

Stereodynamics of *trans*-[(*t*-C₄H₉)₂PR]₂M(CO)X Systems (R = H, CH₃; M = Rh(I), Ir(I); X = Cl, Br, I). Assignment of Conformational Preferences and Conformational Exchange Itineraries

C. Hackett Bushweller,* Christopher D. Rithner,* and David J. Butcher

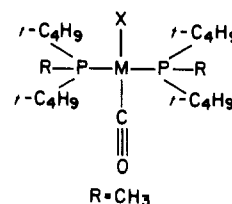
Received October 7, 1985

³¹P{¹H} and ¹³C{¹H} dynamic NMR (DNMR) spectra of *trans*-[(*t*-C₄H₉)₂PR]₂M(CO)X (R = H, CH₃; M = Rh(I), Ir(I); X = Cl, Br, I) show decoalescence phenomena that are assigned to slowing rotation about P-M and P-(*t*-C₄H₉) bonds. Rotation about the P-M bonds is slow on the NMR chemical exchange time scale (rate constants less than 1 s⁻¹) at about 210 K (Δ*G*[‡] = 12-16 kcal/mol). *tert*-Butyl rotation is slow at about 140 K (Δ*G*[‡] ≈ 7.5 kcal/mol). ³¹P{¹H} spectra at about 210 K (slow P-M rotation) show as many as three subspectra (three conformations). There is a dominant subspectrum at higher frequency (lower field), which is due to a conformation that has equivalent phosphorus atoms, and a second subspectrum with a lower population, which is due to a conformation with diastereotopic phosphorus atoms. Except for those complexes having R = H and X = I, there is a third, minor subspectrum at lower frequency (higher field), which is due to a conformation having equivalent phosphorus atoms. In conjunction with other evidence, the observed variations in subspectral populations as a function of halogen van der Waals radius provide circumstantial evidence for assigning the major subspectrum (equivalent phosphorus atoms) to a conformation having both R groups syn to halogen. The minor subspectrum (equivalent phosphorus atoms) is assigned to a conformation having both R groups syn to carbon monoxide. The subspectrum that shows diastereotopic phosphorus atoms is assigned to an anti conformation having one R group syn to halogen and the other syn to carbon monoxide. DNMR line-shape simulations reveal a preferred intramolecular exchange itinerary that involves rotation of one phosphine at a time about a P-M bond. ³¹P and ¹³C NMR chemical shift correlations with molecular geometry are also deduced.

Introduction

In metal-phosphine complexes, it is now well-established that stereochemical nonrigidity is the rule and is rarely the exception.¹ Establishing trends in conformational preference in solution and determining barriers to conformational exchange could be important in understanding reaction regioselectivity and asymmetric induction.²⁻⁴ In spite of this, there have been relatively few thorough, systematic studies of the stereodynamics of metal-phosphine complexes^{5,6} or the free, uncomplexed phosphines.⁷⁻⁹

This paper concerns ³¹P{¹H} and ¹³C{¹H} dynamic NMR (DNMR) studies of the Ir(I) and Rh(I) complexes 1-12. These are well-characterized, stable species that are not prone to decomposition during long-term DNMR studies and structurally simple enough to make conformational assignments tractable. Coordination at the metal is square planar (dsp²) with the pos-



1: M = Rh, X = Cl	4: M = Rh, X = I
2: M = Ir, X = Cl	5: M = Ir, X = Br
3: M = Rh, X = Br	6: M = Ir, X = I
R = H	
7: M = Rh, X = Cl	10: M = Ir, X = Br
8: M = Ir, X = Cl	11: M = Rh, X = I
9: M = Rh, X = Br	12: M = Ir, X = I

sibility of distortion due to steric crowding.^{10,11} The two bulky phosphines in each complex are *trans*. ³¹P{¹H} DNMR spectra, ¹³C{¹H} DNMR spectra, interatomic distance calculations, and variation of the halide ligand allow a strong circumstantial case for conformational assignments that are different from those presumed previously.^{5,6} ¹³C{¹H} DNMR spectra also allow a measurement of *tert*-butyl rotation barriers.

Results and Discussion

trans-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)Cl (1). The ³¹P{¹H} NMR spectrum (101.2 MHz) of the rhodium complex 1 (0.04 M in toluene-*d*₈) at 360 K shows a broadened doublet at δ 42.2 (¹J_{RhP} = 122 Hz).¹² The doublet results from scalar coupling of ³¹P to ¹⁰³Rh (*I* = 1/2; 100% natural abundance). Below 360 K, the spectrum decoalesces asymmetrically and, at 205 K, is separated into a series of well-resolved signals (Figure 1). A decomposition of the theoretical simulation of the 205 K spectrum (Figure 2) shows the B₂ portion of a B₂X spin system (X = ¹⁰³Rh; 59.9% of the total area of the composite spectrum), the AC part of an

- (1) (a) Cotton, F. A. *Acc. Chem. Res.* **1968**, *1*, 257. (b) Muetterties, E. L. *Acc. Chem. Res.* **1970**, *3*, 266. (c) Beall, H.; Bushweller, C. H. *Chem. Rev.* **1973**, *73*, 465.
- (2) (a) Morrison, J. D.; Masler, W. F.; Newburg, M. K. *Adv. Catal.* **1976**, *25*, 81. (b) Bosnich, B.; Fryzuk, M. D. *Top. Stereochem.* **1981**, *12*, 119. (c) Valentine, D., Jr.; Scott, J. W. *Synthesis* **1978**, 329. (d) Aleya, E. C.; Meek, D. W., Eds. *Catalytic Aspects of Metal Phosphine Complexes*; Advances in Chemistry Series 196; American Chemical Society: Washington, DC, 1982. (e) Crabtree, R. *Acc. Chem. Res.* **1979**, *12*, 331. (f) Jardine, F. H. *Prog. Inorg. Chem.* **1981**, *28*, 63.
- (3) Tolman, C. A. *Chem. Rev.* **1977**, *77*, 313.
- (4) Halpern, J. *Inorg. Chim. Acta* **1981**, *50*, 11.
- (5) (a) Bushweller, C. H.; Hoogasian, S.; English, A. D.; Miller, J. S.; Lourandos, M. Z. *Inorg. Chem.* **1981**, *20*, 3448. (b) Bushweller, C. H.; Rithner, C. D.; Butcher, D. J. *Inorg. Chem.* **1984**, *23*, 1967.
- (6) (a) Mann, B. E.; Masters, C.; Shaw, B. L.; Stainbank, R. E. *J. Chem. Soc. D* **1971**, 1103. (b) Bright, A.; Mann, B. E.; Masters, C.; Shaw, B. L.; Slade, R. M.; Stainbank, R. E. *J. Chem. Soc. A* **1971**, 1826. (c) Mann, B. E.; Shaw, B. L.; Slade, R. M. *Ibid.* **1971**, 2976. (d) Empsall, H. D.; Hyde, E. M.; Mentzer, E.; Shaw, B. L. *J. Chem. Soc., Chem. Commun.* **1977**, 2285. (e) Smith, J. G.; Thompson, D. T. *J. Chem. Soc. A* **1967**, 1694. (f) Bennett, M. A.; Tomkins, I. B. *J. Organomet. Chem.* **1973**, *51*, 289. (g) Gill, D. F.; Mann, B. E.; Shaw, B. L. *J. Chem. Soc., Chem. Commun.* **1973**, 311. (h) Faller, J. W.; Johnson, B. V. *J. Organomet. Chem.* **1975**, *96*, 99. (i) Reed, C. A.; Roper, W. R. *J. Chem. Soc., Dalton Trans.* **1973**, 1365.
- (7) (a) Bushweller, C. H.; Brunelle, J. A. *J. Am. Chem. Soc.* **1973**, *95*, 5949. (b) Brunelle, J. A.; Bushweller, C. H.; English, A. D. *J. Phys. Chem.* **1976**, *80*, 2598.
- (8) (a) Bushweller, C. H.; Brunelle, J. A. *Tetrahedron Lett.* **1974**, 893. (b) Bushweller, C. H.; Lourandos, M. Z. *Inorg. Chem.* **1974**, *13*, 2514.
- (9) (a) Rithner, C. D.; Bushweller, C. H. *J. Am. Chem. Soc.* **1985**, *107*, 7823.

