was raised above **7.** In those cases, the large changes were attributed to the onset of *terdentate* chelation by at least one or two of the bound malate ligands (via two carboxylate groups *plus* the α -OH, *or* hydroxylate, group). The change from bidentate to terdentate chelation by the malate ligands would be expected to significantly alter the ligand field sensed by the Ln³⁺ 4f electrons and, thereby, induce major changes in the be expected to significantly after the ligand rield sensed by the $Ln³⁺$ 4f electrons and, thereby, induce major changes in the 4f \rightarrow 4f optical spectra (especially in transition regions known to exhibit hypersensit to exhibit hypersensitive behavior).

Conclusions

The experimental results reported here clearly demonstrate the hypersensitive behavior of the ${}^4I_{9/2} \rightarrow {}^4G_{5/2}$, ${}^2G_{7/2}$ transitions of Nd(III), and it is shown how the absorption spectra associated with these transitions can be used to elicit information about complex formation (as a function of solution associated with these transitions can be used to elicit information about complex formation (as a function of solution pH). Furthermore, it was shown how the *relative* ${}^4I_{9/2} \rightarrow {}^4G_{5/2}$, ${}^2G_{7/2}$ absorption inten systems could be rationalized (at least qualitatively) in terms of specific ligand structural features such as (1) the nature and number of donor groups, (2) the nature and number of

nonligating substituent groups, and **(3)** chelate-ring conformational properties. Our direct calculations of transition oscillator strengths (for model structures presumed to be similar to the real structures present in solution at $pH > 7$) met with mixed success. Agreement between the calculated vs. experimentally determined oscillator strengths was good (in some cases, excellent) for the tris terdentate chelate structures **2-5.** However, our calculations grossly underestimated the oscillator strengths for the $NdCl₃(aq)$ system (modeled as a 9-coordinate $Nd(H_2O)_9^{3+}$ complex with exact D_{3h} symmetry). This led to significant overestimates of f - $(complex):f(NdCl₃(aq))$ ratios and hypersensitive behavior. It is likely that this problem can be attributed primarily to the inappropriateness of using our idealized structure **1** to model the complexes present in the $NdCl₃(aq)$ system.

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Registry No. $Nd(ODA)_3^2$, 43030-80-4; $Nd(DPA)_3^3$, 38721-35-6; Nd(IDA)₃³⁻, 12561-55-6; Nd(MIDA)₃³⁻, 89746-87-2; Nd(MAL)₃³⁻, **89773-17-1;** NdEHPG-, **89746-88-3.**

Contribution from the Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania **16802**

Europium(111) Luminescence Excitation Spectroscopy. A Species-Specific Method for the Quantitation of Lanthanide Ion Binding to Chelating Agents. Complexes of (**1,2-Ethanediyldioxy)diacetate**

MICHAEL ALBIN, GREGORY K. FARBER, and WILLIAM DEW. HORROCKS, **JR.***

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Laser excitation spectroscopy was used to monitor the complexation of (1,2-ethanediyIdioxy)diacetate (EDA²⁻) with lanthanide(III) ions. Relative formation constants for the 1:1 species were determined in competition experiments that monitor the decrease in the excitation maximum due to $Eu(EDA)^+$ as $Ln(EDA)^+$ is formed. The formation constant, K_2 , for the bis complex, Eu(EDA)₂⁻, was evaluated by following the changes in the ${}^7F_0 \rightarrow {}^5D_0$ excitation spectrum as a function of the concentration of added EDA²⁻. Excited-state lifetime measurements in H₂O and D₂O solution provided the number of coordinated water molecules in the two complexes **(7** and **3** for the mono and **bis** complexes, respectively). Structural comparisons between solid-state and solution species were made from the excitation spectra.

Introduction

Classical methods for the determination of the binding constants of metal ions to chelating ligands generally involve fitting the results of potentiometric titrations of the ligand in the presence and absence of metal ion to a sometimes complex scheme requiring the knowledge of multiple proton dissociation constants.' These methods are necessarily indirect, and their validity depends upon the correctness of assumptions made concerning the species present in solution. Work in this laboratory has involved the exploitation of laser-excited europium(II1) luminescence to monitor the binding of this ion to specific sites in proteins or individual complexes in solution. um(III) luminescence to monitor the binding of this ion to specific sites in proteins or individual complexes in solution.
Our experiment involves excitation of the ${}^{7}F_0 \rightarrow {}^{5}D_0$ transition of F_0 (TI) by accoratio of Eu(II1) by scanning a pulsed dye laser through the transition region (577-581 nm) while emission corresponding to the relatively intense ${}^5D_0 \rightarrow {}^7F_2$ transition (~614 nm) is monitored. Since the absorptive transition is between nondegenerate levels, the observation of more than one peak in the excitation spectrum implies the presence of more than one Eu(II1) environment. These methods have been employed in this laboratory to quantitate Eu(III) ion binding to proteins, 2^{-8} iono-

