* = 7.8, 2.4 Hz, H-2'), 2.03 (*J* = 13.5, 2.4 Hz, *trans*-H) and δ 1.94 (*J* = 13.5, 7.8 Hz, *cis*-H) and the HMBC correlations of H-1' (δ 2.03 and 1.94) with C-4 (δ 110.6), confirmed that a 2-(1-hydroxy-1-methylethyl)-2,3-dihydrofuran moiety was fused to the xanthone nucleus at the C-3 and C-4 positions.

The remaining compounds were identified as β -mangostin (4),¹² pancixanthone-A (5),¹³ assiguxanthone-A (6),¹⁴ trapezifolixanthone (7),¹⁵ 5-*O*-methylxanthone V₁ (8),¹⁶ pancixanthone-B (9),¹³ 5-*O*-methyl-2-deprenylrheediaxanthone B (10),¹⁷ and (+)-4-oxa-tricyclo[4.3.1.0]decan-2-one scaffold (11),^{18,19} by 1D and 2D NMR techniques and compared with previous reports of these known compounds.

EXPERIMENTAL

GENERAL

Melting points were measured with using a Bibby Stuart Scientific melting point apparatus SMP3. UV spectra were measured on a UNICAM UV-310 spectrophotometer. Infrared spectra (IR) were recorded on a Perkin-Elmer 1750 FT-IR spectrophotometer. ESI-TOF mass spectra were recorded on a Micromass LCT spectrometer. NMR spectra (¹H, ¹³C, DEPT, ¹H-¹H COSY, NOESY, HMQC and HMBC) were recorded on Bruker AV300 or AV500 spectrometers. Chromatography was performed with the use of Merck prep-PLC and TLC, Merck silica gel 100 column and Sephadex LH-20 column. Optical rotation was measured in acetone solution with a sodium D line (589 nm) on an AUTOPOL[®] P-1020 (A068860638) automatic polarimeter.

PLANT MATERIAL

Barks of *C. sumatranum* ssp. *neriifolium* (Clusiaceae) were collected at Amphur Bannasan, Suratthani Province, Thailand, in February 2009. The plant was identified by Assistant Professor Dr. Maruay Mekanawakul, a specialist in the botanics, and the voucher specimen (WU 1453) is deposited at the botanic garden, Walailak University, Nakhon Si Thammarat, Thailand.

EXTRACTION AND ISOLATION

Chopped dried barks of *C. sumatranum* ssp. *neriifolium* (18.5 kg) were extracted with CH_2Cl_2 at room temperature for 3 days. After a removal of solvents, a yellow-brown viscous CH_2Cl_2 extract (366.28 g) was obtained. This extract (366.28 g) was dissolved in acetone. The acetone-soluble portion (255.28 g) was fractionated by quick column chromatography over silica gel 60H with the use of hexane, hexane- CH_2Cl_2 , CH_2Cl_2 -acetone, acetone and acetone-MeOH as eluents to afford 10 fractions (FCN1-FCN10).

Fraction FCN5 (8.01 g) was subjected to Sephadex LH-20 column chromatography with MeOH as an eluent, yielding 5 subfractions (FCN5a- FCN5e). **7** (6.4 mg) was obtained from repeated Sephadex LH-20 column chromatography with MeOH as an eluent and then by prep-TLC (CH₂Cl₂: hexane, 3:2 v/v) of subfraction FCN5e (218.7 mg). Repeated Sephadex LH20 gel filtration of fraction FCN6 (20.14 g) gave **4** (32.9 mg), **8** (1.4 mg) and **9** (110.6 mg). Fraction FCN7 (15.52 g) was separated by Sephadex LH-20 column chromatography with MeOH as an eluent, yielding **10** (40.0 mg). Fraction FCN8 (9.16 g) was purified by Sephadex LH-20 column chromatography with MeOH as an eluent, yielding **10** (40.0 mg). Fraction FCN8 (9.16 g) was purified by Sephadex LH-20 column chromatography with MeOH as an eluent, yielding **6** subfractions (FCN8e1- FCN8e6). Subfraction FCN8e3 (40.4 mg) was purified by prep-PLC (100% CH₂Cl₂) to afford **1** (19.3 mg) while **5** (32.7 mg) and **3** (6.6 mg) were derived from subfraction FCN8e4 (160.0 mg) by prep-PLC (100% CH₂Cl₂). Subfraction FCN8e5 (27.0 mg) and subfraction FCN8-f (50.0 mg) were purified by prep-PLC (100% CH₂Cl₂) to afford **11** (15.0 mg) and **6** (18.9 mg), respectively.

Neriifolone A 1: Yellow solid. mp 146–147 °C. UV λ_{max} (MeOH) (log ε): 246 (4.08), 256 (3.92), 320 (3.86), 354 (3.38) nm. IR (neat) v_{max} : 3392 (OH), 1647 (C=O) cm⁻¹. ¹H-NMR (300 MHz, acetone- d_6) and ¹³C-NMR (75 MHz, acetone- d_6) see Table 1. TOF-MS [M+H]⁺ m/z: 343.1194 for C₁₉H₁₉O₆ (calcd. 343.1176).