- (10) Empsall, H. D.; Mentzer, E.; Pawson, D.; Shaw, B. L. *J. Chem. Soc., Chem. Commun.* **1977**, 311.
- (11) Hoffmann, P. R.; Yoshida, T.; Okano, T.; Otsuka, S.; Ibers, J. A. *Inorg. Chem.* **1976**, *15*, 2462.
- (12) ³¹P NMR chemical shifts are calibrated relative to trimethyl phosphite in a pertinent solvent, which is sealed in a coaxial reference tube. The ³¹P chemical shift of 85% H₃PO₄ is 142.1 ppm lower frequency (higher field) from trimethyl phosphite in toluene-*d*₈ at 310 K and 142.8 ppm lower frequency from trimethyl phosphite in 33% CD₂Cl₂/33% CF₂ClH/33% CFC₂H at 310 K.

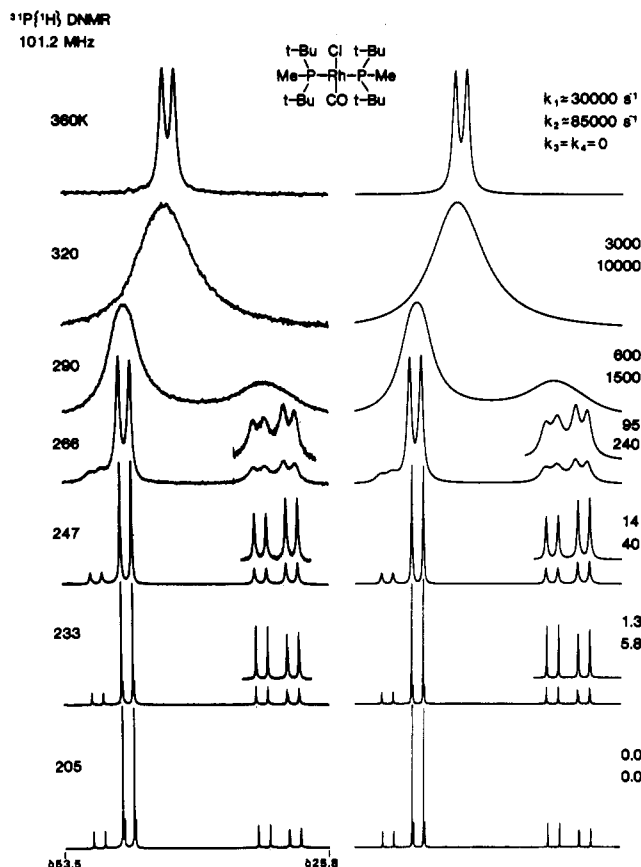


Figure 1. ³¹P{¹H} DNMR spectra of *trans*-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)Cl (1). Rate constants for theoretical simulations are defined in eq 1 and 2.

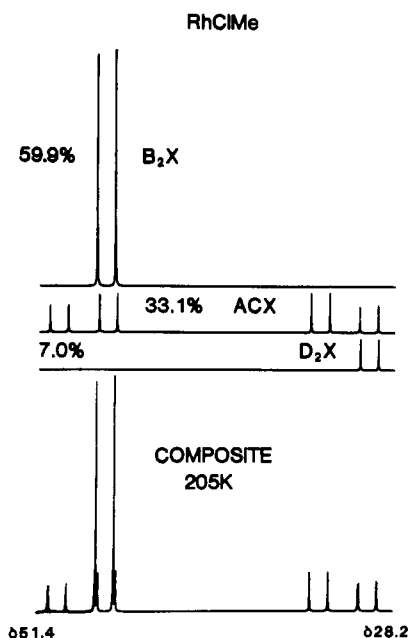


Figure 2. Decomposition of the ³¹P{¹H} NMR spectrum of 1 at 205 K.

ACX spin system (33.1%), and the D₂ portion of a D₂X spin system (7.0%).¹³ The minor D₂X doublet and the outer doublet of the C resonance almost superimpose (Figure 2). NMR parameters and subspectral populations are compiled in Table I.

(13) The computer program DNMR4 was used to simulate these static NMR line shapes and all the exchange-broadened DNMR spectra in this report: Bushweller, C. H.; Letendre, L. J.; Brunelle, J. A.; Bilofsky, H. S.; Whalon, M. R.; Fleischman, S. H. Quantum Chemistry Program Exchange, Indiana University, 1983; Program 466.

Table I. ³¹P NMR Parameters and Subspectral Populations for Complexes 1–12 under Conditions of Slow Rotation about Metal–Phosphorus Bonds

compd	spin syst obsd at slow exchange ^a	³¹ P NMR chem shifts, ppm	¹ J _{RhP} , Hz	² J _{PP} , Hz	rel population (temp, K)
1 ^b	B ₂ X	47.1	119.3		0.599 (205)
	D ₂ X	29.8	121.0		0.070
	ACX	48.4	118.2	323.0	0.331
2 ^b	B ₂	33.7			0.607 (214)
	D ₂	18.5			0.063
	AC	36.1		319.2	0.330
3 ^b	B ₂ X	39.7	117.2		0.666 (214)
	D ₂ X	23.7	119.7		0.034
	ACX	40.8	117.2	319.2	0.300
4 ^b	B ₂ X	26.1	120.8		
	D ₂ X	39.7	116.2		0.838 (223)
	ACX	26.8	118.0	311.0	0.006
5 ^b	B ₂ X	40.7	115.4		0.156
	D ₂ X	29.7	119.2	311.0	
	AC	33.4			0.705 (214)
6 ^b	D ₂	19.4			0.032
	AC	35.4		311.9	0.263
	B ₂	20.7		311.9	
7 ^c	B ₂	28.1			0.845 (223)
	D ₂	16.7			0.006
	AC	29.4		304.2	0.149
8 ^d	B ₂ X	19.2			
	D ₂ X	80.5	114.6		0.782 (220)
	ACX	40.8	118.1	320.7	0.010
9 ^d	B ₂ X	85.1	116.2		0.208
	D ₂ X	45.0	116.6	320.7	
	AC	74.2			0.900 (223)
10 ^d	B ₂	34.2			0.002
	D ₂	80.3		315.7	0.098
	AC	37.2		315.7	
11 ^d	B ₂ X	76.9	112.3		0.851 (214)
	D ₂ X	37.5	116.0		0.003
	ACX	83.2	113.5	315.5	0.146
12 ^d	B ₂	42.7			
	D ₂	72.5			0.946 (214)
	AC	33.0		309.7	0.053
13 ^d	B ₂ X	80.6			
	D ₂ X	35.8	309.7		0.935 (214)
	ACX	72.3	109.9	310.1	0.065
14 ^d	B ₂ X	81.0	112.3		
	D ₂ X	43.0	112.7	310.1	
	AC	67.2			0.975 (214)
15 ^d	B ₂	77.8			0.025
	D ₂	33.5		301.5	
	AC	33.5		301.5	

^aX = ¹⁰³Rh (*I* = 1/2). ^bSolvent toluene-*d*₈. ^cSolvent 20% CDCl₃/80% toluene-*d*₈. ^dSolvent CDCl₃.