The present research illustrates the application of these species-specific methods to obtain two additional classes of

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phoric ligands,⁹ and monodentate anions¹⁰ as well as to monitor ligand-exchange kinetic processes in chelate systems.¹¹ In addition, from measurements of excited-state lifetimes made separately in H_2O and D_2O solution, it is possible to determine the number of water molecules coordinated to the Eu(II1) ion in any given species. $2,12$

Figure 1. Plot of intensity ($\lambda_{ex} = 579.50$ nm, $\gamma_{em} = 614$ nm) vs. $[EDA²⁻] ([Eu]_i = 1.27 mM):$ circles, experimental points; solid line, best fit for $log K_2 = 2.50$. The excitation spectrum at $[EDA²] =$ **3.44** mM (solid circle) is shown in the insert.

information. The first is the measurement of relative (to Eu^{3+}) constants for the binding of various ions to a chelating ligand under conditions where complex formation is essentially quantitative (e.g., an equimolar mixture of metal ion and quantitative (e.g., an equimolar mixture of metal fon and
ligand contains no unbound species). The ligand $(1,2-$
ethanediyldioxy)diacetate, EDA (1) , has been chosen as the **ethanediyldioxy)diacetate,** EDA **(l),** has been chosen as the

object of this study. Relative binding constants, K_{rel} , corresponding to the equilibrium of eq 1 (Ln^{3+}) represents any

$$
Eu(EDA)^{+} + Ln^{3+} \xleftarrow{K_{rel}} Ln(EDA)^{+} + Eu^{3+} \qquad (1)
$$

aquated ion of the lanthanide series other than Eu^{3+}), are obtained by monitoring the concentration of Eu(EDA)+ with laser excitation spectroscopy.

The second class of experiment exemplified here involves the measurement of the formation constant, K_2 , of the bis-(ligand) complex, according to eq 2. This is accomplished

$$
Eu(EDA)^{+} + EDA^{2-} \xrightarrow{K_2} Eu(EDA)_2^{-} \qquad (2)
$$

by monitoring the concentrations of Eu(EDA)+ and Eu- $(EDA)₂$ as a function of solution composition by means of their two distinct excitation spectral peaks (see insert, Figure 1).

Experimental Section

(1,2-Ethanediyldioxy)diacetic acid (H2EDA) was prepared by oxidizing triethylene glycol with nitric acid.¹³ Europium chloride solutions were prepared in doubly distilled deionized water with EuCl₃.6H₂O (Aldrich Chemical Co.) and standardized by titration with EDTA, using arsenazo as indicator.¹⁴ H₂EDA stock solutions were standardized by titration with a sodium hydroxide solution **standardized** with potassium hydrogen phthalate. All laser experiments were performed at 25 ± 0.1 °C on solutions with the ionic strength adjusted to 0.10 with sodium perchlorate, pH 5.5-6.0, and a total Eu(II1) concentration of 1.27 mM. Excitation spectra and lifetime

Figure 2. Excitation spectra of Eu(EDA)⁺ ([EDA²⁻]/[Eu] = 0.8;
pH 6): (a) ⁷F₀ \rightarrow ⁵D₀; (b) ⁷F₀ \rightarrow ⁵D₁; (c) ⁷F₀ \rightarrow ⁵D₂.

measurements were obtained by using a previously described nitrogen laser-pumped pulsed dye laser system.^{2,15} All intensities were recorded in a ratio mode with a photodynode used to monitor laser power fluctuations.

Determination of Relative Binding Constant, K_{rel} **.** These correspond to the equilibrium of *eq* 1 and are defined by eq **3.** Experimentally

$$
K_{\text{rel}} = \frac{[\text{Ln}(\text{EDA})^{+}][\text{Eu}^{3+}]}{[\text{Eu}(\text{EDA})^{+}][\text{Ln}^{3+}]} \tag{3}
$$

