

82090-98-0; Ni(4-MeBPI)₂, 78065-33-5; Ni(5-MeBPI)₂, 87802-48-0; Ni(4-*t*-BuBPI)₂, 87802-49-1; Fe(BPI)₂, 79062-05-8; Fe(3-MeBPI)₂, 79062-04-7; Fe(4-MeBPI)₂, 78065-31-3; Fe(5-MeBPI)₂, 87802-50-4; Fe(4-*t*-BuBPI)₂, 87802-51-5; Mn(3-MeBPI)₂·¹/₂CH₂Cl₂, 87802-52-6; Zn(BPI)₂, 87802-53-7; 4-*tert*-butylphthalic acid, 14236-13-6; 4-*tert*-butylphthalonitrile, 32703-80-3; 2-aminopyridine-*d*₄, 87802-54-8; phthalonitrile-*d*₄, 69299-71-4.

Supplementary Material Available: Details of the crystal structure analysis, space group, and unit cell data, listings of positional and thermal parameters and selected individual bond distances, angles, torsional angles, and least-squares planes, additional perspective and stereoscopic views of the molecules, and a table of structure factor amplitudes (71 pages). Ordering information is given on any current masthead page.

Contribution from the Department of Chemistry, University of Arizona, Tucson, Arizona 85721, and Lehrstuhl für Anorganische Chemie I, Ruhr Universität Bochum, 4630 Bochum Querenburg, Federal Republic of Germany

⁹⁵Mo NMR Studies of Dioxo, Oxo-Sulfido, Oxo-Selenido, and Disulfido Complexes of Molybdenum(VI)

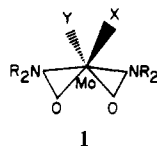
MARTIN MINELLI, JOHN H. ENEMARK,* KARL WIEGHARDT, and MANFRED HAHN

Received April 21, 1983

The ⁹⁵Mo NMR spectra of a number of Mo(VI) complexes with the general formula MoXY[ONHR]₂ and MoXY[ONR₂]₂ (X = O, S, Se; Y = O, S; R = CH₃, C₂H₅, C(CH₃)₃, CH₂C₆H₅; R₂ = C₅H₁₀) have been measured. Change of the terminal groups from oxygen to sulfur to selenium causes deshielding of the molybdenum nucleus in the order O < S < Se. A narrowing of the line width accompanies this deshielding. Bulky R groups also seem to have some deshielding effect. The quantitative conversion of the dioxo (MoO₂) complexes to the corresponding monooxo-monosulfido (MoOS) and disulfido (MoS₂) complexes by reaction with [(CH₃)₃Si]₂S was followed by ⁹⁵Mo NMR. Under similar conditions MoOSe[ONR₂]₂ was converted to MoOS[ONR₂]₂, MoS₂[ONR₂]₂, and several other species.

Introduction

Recently we have shown that low-symmetry, six-coordinate Mo(VI) complexes containing the [MoO₂]²⁺ unit can be conveniently studied by ⁹⁵Mo NMR.^{1,2} Another important group of [MoO₂]²⁺ complexes are the Mo(VI) complexes of structure 1 that result from the reaction of substituted hy-



droxylamines with molybdate.³ Our special interest in these complexes stems from the fact that their terminal oxo groups can be replaced by terminal sulfido and terminal selenido groups. Such complexes are relevant to understanding the molybdenum site of the oxidized form of xanthine oxidase. The molybdenum center of the active form of this enzyme is believed to contain a Mo(VI) atom with one terminal oxo and one terminal sulfido group (MoOS), whereas the inactive, cyanolyzed form of the enzyme is postulated to have two terminal oxo groups (MoO₂).⁴ A ⁹⁵Mo NMR study of complexes of type 1 in which the terminal oxo groups can be replaced by sulfido groups is therefore an important prerequisite to future ⁹⁵Mo NMR studies of molybdenum in enzymes and their cofactors.⁵ The ⁹⁵Mo NMR spectra for several type 1 complexes appeared while this work was in progress.⁶ Here we present results for additional examples and show that the formation and the interconversions of the terminal O, S, and Se groups in type 1 complexes can be followed by ⁹⁵Mo NMR.

Experimental Section

The complexes were synthesized as previously described.³ The ⁹⁵Mo NMR data were obtained on a Bruker WM250 NMR spectrometer. A 10-mm molybdenum probe (16.3 MHz) was used. A delay of 200 μs prior to acquisition was used to reduce the effects of probe ringing.