Below 205 K, the ³¹P{¹H} spectrum of 1 (0.04 M in 33% CD₂Cl₂/33% CFCl₂H/33% CF₂ClH) shows changes in subspectral populations but *no additional decoalescence phenomena* down to 125 K.

The DNMR spectra in Figure 1 might reflect *intermolecular* or *intramolecular* exchange processes. It is necessary to distinguish between the two possibilities. For all of the Rh(I) complexes in this study, ¹⁰³Rh–³¹P scalar coupling (¹J_{RhP} = 110–122 Hz; Table I) is observed at room temperature *and above*. The addition of free phosphine to the NMR sample causes no coalescence of ³¹P{¹H} NMR signals for free and bound phosphine and causes no change in the ³¹P{¹H} DNMR spectra of 1. From both of these observations, it must be concluded that Rh–P bond dissociation (or *intermolecular* phosphine exchange) does not occur even at a slow rate on the NMR chemical exchange time scale at temperatures as high as 360 K (Figure 1). The ¹³C{¹H} NMR spectrum of the carbon monoxide ligand of 1 at 310 K shows both ¹⁰³Rh–¹³C (¹J_{RhC} = 74 Hz) and ³¹P–¹³C (²J_{PC} = 16 Hz) scalar coupling which reveals no metal–carbon monoxide dissociation. Removal or exchange of chloride in complexes such as 1 requires the use of a Lewis acid or excess ionic halide *in polar solvents*.⁶¹

Table II. ^{13}C NMR Chemical Shifts for Complexes **1** and **2** under Conditions of Slow Rotation about Metal-Phosphorus Bonds

compd (temp, K)	$^*\text{C}(\text{CH}_3)_3$ ($^{\nu}J_{\text{PC}}$, Hz)	$\text{C}(\text{CH}_3)_3$	PCH_3 ($^{\nu}J_{\text{PC}}$, Hz)	CO
1 (205 K) ^a	34.7 (t, 20) ^b	29.4	4.12 (t, 22.8)	189.7 (m)
	35.5 (m) ^c	29.7	8.17 (t, 22.0)	
			2.84 (d, 21.7) ^d	
			7.66 (d, 22.8)	
2 (214 K) ^a	35.4 (t, 26)	29.3	2.40 (t, 29.6)	173.5 (m)
		29.8	8.00 (t, 26.0)	
			0.98 (d, 27.7)	
			7.38 (d, 29.6)	

^aSolvent CDCl_3 . ^bt = triplet. ^cm = multiplet. ^dd = doublet.

In the *nonpolar* solvents used in our DNMR studies and in the absence of Lewis acids or ionic halides, metal-chloride dissociation does not occur. Thus, for spectroscopic and chemical reasons, it must be concluded that the rate process observed in Figure 1 is intramolecular and involves conformational exchange among at least three species, which give the B_2X , D_2X , and ACX subspectra. It is important to note that the $^2J_{\text{PP}}$ value (323 Hz; Table I) for the diastereotopic phosphorus atoms that give the ACX subspectrum verifies that the two phosphines are *trans* and not *cis*.¹⁴ The $^2J_{\text{PP}}$ value for *cis* phosphines is usually much smaller in magnitude.¹⁴

The $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (62.9 MHz) of **1** (0.04 M in CDCl_3) at 303 K shows signals at δ 189.4 (CO; $^1J_{\text{RhC}} = 74$ Hz, $^2J_{\text{PC}} = 16$ Hz), δ 35.0 (*t*- C_4H_9 quaternary; $^{\nu}J_{\text{PC}} = 17.6$ Hz), and δ 29.7 (*t*- C_4H_9 methyls; $^{\nu}J_{\text{PC}} = 5.2$ Hz) and an *exchange-broadened* resonance at δ 5.1 (PCH_3). Part of the spectrum at 303 K is shown in Figure 3. $^{13}\text{C}\{^1\text{H}\}$ NMR parameters are compiled in Table II.

Below 303 K, the quaternary carbon, *tert*-butyl methyl and *P*-methyl resonances all decoalesce and give sharp signals at 214 K (Figure 3). In particular, the *P*-methyl resonance is separated at 214 K into four signals including a dominant virtual triplet, a minor virtual triplet, and two doublets of equal area, which are marked respectively with a solid circle, an \times , and open circles in Figure 3. From systematic calculations of a series of four-spin spectra,¹⁵ the virtual triplet pattern for the dominant signal at δ 4.12 ($^{\nu}J_{\text{PC}} = 22.8$ Hz) is consistent with a $^{13}\text{CPRhP}'$ system that has *equivalent* phosphorus atoms. It corresponds to the dominant B_2X doublet in the $^{31}\text{P}\{^1\text{H}\}$ spectrum at 205 K (Figure 2). The minor triplet at δ 8.17 ($^{\nu}J_{\text{PC}} = 22$ Hz) labeled with an \times in Figure 3 is also consistent with equivalent phosphorus atoms and corresponds to the minor D_2X subspectrum in the $^{31}\text{P}\{^1\text{H}\}$ spectrum. The doublets at δ 7.66 ($^{\nu}J_{\text{PC}} = 22.8$ Hz) and δ 2.84 ($^{\nu}J_{\text{PC}} = 21.7$ Hz) labeled with open circles in Figure 3 result from a $^{13}\text{CPRhP}'$ spin system that has *different* chemical shifts for the two phosphorus atoms. Thus, the two $^{13}\text{C}\{^1\text{H}\}$ doublets of equal area at δ 7.66 and 2.84 correspond directly to the $^{31}\text{P}\{^1\text{H}\}$ ACX subspectrum (Figure 2), which clearly shows diastereotopic phosphorus atoms.

The quaternary carbon resonance decoalesces below 303 K and is separated at 214 K into a dominant virtual triplet at δ 34.7 ($^{\nu}J_{\text{PC}} = 20$ Hz; see solid circle in Figure 3), indicating equivalent phosphorus atoms, and a multiplet centered at δ 35.5 (open circle). The chemical shift differences are quite small, and the spectrum has limited value. The *tert*-butyl methyl signal is decoalesced at 214 K into a dominant signal at δ 29.4 (solid circle) and a smaller signal at δ 29.7 (open circle), which are also very closely spaced and have limited informational content.