 $[Eu(EDA)^+]$ is measured by recording the intensity, *I*, of the excitation peak at 579.00 nm (Figure 2), where

$$
I = k[\text{Eu}(\text{EDA})^+] \tag{4}
$$

and *k* is a proportionality constant determined from a plot of *I* vs. $[Eu(EDA)^+]$ over the 1-10 mM concentration range. Equation 4, in conjunction with relationships **5-7,** reduces the equilibrium ex-

$$
[Eu3+]_i = [Eu3+] + [Eu(EDA)+] \qquad (5)
$$

$$
[Ln3+]_{i} = [Ln3+] + [Ln(EDA)+] \qquad (6)
$$

$$
[EDA2-]i = [Ln(EDA)+] + [Eu(EDA)+] \qquad (7)
$$

pression *(eq* **3)** to a function of *I, k,* and initial concentrations (eq **8),** where the subscript i denotes the initial concentration of the species

$$
K_{\text{rel}} = \frac{([\text{EDA}^2^-]_i - (I/k))([\text{Eu}^{3+}]_i - (I/k))}{(I/k)([\text{Ln}^{3+}]_i - [\text{EDA}^{2-}]_i + (I/k))}
$$
(8)

indicated. Under the conditions employed (pH 5.5–6.0, $[Ln^{3+}]_i$ = $[Eu³⁺]$ _i = $[EDA²⁻]$ _i = 1.27 mM), the contribution from HEDA⁻, H_2EDA and $Ln(EDA)_2$ ⁻ are negligible and are not considered in eq 5-7. Equation **8** is further reduced to *eq* 9 in the case where the initial concentrations of $EDA²$, $Eu³⁺$, and competing ion, $Ln³⁺$, are equal.

$$
K_{\rm rel} = \frac{([Eu^{3+}]_i - (I/k))^2}{(I/k)^2}
$$
 (9)

Determination of the Formation Constant of Eu(EDA)₂, *K***₂. This value was determined by monitoring changes in the ⁷F₀** \rightarrow **⁵D₀ excitation spectra as a function of [EDA²⁻]_i, while [Eu³⁺]_i was held** constant. Only two excitation peaks are detected (Figure 1, insert), one due to $Eu(EDA)^+$ and the other due to $Eu(EDA)_2^-$ (no free Eu^{3+} , $Eu(EDA)₃³⁻$, or other Eu(III)-containing species were observed under our conditions). K_2 was obtained from intensity measurements at 579.50 nm, the excitation maximum for $Eu(EDA)_2$ ⁻. At this wavelength, Eu(EDA)+ makes a nonnegligible contribution to the intensity, *I*, so one must express *I* as a sum of terms: $I = k'[Eu (EDA)^+$] + k'' $[Eu(EDA)_2^-]$. In a manner analogous to the procedure previously employed in this laboratory,¹⁰ this intensity relationship can be expressed as a function of k' , k'' , K_2 , and the initial concen-

trations of EDA²⁻ and Eu³⁺

$$
I = k'[Eu3+]_{i} - \frac{k'P}{2} + \frac{k''P}{2}
$$
(10)

where

- $P = [\text{Eu}^{3+}]_i + [\text{EDA}^{2-}]_i + K_2^{-1} ([\text{Eu}^{3+}]_i^2 2[\text{Eu}^{3+}]_i[\text{EDA}^{2-}]_i]$ + $2[Eu^{3+}]_iK_2^{-1}$ + $[EDA^{2-}]_i^{2}$ + $2[EDA^{2-}]_iK_2^{-1}$ + $K_2^{-2})^{1/2}$ (11)
- By the use of a nonlinear regression analysis with k' , k'' , and K_2 as

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Figure 3. Excitation spectra of $Eu(EDA)_2^-$ ($[EDA^{2-}]/[Eu] = 30$; **pH** *6).* Transitions are as in Figure **2.**

Table **I.** Frequencies of Electronic Transitions

	freq, cm^{-1}		
transition	$Eu(EDA)^+$	$Eu(EDA)$, $\overline{}$ 17257	
	17272		
${}^7F_0 \rightarrow {}^5D_0$ ${}^7F_0 \rightarrow {}^5D_1$	19032	19016	
	19019	19005	
${}^{7}F_{0} \rightarrow {}^{5}D_{2}$	21528	21.507	
	21512	21494	
	21500 (sh)	21469	

adjustable parameters, the "best fit" value of K_2 was obtained for eight data points (see Figure 1).