Table I. ⁹⁵Mo NMR Data

compd	chem shift, ^a ppm	line width, Hz
MoO ₂ [ONH(CH ₃) ₂] ₂	-219	200
MoO ₂ [ONH(C(CH ₃) ₃) ₂] ₂	-218	150
MoO ₂ [ON(CH ₃) ₂] ₂	-165	120
MoO ₂ [ON(C ₂ H ₅) ₂] ₂	-169	160
MoOS[ON(C ₂ H ₅) ₂] ₂	+544	140
MoOSe[ON(C ₂ H ₅) ₂] ₂	+865	120
MoS ₂ [ON(C ₂ H ₅) ₂] ₂	+1234	100
MoO ₂ [ONC ₅ H ₁₀] ₂	-184	160
MoOS[ONC ₅ H ₁₀] ₂	+531	100
MoOSe[ONC ₅ H ₁₀] ₂	+855	70
MoS ₂ [ONC ₅ H ₁₀] ₂	+1219	70
MoO ₂ [ON(CH ₂ C ₆ H ₅) ₂] ₂	-161	260

^a Measured at room temperature in DMF, concentration 0.4 M; if less soluble, saturated; chemical shifts relative to 2 M Na₂MoO₄ in H₂O at pH 11. The estimated uncertainty of the chemical shifts is ±1 ppm.

A 2 M Na₂MoO₄ solution at pH 11 served as external standard. All solvents were dried prior to use. In the case of the oxo-selenido complexes the NMR spectra were measured in sealed 10-mm NMR tubes under nitrogen; in the other cases regular 10-mm NMR tubes were used.

In Situ Thiolation Reactions. MoO₂[ON(C₂H₅)₂]₂ (1.5 m Mol) was dissolved in 2.5 mL of dry DMF under nitrogen in a 10-mm sealed screw top NMR tube (Wilmad) and the ⁹⁵Mo NMR spectrum measured. An equimolar amount of [(CH₃)₃Si]₂S (Fluka Chemical

- (1) Christensen, K. A.; Miller, P. E.; Minelli, M.; Rockway, T. W.; Enemark, J. H. *Inorg. Chim. Acta* **1981**, *56*, L27.
- (2) For a complete list of ⁹⁵Mo NMR studies see: Minelli, M.; Hubbard, J. L.; Christensen, K. A.; Enemark, J. H. *Inorg. Chem.* **1983**, *22*, 2652.
- (3) (a) Wiegardt, K.; Holzbach, W.; Weiss, F.; Nuber, B.; Prikner, B. *Angew. Chem., Int. Ed. Engl.* **1979**, *18*, 548. (b) Wiegardt, K.; Hahn, M.; Weiss, J.; Swiridoff, W. *Z. Anorg. Allg. Chem.*, in press.
- (4) Cramer, S. P.; Wahl, R.; Rajagopalan, K. V. *J. Am. Chem. Soc.* **1981**, *103*, 7721.
- (5) Johnson, J. L. In "Molybdenum and Molybdenum Containing Enzymes"; Coughlan, M. P., Ed.; Pergamon Press: Oxford, 1980; p 345.
- (6) Gheller, S. F.; Hambley, T. W.; Traill, P. R.; Brownlee, R. T. C.; O'Connor, M. J.; Snow, M. R.; Wedd, A. G. *Aust. J. Chem.* **1982**, *35*, 2183.

* To whom correspondence should be addressed at the University of Arizona.

Corp.) was added with a gastight syringe. (*Caution:* [(CH₃)₃Si]₂S has a terrible stench: All manipulations must be carried out in an efficient hood. The NMR tubes must be tightly sealed before transfer to the NMR spectrometer.) The reaction was followed for 4 h by ⁹⁵Mo NMR after which a second equivalent of [(CH₃)₃Si]₂S was added. The reaction was then checked periodically for 17 h and again after 1 week. A similar procedure was followed for MoOSe[ONC₅H₁₀]₂ except that the final solution resulted from a fourfold excess of the thiolating reagent. The identities of the products were inferred by comparison to the ⁹⁵Mo NMR spectra of known pure compounds.

Results and Discussion

These MoXY[ONHR]₂ and MoXY[ONR₂]₂ complexes (X = O, S, Se; Y = O, S; R = CH₃, C₂H₅, C(CH₃)₃, CH₂C₆H₅, R₂ = C₅H₁₀) are the only known six-coordinate Mo(VI) complexes where the terminal oxo groups can be replaced by terminal sulfido and selenido groups. Their geometry (**1**) is best described as a skew trapezoid bipyramid.³

The ⁹⁵Mo NMR signals of the dioxo species of these complexes occur around -200 ppm (Table I). As the R groups become bulkier, some deshielding of the ⁹⁵Mo NMR center is observed. The chemical shifts of these complexes are more negative than those of the pseudooctahedral Mo(VI) complexes with nitrogen and oxygen ligands.^{1,6} When one terminal oxo group is replaced by a sulfido group, a deshielding of the Mo center of about 700 ppm is observed. The magnitude of this deshielding is somewhat larger than that observed in tetrahedral MoX₄²⁻ complexes (X = O or S)⁷ when O is replaced by S. Replacement of the terminal oxygen by selenium causes deshielding of the molybdenum nucleus of over 1000 ppm, i.e. selenium causes more deshielding than sulfur as occurs for the tetrahedral MoX₄²⁻ complexes.⁸ The ⁹⁵Mo NMR signals for the disulfido complexes occur around 1200 ppm, about another 700 ppm downfield from the monooxo-monosulfido complexes. The line widths decrease with the deshielding of the molybdenum center. The deshielding is accompanied by a bathochromic shift of the lowest energy electronic transition in the UV-Vis spectra (OS 413 nm, OSe 457 nm; SS 514 nm, for R₂ = C₅H₁₀³). While this study was in progress we received a preprint from Dr. A. Wedd, who independently studied several complexes of the same type.⁶ The results for compounds common to both studies are in good agreement.