Below 214 K (Figure 3), the quaternary carbon and *P*-methyl carbon resonances (67.9 MHz) of **1** (0.04 M in 33% $\text{CD}_2\text{Cl}_2/33\%$ $\text{CFCl}_2\text{H}/33\%$ CF_2ClH) undergo no further decoalescence. The dominant *P*-methyl and quaternary carbon triplets do become

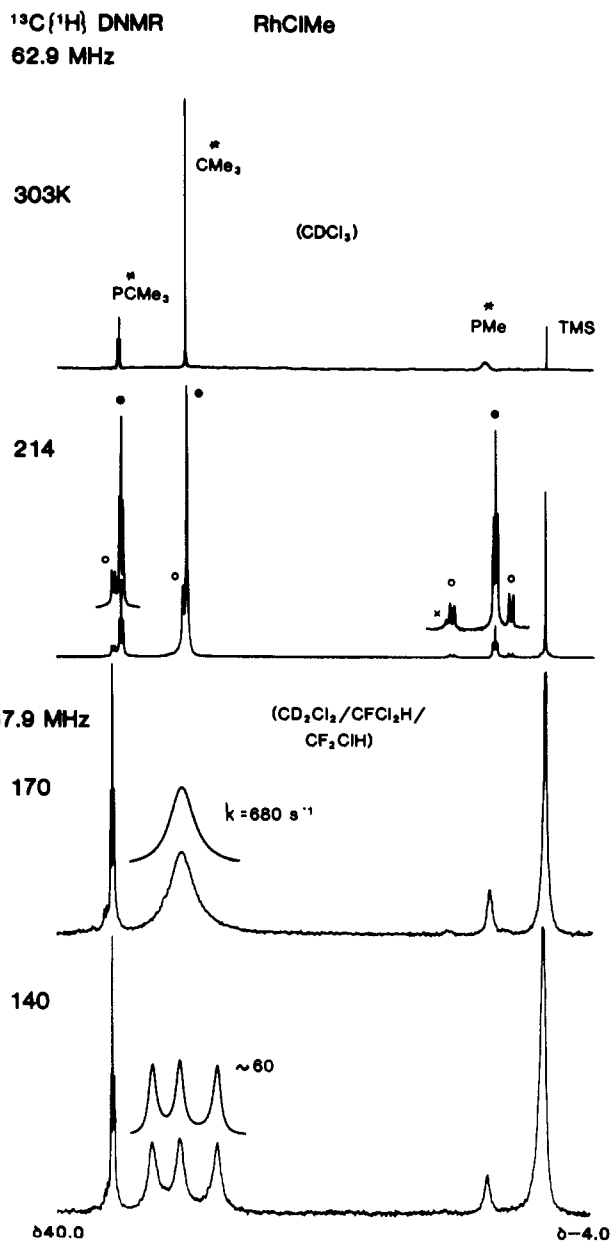
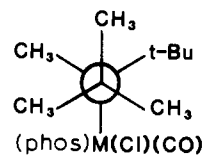


Figure 3. $^{13}\text{C}\{^1\text{H}\}$ DNMR spectra of *trans*- $[(t\text{-C}_4\text{H}_9)_2\text{PCH}_3]_2\text{Rh}(\text{CO})\text{Cl}$ (**1**). The spectra at 303 and 214 K were run in CDCl_3 and, at 170 and 140 K, in $\text{CD}_2\text{Cl}_2/\text{CFCl}_2\text{H}/\text{CF}_2\text{ClH}$.

more dominant at lower temperatures. In addition, the $^{31}\text{P}\{^1\text{H}\}$ spectrum undergoes no further decoalescence down to 125 K. However, the $^{13}\text{C}\{^1\text{H}\}$ spectrum of the *tert*-butyl methyl groups undergoes a clear-cut decoalescence and, at 140 K, is separated into three broad singlets of equal area at δ 26.6, 29.7 and 32.0 (Figure 3). In light of the local symmetry proximate to a *tert*-butyl group (**13a**) and the observation of three *tert*-butyl methyl signals



13a: M = Rh

13b: M = Ir

of equal area at 140 K, we are compelled to assign this decoalescence to threefold *tert*-butyl rotation. Threefold *tert*-butyl rotation does not effect any net change in molecular geometry and therefore will have no effect on the *tert*-butyl quaternary and *P*-methyl $^{13}\text{C}\{^1\text{H}\}$ resonances or on the $^{31}\text{P}\{^1\text{H}\}$ resonances of **1**.¹⁶

(14) For a review, see: Diehl, P.; Fluck, E.; Kosfeld, R., Eds. *NMR Basic Principles and Progress* Springer-Verlag: New York, 1979.

(15) The computer program UEA1TR was used for simulating static spectra with four or more spins: Johannesen, R. B.; Ferretti, J. A.; Harris, R. K. Quantum Chemistry Program Exchange, Indiana University, 1979; Program 188.

A three-site DNMR simulation at 170 K gives a *tert*-butyl rotation barrier (ΔG^\ddagger) of 7.6 ± 0.4 kcal/mol.¹³ It is interesting to note that the rather large Rh(phos)(CO)Cl moiety raises the *tert*-butyl rotation barrier only 1.4 kcal/mol above that in the free, uncomplexed di-*tert*-butylmethylphosphine ($\Delta G^\ddagger = 6.2 \pm 0.3$ kcal/mol at 120 K).⁹

Therefore, we assign the higher temperature decoalescence for **1** (Figure 1 and 3) to rotation about Rh–P bonds ($\Delta G^\ddagger \approx 13$ kcal/mol) and the lower temperature decoalescence (Figure 3) to *tert*-butyl rotation ($\Delta G^\ddagger = 7.6$ kcal/mol). The three different ³¹P{¹H} subspectra (B₂X, ACX, D₂X) observed at 205 K (Figure 2) correspond to three different equilibrium conformations associated with restricted rotation about the Rh–P bonds.

trans-[(t-C₄H₉)₂PCH₃]₂Ir(CO)Cl (2). The ³¹P{¹H} NMR spectrum (101.2 MHz) of the iridium complex **2** (0.04 M in toluene-*d*₆) at 360 K is a broad singlet at δ 29.4.¹² Although ¹⁹¹Ir (37.3% natural abundance) and ¹⁹³Ir (62.7% natural abundance) both have a spin of ³/₂, efficient nuclear quadrupolar relaxation decouples iridium from phosphorus in **2** and in all the other iridium complexes in this study. Below 360 K, the spectrum decoalesces and, at 214 K, is separated into a B₂ singlet (60.7%; equivalent phosphorus atoms), a smaller D₂ singlet (6.3%; equivalent phosphorus atoms), and an AC subspectrum (33.0%; ²J_{PP} = 319 Hz; diastereotopic and trans phosphorus atoms) as shown in Figures 4 and 5 (supplementary material). NMR parameters are compiled in Table I.

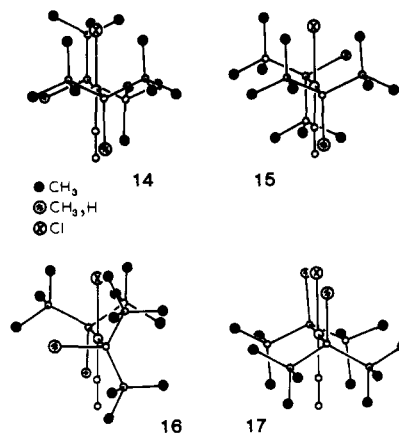
The ³¹P{¹H} spectrum of **2** (0.03 M in 33% CD₂Cl₂/33% CF₂ClH/33% CFC₂H) shows no additional decoalescence down to 130 K.