Results and Discussion

Characterization of $Eu(EDA)^+$ **and** $Eu(EDA)_2^-$ **.** Figures **2** and 3 show ⁷F₀ \rightarrow ⁵D₀, ⁷F₀ \rightarrow ⁵D₁, and ⁷F₀ \rightarrow ⁵D₂ excitation spectra of Eu(EDA)⁺ and Eu(EDA)₂⁻, respectively. The composition of the solution was so adjusted (see captions to Figures **2** and **3)** that the complexes indicated were the only detectable Eu(II1)-containing species present. In each case the ${}^{7}F_0 \rightarrow {}^{5}D_0$ region contains only a single peak indicative of a single Eu(II1) species in solution. This transition occurs at 15 cm⁻¹ lower energy in Eu(EDA)₂ compared with Eu- $(EDA)^+$ (Table I). This is in accord with a previously noted trend^{15,16} wherein this transition occurs at lower energy the greater the negative charge on a complex in a homologous series. The peak width for the ${}^{7}F_0 \rightarrow {}^{5}D_0$ transition of Eu- $(EDA)^+$ is 13.1 cm⁻¹ fwhm while that of $Eu(EDA)_2$ ⁻ is 6.3 cm⁻¹ fwhm. For both complexes the ${}^7F_0 \rightarrow {}^5D_1$ and ${}^7F_0 \rightarrow$ 5D, transitions (Figures **2** and **3)** show structure owing to ligand field splittings of the excited states. Fewer than the maximum possible number of transitions (three in the case of 5D_1 and five in the case of 5D_2) are observed in all cases. The transitions for $Eu(EDA)₂$ again tend to occur at slightly lower energies than those of $Eu(EDA)^+$.

The number of water molecules coordinated to Eu(II1) in the two complexes was detemined by measuring the reciprocal excited-state lifetimes, τ^{-1} , separately in H₂O and D₂O solution.¹² The results were as follows: $Eu(EDA)^{+}$, $\tau_{H_2O}^{-1} = 7.72$ ms^{-1} , $\tau_{\text{D}_2\text{O}}^{-1}$ = 0.51 ms⁻¹; Eu(EDA)₂⁻, $\tau_{\text{H}_2\text{O}}^{-1}$ = 4.37 ms⁻¹, $\tau_{D_2O}^{-1} = 1.30 \text{ ms}^{-1}$. From the previously determined linear correlation between $\tau_{H_2O}^{-1} - \tau_{D_2O}^{-1}$ and the number of coordinated water molecules it is indicated that 7 ± 0.5 water molecules are corrdinated in $Eu(EDA)^+$ and 3 ± 0.5 in Eu- $(EDA)₂$. Four water molecules are apparently displaced when a second EDA²⁻ ligand binds to $Eu(\text{EDA})^+$, consistent with its potential tetradentate nature (vide infra). Reciprocal lifetime measurements on a crystalline solid of composition $Na[Eu(EDA)₂(H₂O)_x]$ isolated separately from $H₂O$ and $D₂O$ solution ($\tau_{H_2O}^{-1}$ = 3.55 ms⁻¹ and $\tau_{D_2O}^{-1}$ = 1.20 ms⁻¹) suggest the presence of two coordinated water molecules in the solid state, consistent with the X-ray structure of sodium diaquabis((**1,2-phenylenedioxy)diacetato)lanthanate(III)** tetra-

Figure 4. Relative formation constants for Ln(EDA)⁺: (O) laser results; (x) literature titration data.¹³

Table **II.** $\log K_1$ Values Obtained from Competition Studies^a

Ln	$log K$,	σ^b	Ln	log K	σ^{b}
Ce	4.38	0.16	Dy	4.54	0.02
P _T	4.89	0.15	Ho	4.51	0.02
Nd	5.10	0.13	Er	4.53	0.09
Sm	5.07		Tm	4.69	0.06
Eu	4.94	0.15	Yb	4.83	0.05
Gd	4.84	0.23	Lu	4.84	0.02
Tb	4.77	0.01			

standard deviation for six K_1 determinations. ^{*a*} Numbers are relative to literature value for Sm^{13} ^b σ is the

hydrate,¹⁷ where the total coordination number of the lanthanide ion is 10. In solution the first coordination sphere apparently expands to a coordination number of **11,** which is not unkown in lanthanide ion structural chemistry.¹⁸

Formation Constants for Ln(EDA)+ across the Lanthanide Series. Relative formation constants for $Ln(EDA)^+$ complexes with respect to $Eu(EDA)^+$ were obtained from experiments wherein equimolar concentrations of Eu^{3+} and another Ln^{3+} ion compete for $EDA²⁻$ ligand while the concentration of Eu(EDA)+ is monitored via laser methods (see Experimental Section). The logarithms of the formation constants, K_1 , for the entire series, based on the measured relative constants, are given in Table I1 and plotted in Figure **4.** The relative formation constants were placed on an absolute scale by using the literature value¹³ for $Sm(EDA)^+$. It was not possible for us to obtain an absolute value of K_1 for $Eu(EDA)^+$ by directly using laser methods owing to the detection limits of our current apparatus and interference from the signal due to Eu_{aq}^{3+} under the necessary conditions of the experiment. It can be seen in Figure **4** that good agreement between the direct spectroscopic results of this study (0) and the potentiometric titration data in the literature **(X)** is obtained. The errors in the observed $log K₁$'s quoted in Table II are based on a minimum of six separate runs for each $Ln³⁺$ ion.