The large chemical shift differences among the type **1** complexes containing the MoO₂, MoOS, and MoS₂ groups prompted us to use ⁹⁵Mo NMR to investigate the reaction of MoO₂[ONR₂]₂ complexes with a thiolating reagent in aprotic

media. Hexamethyldisilathiane, [(CH₃)₃Si]₂S, was added to MoO₂[ON(C₂H₅)₂]₂ in a 1:1 ratio in DMF at room temperature. After about 2 h the corresponding MoOS species appeared in the ⁹⁵Mo NMR spectrum, and the intensity of the MoO₂ peak decreased. When the ratio of hexamethyldisilathiane was increased to 2:1, the MoS₂ species began to appear. After 1 week no dioxo complex was left and the ratio of MoOS:MoS₂ was about 3:1.

The clean conversion of MoO₂[ON(C₂H₅)₂]₂ to the corresponding MoOS and MoS₂ complexes by [(CH₃)₃Si]₂S led us to examine the reaction of MoOSe[ONC₅H₁₀]₂ with this reagent. Addition of an equimolar amount of hexamethyldisilathiane to the MoOSe complex resulted in formation of the MoOS complex after about 45 min. Increasing the ratio of disilthiane to complex to 4:1 produced the corresponding MoS₂ complex, two unidentified species at 1804 ppm and at 1840 ppm, and finally MoS₄²⁻. After 7 h no MoOSe starting complex remained.

The facile thiolation reactions of the type **1** complexes in aprotic media raised the question as to whether these complexes exchange terminal O and S groups in the absence of thiolating reagents. No comproportionation to the MoOS complex was observed after 2 days when the MoO₂ and MoS₂ complexes were both present in the same solution at room temperature.

This work shows that six-coordinate MoO₂L₄, MoOSL₄, MoOSeL₄, and MoS₂L₄ complexes can be easily distinguished by ⁹⁵Mo NMR. In addition, ⁹⁵Mo NMR can be used to directly follow the formation and the interconversion of these complexes. The results also provide encouragement that ⁹⁵Mo NMR should be able to distinguish between the postulated six-coordinate Mo(VI) sites of the active (MoOS) and the inactive (MoO₂) forms of xanthine oxidase (or its cofactor) provided that the line widths are sufficiently narrow.

Acknowledgment. We thank the U.S. Department of Agriculture for supporting this work with Grant No. 59-2401-1-626 and the National Science Foundation for funds for the NMR spectrometer. We thank Dr. K. A. Christensen for his technical assistance with the NMR and Dr. J. T. Spence for helpful discussions. We thank Dr. A. G. Wedd for a preprint of ref 6. J.H.E. thanks the Alexander von Humboldt Foundation for a Senior Scientist Award, which gave rise to this collaborative research.

Registry No. MoO₂[ONH(CH₃)₂], 70631-31-1; MoO₂[ONH(C(CH₃)₃)₂], 75701-12-1; MoO₂[ON(CH₃)₂]₂, 74081-85-9; MoO₂[ON(C₂H₅)₂]₂, 74081-86-0; MoOS[ON(C₂H₅)₂]₂, 76900-51-1; MoOSe[ON(C₂H₅)₂]₂, 84133-90-4; MoS₂[ON(C₂H₅)₂]₂, 76900-53-3; MoO₂[ONC₅H₁₀]₂, 84120-41-2; MoOS[ONC₅H₁₀]₂, 84133-89-1; MoOSe[ONC₅H₁₀]₂, 84133-41-5; MoS₂[ONC₅H₁₀]₂, 84146-55-4; MoO₂[ON(CH₂C₆H₅)₂]₂, 74081-87-1; ⁹⁵Mo, 14392-17-7.

(7) Lutz, O.; Nolle, A.; Kroneck, P. Z. *Naturforsch.*, A **1976**, *31A*, 454; **1977**, *32A*, 505.

(8) Gheller, S. F.; Gazzana, P. A.; Masters, A. F.; Brownlee, R. T. C.; O'Connor, M. J.; Wedd, A. G.; Rodgers, J. R.; Snow, M. R. *Inorg. Chim. Acta* **1981**, *54*, L131.