The ¹³C{¹H} DNMR spectra of **2** are shown in Figure 6 (supplementary material). The spectrum of **2** at 305 K (0.03 M in CDCl₃) shows resonances at δ 173.5 (CO; ²J_{PC} = 9 Hz), δ 35.6 (*t*-C₄H₉ quaternary; ^νJ_{PC} = 25.0 Hz), and δ 29.7 (*t*-C₄H₉ methyls) and an exchange-broadened peak at δ 3.1 (PCH₃). The observation of scalar coupling between the carbon monoxide carbon-13 and the phosphorus atoms reveals no carbon monoxide or phosphine dissociation at 305 K. In the relatively nonpolar CDCl₃, chloride dissociation also should not occur (vide supra). Between 305 and 214 K, the spectrum of **2** (Figure 6) decoalesces in a manner that is essentially identical with that for **1** (Figure 3). Although the signals for **2** are shifted slightly upfield as compared to those for **1**, the *P*-methyl group of **2** at 214 K gives a dominant virtual triplet (δ 2.4; ^νJ_{PC} = 29.6 Hz; equivalent phosphorus atoms), a minor virtual triplet (δ 8.0; ^νJ_{PC} = 26.0 Hz), and two doublets of equal area at δ 7.38 (^νJ_{PC} = 29.6 Hz) and δ 0.98 (^νJ_{PC} = 27.7 Hz), which reveal diastereotopic phosphorus atoms. The *tert*-butyl quaternary and methyl resonances also decoalesce into groups of very closely spaced signals (Table II). The *P*-methyl ¹³C{¹H} spectrum of **2** at 214 K correlates precisely with the ³¹P{¹H} spectrum at 214 K. The decoalescence phenomena that occur in the ³¹P{¹H} and ¹³C{¹H} spectra above 214 K are assigned to slowing rotation about the Ir–P bonds ($\Delta G^\ddagger \approx 14$ kcal/mol). Only the *tert*-butyl methyl ¹³C{¹H} resonance of **2** (0.03 M in 33% CD₂Cl₂/33% CFC₂H/33% CF₂ClH) decoalesces below 214 K and is separated at 130 K into three singlets or equal area at δ 27.0, 29.5, and 32.2 (Figure 6; Table II). This decoalescence is attributed to *tert*-butyl rotation (see **13b**; $\Delta G^\ddagger = 7.2 \pm 0.4$ kcal/mol at 160 K).

Assignment of Conformational Preferences. The three subspectra observed in the ³¹P{¹H} spectra of **1** and **2** at about 210 K must be due to different molecular conformations associated with restricted rotation about the metal–phosphorus bonds.

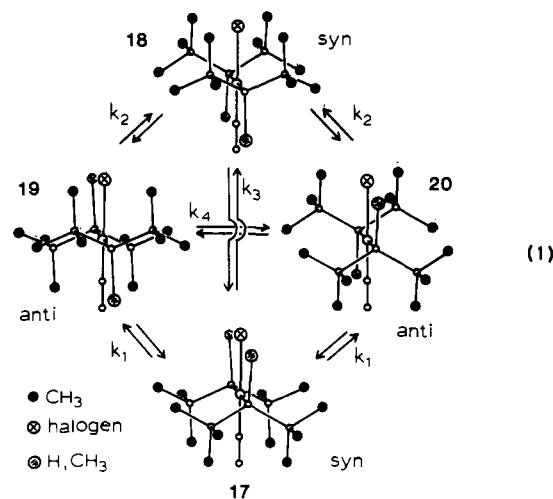
Assignments are problematic because NMR parameters that probe molecular geometry unequivocally (e.g., a Karplus relationship) are not available. Thus, we will present below a circumstantial case for conformational assignments that is based on observed variations of subspectral populations as a function of halogen steric bulk and other considerations.

One premise to be used in predicting equilibrium conformations in these systems has much precedent. The *tert*-butyl group is sterically large and will prefer to stagger and not eclipse the chlorine and carbon monoxide ligands. Staggering by *tert*-butyl groups is observed in the crystal structures of *trans*-[(*t*-C₄H₉)₂PC₆H₅]₂Rh(N₂)H¹¹ and the cyclic *trans*-[(*t*-C₄H₉)₂PC≡C(CH₂)₅C≡CP(*t*-C₄H₉)₂]Ir(CO)Cl.¹⁰ Conformations such as **14** (*t*-C₄H₉/Cl eclipsing) and **15** (*t*-C₄H₉/CO eclipsing) are pre-



dicted to be unstable. A conformation such as **16** in which the *P*-methyl bond is in a plane perpendicular to the metal coordination plane and the *tert*-butyl groups are skewed only slightly away from perfect eclipsing also involves severe *t*-C₄H₉/Cl and *t*-C₄H₉/CO nonbonded repulsions. Thus, **16** is also predicted to be unstable. An example of a stable conformation with optimally staggered *tert*-butyl groups is **17**, which has both *P*-methyl groups syn to chlorine. A requirement of optimized staggering for a (*t*-C₄H₉)₂P moiety about the Cl–M–CO axis then requires an eclipsing of the *P*-methyl groups by chlorine or carbon monoxide.

Therefore, we predict four equilibrium conformations for **1** and **2**, which are illustrated in eq 1: two diastereomeric syn forms (**17**, **18**) and two equivalent anti forms (**19**, **20**). A complete



(16) It is inevitable in these complexes that twisting away from perfect staggering about P–(*t*-C₄H₉) bonds will occur in the equilibrium conformations.⁹ This introduces rate processes involving low-torsion *tert*-butyl libration (or twisting) as well as threefold *tert*-butyl rotation. Libration does not involve vicinal eclipsings in the transition state and consistently has a much lower barrier than threefold *tert*-butyl rotation.⁹ It is apparent that, at 140 K, *tert*-butyl rotation is slow while libration is fast. In fact, slow *tert*-butyl rotation and slow libration would give, in principle, six *tert*-butyl methyl resonances for a given phosphine moiety.

accounting of all the stable conformations of **1** or **2** must include two equivalent anti forms, which are interconverted directly by two simultaneous 180° rotations of both phosphines about the P–M bonds. There is a statistical preference of **2** for the anti forms over either one of the syn forms. In addition, an accurate kinetic model for conformational exchange to be used in any DNMR simulations must include two anti forms (vide infra).

In the ³¹P{¹H} and ¹³C{¹H} spectra of **1** and **2** at about 210 K, which show slow rotation about the metal–phosphorus bonds,

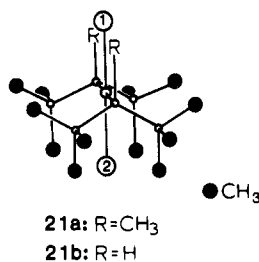
Table III. Cone Angles^a and van der Waals Radii

group	cone angle, deg	van der Waals radius, Å	group	cone angle, deg	van der Waals radius, Å
H	75	1.2	Cl	102	1.8
CO	95	1.4 ^b	Br	105	2.0
CH ₃	90	2.0	I	107	2.2

^aReference 3. ^bReference 17.

assignment of the anti resonances is trivial. In an anti form, the two phosphorus atoms are diastereotopic and will give anisochronous ³¹P chemical shifts. Thus, the ACX subspectrum for **1** and the AC subspectrum for **2** are assigned to anti conformers **19** and **20** (eq 1). Likewise, the two *P*-methyl groups in an anti form are diastereotopic. The two *P*-methyl doublets of equal area observed in each of the 214 K ¹³C{¹H} spectra for **1** and **2** are assigned to the anti forms.