A variety of trends in complex formation constants across the lanthanide series have been observed for chelating ligands.^{19,20} Sinha¹⁹ has compiled data for many systems and discussed the factors that are involved in the variation of *K* across the lanthanide series. No simple relationship is observed between ionic radii, which decrease monotonically, and formation constants. Contributing factors include ionic size in relation to ligand cavity size or bite constraints and changes in total coordination number at a certain point in the series, as well as entropy and enthalpy effects. Depicted graphically in Figure 5 (literature values from ref **13, 23-26)** are the formation constant (K_1) trends for the series of chelaing lig-

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Figure 5. Trends in lanthanide ion formation constants with dicarboxylate ligands.

ands, A-H, all of which have carboxylate groups separated by one or two potentially liganding atoms (average log *K,* values are indicated in parentheses under the structures). The

trend observed for H_2EDA (ligand D) is clearly not exactly matched by any of the others, although those of ligands C and E are fairly similar. Several aspects of the interaction of Ln(II1) ions and these dicarboxylate ligands are apparent from the trend in average formation constant. Assuming a strong interaction with the two carboxylate moieties, the trend in preference for donor groups is

$$
-i - \xi - \xi - \xi -
$$

Perusual of a compendium²¹ of chelate stability constants for $Ln(III)$ ions reveals that log $K₁$ values are typically in the ranges 2-4 for bidentate ligands, 5-7 for tridentates, and 8-10 for tetradentates. According to this correlation, ligands A and

B appear to be bidentate with no sulfur coordination. Ligands E and F fall in the tridentate range, while G appears to be tetradentate. Ligands C and D have formation constant values intermediate between those usually observed for bi- and tridentate ligands. With seven coordinated water molecules indicated from our lifetime data, bidentate coordination by D is not unreasonable (a total coordination number of 9); however, an analysis of the paramagnetically shifted 'H and ¹³C nuclear resonances of C shows²² that this ligand coordinates in a tetradentate fashion. Additionally, our finding that upon coordination of a second $EDA²⁻$ ligand to $Eu³⁺$ four water molecules are displaced suggests tetradentate binding by ligand D. Clearly, a variety of factors contribute to the magnitude of a stability constant, and simplified interpretations of any particular result or trend must be viewed with caution.

Direct Spectroscopic Determination of the Formation Conparticular result or trend must be viewed with caution.
 Direct Spectroscopic Determination of the Formation Constant of Eu(EDA)₂, K_2 **. ⁷F₀** \rightarrow **⁵D₀ excitation spectra of solutions containing EDA²⁻ to Eu^{3+**} hibit two distinct excitation peaks due to Eu(EDA)⁺ and $Eu(EDA)₂$ ⁻ (Figure 1, insert). By monitoring the intensity of the excitation spectra at 579.5 nm (where $Eu(EDA)₂$ is the principal absorber) as a function of the concentration of added $EDA²⁻$ and fitting the results to eq 10 and 11 (see Experimental Section), we obtained a value for $log K_2$ of 2.50 \pm 0.71. The theoretical curve obtained from this value of K_2 and $k' = 2.76 \times 10^4$ and $k'' = 1.50 \times 10^5$ is compared with the experimental data points in Figure 1. The $log K_2$ value found by us for the Eu^{3+}/EDA^{2-} system compares well with that obtained by means of potentiometric titration for the corresponding Sm^{3+} (2.91) and Gd³⁺ (2.98) systems as well as for the Eu3+/(**1,2-phenylenedioxy)diacetate** (C) complex equilbrium (2.85).

Conclusions

We have demonstrated the utility of $Eu³⁺$ excitation spec-**Conclusions**
We have demonstrated the utility of Eu³⁺ excitation spec-
troscopy of the ${}^7F_0 \rightarrow {}^5D_0$ transition in determining relative
formation constants by means of motel is a competition information constants by means of metal ion competition experiments in the case of very tight binding and an absolute formation constant in a more weakly binding situation. The advantage over other methods is that the present experiment measures a quantity that is species-specific and directly proportional to concentration. Extension of quantitative binding constant studies to more tightly binding systems should be possible by using a more intense laser source (e.g., a Nd-YAG laser-pumped dye laser), thus increasing the general applicability of the method.

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