Neriifolone B **2**: Yellow solid. mp 231–232 °C. UV λ_{max} (MeOH) (log ε): 244 (4.42), 314 (4.00), 348 (3.77) nm. IR (neat) ν_{max} : 3368 (OH), 1780 (C=O), 1651 (C=O) cm⁻¹. ¹H-NMR (300 MHz, acetone- d_6) and ¹³C-NMR (75 MHz, acetone- d_6) see Table 1. TOF-MS [M+Na]⁺ m/z: 379.0789 for C₁₉H₁₆O₇Na (calcd. 379.0788).

Neriifolone C **3**: Yellow solid. mp 297–298 °C. Optical rotation: $[\alpha]_D^{23.6}$ -27.42° (*c* 0.2600 %w/v in acetone). UV λ_{max} (MeOH) (log ε): 248 (4.28), 256 (4.27), 318 (4.02), 366 (3.07) nm. IR (neat) v_{max} : 3392 (OH), 1651 (C=O) cm⁻¹. ¹H-NMR (300 MHz, acetone-*d*₆) and ¹³C-NMR (75 MHz, acetone-*d*₆) see Table 1. TOF-MS [M+H]⁺ *m/z*: 329.1018 for C₁₈H₁₇O₆ (calcd. 329.1020).

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REFERENCES

- 1. E. F. Aderson, *Econ. Bot.*, 1986, **40**, 442.
- 2. G. M. Kitanov, I. Assenov, and D. The Van, *Pharmazie*, 1988, 43, H12.
- 3. S. Laphookhieo, W. Maneerat, W. Narmdorkmai, and S. Koysomboon, *Heterocycles*, 2009, 78, 1299.
- 4. S. Laphookhieo, W. Maneerat, and S. Koysomboon, *Molecules*, 2009, 14, 1389.
- 5. S. Laphookhieo, W. Maneerat, T. Buatip, and J. K. Syers, Can. J. Chem., 2008, 86, 757.
- S. Laphookhieo, J. K. Syers, R. Kiattansakul, and K. Chantrapromma, *Chem. Pharm. Bull.*, 2006, 54, 745.
- N. Boonnak, C. Karalai, S. Chantrapromma, C. Ponglimanont, H. K. Fun, A. Kanjana-Opas, and S. Laphookhieo, *Tetrahedran*, 2006, 62, 8850.
- 8. S. Boonsri, C. Karalai, C. Ponglimanont, A. Kanjana-Opas, and K. Chantrapromma, *Phytochemistry*, 2006, **67**, 723.
- 9. W. Mahabusarakam, W. Nuangnaowarat, and W. C. Taylor, *Phytochemistry*, 2006, 67, 470.
- E. K. Seo, N. C. Kim, M. C. Wani, M. E. Wall, H. A. Navarro, J. P. Burgess, K. Kawanishi, L. B. S. Kardono, S. Riswan, W. C. Rose, C. R. Fairchild, N. R. Farnsworth, and A. D. Kinghorn, *J. Nat. Prod.*, 2002, 65, 299.
- W. Mahabusarakam, S. Rattanaburi, S. Phongpaichit, and A. Kanjana-Opas, *Phytochemistry Letters*, 2008, 1, 211.
- 12. K. Likhitwitayawuid, T. Phadungcharoen, and J. Krungkrai, *Planta Med.*, 1998, 64, 70.
- 13. C. Ito, Y. Miyamoto, K. Sunder Rao, and H. Furukawa, Chem. Pharm. Bull., 1996, 44, 441.
- C. Ito, Y. Miyamoto, M. Nakayama, Y. Kawai, K. Sunder Rao, and H. Furukawa, *Chem. Pharm. Bull.*, 1997, 45, 1403.
- E.-K. Seo, M. E. Wall, M. C. Wani, H. Navarro, R. Mukherjee, N. R. Farnsworth, and A. D. Kinghorn, *Phytochemistry*, 1999, **52**, 669.
- O. Thoison, D. D. Cuong, A. Gramain, A. Chiaroni, N. V. Hung, and T. Sévenet, *Tetrahedron*, 2005, 61, 8529.
- 17. G. Rath, O. Potterat, S. Mavi, and K. Hostettmann, *Phytochemistry*, 1996, 43, 513.
- O. Thoison, J. Fahy, V. Dumontet, A. Chiaroni, C. Riche, M. V. Tri, and T. Sévenet, *J. Nat. Prod.*, 2000, 63, 441.
- N.-G. Li, J.-X. Wang, X.-R. Liu, C.-J. Lin, Q.-D. You, and Q.-L. Guo, *Tetrahedron Lett.*, 2007, 48, 6586.