Assignment of the syn forms is more equivocal. The relative stabilities of the two syn forms (**17**, **18**; eq 1) will be determined by the relative sizes of halogen and carbon monoxide. In addition, the syn conformational preference will be determined by the relative degrees of crowding at the position syn to the two methyl groups (site 1 in structure **21a**) and the position gauche to the



four *tert*-butyl groups (site 2 in **21a**). The larger ligand will prefer to occupy the less crowded of sites 1 and 2 in **21a**. With use of both van der Waals radii and cone angle values for carbon monoxide (1.4 Å; 95°) and chlorine (1.94 Å; 102°), chlorine is determined to be the larger ligand (Table III).^{3,17}

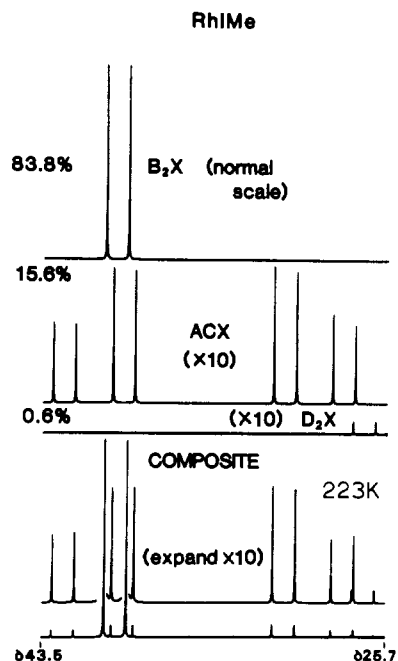
Interatomic distance calculations for **1** and **2** were done by using the computer program COORD.¹⁹ An idealized geometry with standard bond lengths, perfect C_{2v} symmetry, tetrahedral bonding to phosphorus, and a 90° value for all *cis*-L-M-L bond angles was used. For chlorine at site 1 in **21a**, the two *P*-methyl carbons are each 3.0 Å away from the chlorine. A chlorine at site 2 is also 3.0 Å away from four proximate *tert*-butyl methyl carbons. For each of these CH₃/Cl interactions, the van der Waals radii of methyl and chlorine overlap by about 0.8 Å, which should be repulsive. Admittedly, the actual equilibrium geometries of **1** and **2** do not have C_{2v} symmetry. Twisting about bonds and bond angle distortions will occur.⁹⁻¹¹ Nevertheless, it is apparent that the greater number of methyl groups proximate to site 2 in **21a** renders site 2 more crowded than site 1. The smaller carbon monoxide should prefer to locate at site 2 and the larger chlorine at site 1. Thus, we assign the major syn form of **1** or **2** to **17** (eq 1). The major B₂X and B₂ signals in the ³¹P{¹H} spectra of **1** and **2** at about 210 K (Table I) are assigned to **17** and the minor D₂X and D₂ signals to **18**.

In an anti form, the larger chlorine ligand will experience three CH₃/Cl nonbonded repulsions, which should render the anti forms intermediate in stability between the two syn conformations. Indeed, this is observed (Table I).

(17) The carbon monoxide ligand was treated, to a first approximation, as a cylinder with a length (*l*), radius (*r*) and volume (*V*). The length (*l*) was derived from available C=O bond length and van der Waals radii for carbon and oxygen and the volume from the work of Bondi.¹⁸ With the dimensions thus set, the equation for the volume of a cylinder was solved for *r* = 1.4 Å.

(18) (a) Bondi, A. *J. Phys. Chem.* **1964**, *68*, 441. (b) Slater, J. C. *J. Chem. Phys.* **1964**, *41*, 3199.

(19) Stevenson, P. E.; Merrill, J. E. Quantum Chemistry Program Exchange, Indiana University; Program 186.

**Figure 10.** Decomposition of the ³¹P{¹H} NMR spectrum of **4** at 223 K.

Now that these assignments have been made, certain structural variations should produce predictable changes in conformer populations. Specifically, increasing the steric size of halogen should enhance the relative stability of conformer **17** and decrease the relative stabilities of **18–20**. In addition, changing the PCH₃ groups to PH moieties should enhance the difference between the degrees of crowding at sites 1 and 2 in **21b** as compared to the case for **21a**. In **21b** site 1 should be substantially less crowded than site 1 in **21a**, while site 2 remains essentially unchanged. Larger halogen ligands will be much better accommodated at site 1 in **21b**. Thus, changing PCH₃ to PH groups should also enhance the relative stability of **17** as compared to **18–20**.

Effect of Halogen Steric Bulk on Conformational Preference. *trans*-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)Br (**3**). The ³¹P{¹H} NMR spectrum (101.2 MHz) of the bromo complex **3** (0.03 M in toluene-*d*₈) at 370 K is a broadened doublet at δ 36.9 (¹J_{RhP} = 120 Hz). Below 370 K, the spectrum decoalesces and, at 214 K, is sharpened into three subspectra (see Figure 7, supplementary material). A decomposition of the 214 K spectrum (see Figure 8, supplementary material) shows a major B₂X doublet (**17**; 66.6%), a minor D₂X doublet (**18**; 3.4%) and an ACX subspectrum (**19** and **20**; 30.0%). NMR parameters are compiled in Table I. This DNMR behavior is analogous to that for **1** but with important differences in subspectral populations. As compared to the case for **1**, the population of **17** has increased by 6.7% while the populations of **18** as well as **19** and **20** have decreased by 3.6% and 3.1%, respectively (Table I). The relative stability of conformer **17** has increased while the stabilities of **18–20** have decreased, as expected.

trans-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)I (**4**). The ³¹P{¹H} NMR spectrum of **4** shows a characteristic decoalescence (see Figure 9, supplementary material). A decomposition of the slow-exchange spectrum at 223 K (Figure 10) shows the typical B₂X (**17**; 83.8%), D₂X (**18**; 0.6%), and ACX (**19** and **20**; 15.6%) subspectra (Table I). In Figure 10, those spectra marked ×10 are displayed at an amplification 10 times larger than the B₂X subspectrum and the bottom composite spectrum. As compared to the case for the bromo complex **3**, there is another significant increase in the relative population of conformer **17** (+17.2%) and decreases for **18** (-2.8%) as well as **19** and **20** (-14.4%), as expected.

trans-[(C₄H₉)₂PCH₃]₂Ir(CO)Br (**5**). The ³¹P{¹H} NMR spectrum of **5** (0.03 M in toluene-*d*₈) at 375 K shows a highly exchange-broadened signal at δ 31.5 (see Figure 11, supplementary material). The degree of exchange broadening for **5** at 375 K is substantially greater than for **2** at 360 K (Figure 4). This is consistent with higher rotation barriers due to a sterically larger

bromine. At 214 K, the spectrum is separated into B₂ (**17**; 70.5%), D₂ (**18**; 3.2%), and AC (**19** and **20**; 26.3%) subspectra (see Figure 12, supplementary material; Table I). As compared to the case for the chloro analogue (**2**), there is an increase in the relative population of **17** (+9.8%) and corresponding decreases for **18** (-3.1%) as well as **19** and **20** (-6.7%).

trans-[(t-C₄H₉)₂PCH₃]₂Ir(CO)I (**6**). The ³¹P{¹H} spectrum of **6** (0.03 M in toluene-*d*₈) at 310 K already shows a substantially decoalesced spectrum (Figure 13, supplementary material). This reveals higher rotation barriers about Ir-P bonds than in the chloro analogue and a sterically larger iodine. The 242 K slow-exchange spectrum shows major B₂ (**17**; 84.5%), very minor D₂ (**18**; 0.6%), and AC (**19** and **20**; 14.9%) subspectra (Figure 14, supplementary material).

At this point, it should be stated that complete DNMR line-shape analyses for iodo complexes **4** and **6** show that the minor D₂X or D₂ signals observed at low temperature do indeed coalesce with the rest of the spectrum at higher temperatures. The D₂X and D₂ signals are not due to minor impurities and are due to molecular conformations associated with intramolecular exchange in **4** and **6**.

A perusal of Table I reveals a clear-cut trend. As the van der Waals radius and cone angle of halogen increases, the relative stability of **17** increases while the stabilities of **18**, **19**, and **20** decrease. This is consistent with increasing steric bulk in proceeding from chlorine to bromine to iodine and an increasing differential between repulsions experienced by halogen at sites 1 and 2 in **21a**. As halogen steric bulk increases, there is a progressively greater preference for halogen to locate at the less sterically crowded site 1 in **21a** than at site 2 and an increasing preference for conformation **17**. Likewise, as halogen steric bulk increases, nonbonded methyl/halogen repulsions should increase in the anti forms **19** and **20**. The relative stabilities of **19** and **20** should decrease, as observed (Table I). These trends are entirely consistent with our conformational assignments.

Effect of the P-R Group on Conformational Preference. Consistent with our rationale for conformational assignments (vide supra), changing from P-CH₃ to P-H groups should enhance the relative stability of syn form **17** as compared to **18-20**.

trans-[(t-C₄H₉)₂PH]₂Rh(CO)Cl (**7**). The ³¹P{¹H} NMR spectrum (101.2 MHz) of **7** (0.04 M in 20% CDCl₃/80% toluene-*d*₆) at 370 K shows an exchange-broadened resonance (δ 74.3), which decoalesces at lower temperatures into a familiar set of three subspectra (see Figures 15 and 16, supplementary material). NMR parameters are compiled in Table I. At 223 K, the spectrum shows B₂X (**17** in eq 1; 78.2%), D₂X (**18**; 1.0%), and ACX (**19** and **20**; 20.8%) subspectra.

One comparison is noteworthy. For **7**, the population of the dominant syn form **17** is substantially higher while minor syn and anti populations are lower than in the di-*tert*-butylmethylphosphine analogue **1** (Table I). This is consistent with the fact that the larger halogen ligand can locate at the substantially less crowded site 1 in **21b** (as compared to site 1 in **21a**). This leads to an enhanced relative stability for conformer **17**.

trans-[(t-C₄H₉)₂PH]₂Ir(CO)Cl (**8**). The ³¹P{¹H} spectrum (101.2 MHz) of **8** (0.03 M in CDCl₃) decoalesces in typical fashion (see Figure 17, supplementary material), revealing at 223 K dominant B₂ (**17**; 90.0%), AC (**19** and **20**; 9.8%), and very minor D₂ (**18**; 0.2%) subspectra (see Figure 18 (supplementary material) and Table I). For **8**, conformer **17** is once again more stable than for the methyl analogue **2**.

trans-[(t-C₄H₉)₂PH]₂Rh(CO)Br (**9**) and trans-[(t-C₄H₉)₂PH]₂Ir(CO)Br (**10**). The ³¹P{¹H} NMR spectra of **9** (0.03 M in CDCl₃) and **10** (0.03 M in CDCl₃) decoalesce in typical fashion. NMR parameters and subspectral populations are compiled in Table I. As expected, the relative stability of **17** has increased for **9** and **10** as compared to that for the chloro complexes **7** and **8**. For **9** and **10**, the minor syn conformer is present at minuscule concentrations (0.1-0.3%).

trans-[(t-C₄H₉)₂PH]₂Rh(CO)I (**11**) and trans-[(t-C₄H₉)₂PH]₂Ir(CO)I (**12**). The ³¹P{¹H} NMR spectra of **11** (0.03 M in CDCl₃) and **12** (0.03 M in toluene-*d*₈) also decoalesce in

typical fashion. For **11** at 214 K, there is a strongly dominant B₂X doublet (**17**; 93.5%) and an ACX subspectrum (**19** and **20**; 6.5%). Even after many hours of signal averaging, we could detect no D₂X subspectrum. For **12** at 214 K, there is an even more strongly dominant B₂ singlet (**17**; 97.5%) and an AC subspectrum (**19** and **20**; 2.5%). We could detect no D₂ singlet. NMR parameters and subspectral populations are compiled in Table I. For **11** and **12**, the minor syn species (**18**; eq 1) is present in too low a concentration to be NMR detectable, and syn form **17** is strongly dominant.

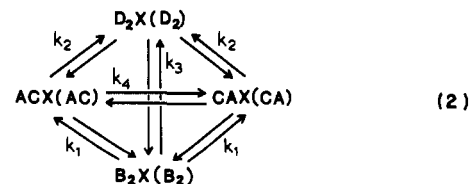
Trends in Conformational Preference. The consistency between our rationale for conformational assignments and the trends in conformational preference determined from the ³¹P{¹H} NMR data (vide supra) strongly suggest conformer **17** to be the most stable equilibrium conformation.

For both series of complexes (**1-6**, **7-12**), proceeding from iodine to bromine to chlorine corresponds to a reduction in steric size for halogen that is progressively more comparable to carbon monoxide. The differential between methyl/halogen and methyl/halogen repulsions should progressively decrease with decreasing halogen van der Waals radius and the population differences between conformers should also decrease, as observed (Table I). The relative stabilities of the minor syn form **18** and anti forms (**19**, **20**) progressively increase as halogen size decreases.

Comparisons between pertinent pairs of the two series **1-6** and **7-12** (Table I) will also show that changing from R = H to R = CH₃ results in a destabilization of the major syn conformer (**17**) and increased relative stabilities for **18-20**. This trend is consistent with the fact that the difference between nonbonded repulsions felt by halogen at sites 1 and 2 in **21a** is smaller than in **21b**. Steric crowding at site 1 in **21a** is more severe than it is in **21b**. The net result is a trend toward equalizing conformer populations in the *P*-methyl analogues.

³¹P{¹H} Chemical Shift Trends. Now that the conformational assignments above have been made, some potentially useful trends in ³¹P{¹H} chemical shifts can be deduced as follows: (1) The chemical shift of the major syn species (**17**) with the P-R groups syn to halogen is in the higher frequency (downfield) region of the spectrum (Table I). (2) The chemical shift of the minor syn species (**18**) with the P-R groups syn to carbon monoxide is in the lower frequency (upfield) region of the spectrum. (3) For the anti species, the higher frequency signal is due to the P-R group that is syn to halogen and the lower frequency signal is due to the P-R group that is syn to carbon monoxide.

Dynamics of Rotation about Metal-Phosphorus Bonds. A ³¹P{¹H} NMR line-shape simulation model for complexes **1-8** requires the incorporation of four unique spin systems: B₂X (or B₂), D₂X (or D₂), ACX (or AC), and CAX (or CA).²⁰ While the two anti forms **19** and **20** are equivalent, they can be interconverted directly by simultaneous 180° rotations of both phosphines about the M-P bonds. During this process, the two diastereotopic phosphorus atoms interchange chemical shifts (i.e., ACX to CAX or AC to CA). Therefore, the direct anti to anti conversion (e.g., **19** to **20**) is in principle a DNMR-visible process, as are all the other possible conformational exchanges shown in eq 1. A general equation that catalogues all possible subspectral exchanges for complexes **1-8** is shown in eq 2. Equation 2



correlates directly to eq 1 in terms of spin systems, the conformational assignments made above, and rate constants. Six rate constants are shown in eq 1 and 2. The corresponding reverse rate constants are all implied and are automatically calculated

(20) DNMR simulations were not performed for complexes **9-12**.

Table IV. Activation Parameters for Rotation about the Metal-Phosphorus Bonds in Complexes 1-8

compd	spin exchange obsd	ΔH^\ddagger , kcal/mol	ΔS^\ddagger , cal/mol-deg	ΔG^\ddagger , kcal/mol (temp, K)
1	B ₂ X to ACX	12.9 ± 1.5	-2 ± 7	13.4 ± 0.2 (266)
	ACX to D ₂ X	11.9 ± 1.5	-3 ± 7	12.7 ± 0.2 (266)
2	B ₂ to AC	14.4 ± 1.5	2 ± 7	13.9 ± 0.2 (271)
	AC to D ₂	12.2 ± 1.5	-5 ± 7	13.5 ± 0.2 (271)
3	B ₂ X to ACX	13.1 ± 1.5	-4 ± 7	14.1 ± 0.2 (271)
	ACX to D ₂ X	12.9 ± 1.5	-4 ± 7	13.9 ± 0.2 (271)
4	B ₂ X to ACX	11.8 ± 1.5	-10 ± 7	14.7 ± 0.2 (290)
	ACX to D ₂ X	12.2 ± 1.5	-7 ± 7	14.4 ± 0.2 (290)
5	B ₂ to AC	13.3 ± 1.5	-6 ± 7	15.2 ± 0.2 (300)
	AC to D ₂	13.6 ± 1.5	-4 ± 7	14.9 ± 0.2 (300)
6	B ₂ to AC	16.0 ± 1.5	1 ± 7	15.8 ± 0.2 (300)
	AC to D ₂	14.5 ± 1.5	-3 ± 7	15.4 ± 0.2 (300)
7	B ₂ X to ACX	11.4 ± 1.5	-7 ± 7	13.5 ± 0.2 (271)
	ACX to D ₂ X	11.1 ± 1.5	-7 ± 7	13.1 ± 0.2 (271)
8	B ₂ to AC	12.0 ± 1.5	-8 ± 7	14.2 ± 0.2 (280)
	AC to D ₂	11.5 ± 1.5	-8 ± 7	13.9 ± 0.2 (280)

in the DNMR line-shape calculations.¹³

The ³¹P{¹H} DNMR spectra of 1-8 decoalesce above 200 K due to slowing rotation about M-P bonds. Excellent line-shape simulations were achieved with k_3 and k_4 values in eq 2 equal to zero and non-zero values for k_1 and k_2 . Incorporation of k_3 or k_4 values that were above 10% of the other rate constants produced subtle but discernible degradations in the DNMR line-shape simulations.

Although the error in temperature measurement at the NMR sample is ±3 K, decoalescence generally occurs over a large temperature range (ca. 120 K) and we feel justified in reporting ΔH^\ddagger , ΔS^\ddagger , and ΔG^\ddagger values for M-P rotation from least-squares fits of Eyring plots (Table IV). Of course, ΔG^\ddagger values must be considered to be the most accurate of the three activation parameters.

There are implications to be derived from the DNMR simulations. The requirement that k_3 and k_4 values equal zero says the direct syn to syn (e.g., 17 and 18) and direct anti to anti (19 to 20) conversions occur at rates that are slower than syn to anti exchanges. The preferred itinerary for exchange involves stepwise interconversions around the periphery of eq 1 or 2. One phosphine at a time rotates 180°, presumably passing through two maxima involving *t*-C₄H₉/halogen and *t*-C₄H₉/CO eclipsings and via an unstable intermediate (e.g., see 16). The DNMR simulations show that M-P rotation barriers do increase with increasing steric size of halogen and the barriers in the iridium complexes are slightly higher than in the rhodium complexes (Table IV).

Experimental Section

The 101.2-MHz ³¹P{¹H} and 62.9-MHz ¹³C{¹H} NMR spectra were recorded with use of a Bruker WM-250 NMR system at the University of Vermont. The 67.9-MHz ¹³C{¹H} spectra were recorded on a Bruker HX-270 NMR system at the NSF Regional Instrumentation Center at Yale University. Variation of NMR sample temperature was achieved with use of a custom-built nitrogen gas delivery system, and temperature was controlled with a Bruker BVT-1000 control unit. Temperatures could be maintained at a constant value (±0.1 K) but are accurate to only ±3 K at the sample. NMR samples were prepared in precision NMR tubes on a vacuum line and were sealed after a minimum of three freeze/pump/thaw cycles (0.01 torr).

Mass spectral analyses were provided by the Analytical Section of Warner-Lambert/Parke-Davis Pharmaceuticals, Ann Arbor, MI. Mass measurements were made by using a high-resolution VG7070 E/HF mass spectrometer operating in either the fast atom bombardment (FAB)

or desorption electron ionization (DEI) mode. For the FAB experiments, the matrix used is thioglycerol and the reagent gas used is xenon.

Syntheses of complexes 1, 2, 7, and 8 have been described previously.^{5b}

trans-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)Br (3) was prepared from 1 by six separate treatments with a stirred 6 M excess of lithium bromide in methanol at reflux for 24 h. After each exchange, the mixture was cooled to 273 K and the precipitated complex isolated by filtration. Examination of the ³¹P{¹H} and ¹³C{¹H} NMR spectra after each exchange showed a smooth and eventually complete conversion to 3. Following the last exchange, 3 was washed with methanol and recrystallized from 50% toluene/50% methanol. NMR data: Table I. Mass spectral data: *m/e* 530 (FAB); calcd *M_r*, 531.4. Anal. Calcd for C₁₉H₄₂BrOP₂Rh: C, 42.9; H, 7.97; P, 11.7; Br, 15.0. Found: C, 43.1; H, 7.92; P, 11.5; Br, 14.7.

trans-[(*t*-C₄H₉)₂PCH₃]₂Rh(CO)I (4) was prepared from 1 with use of sodium iodide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 578 (FAB); calcd *M_r*, 578.4. Anal. Calcd for C₁₉H₄₂IOP₂Rh: C, 39.5; H, 7.32; P, 10.7; I, 21.9. Found: C, 39.4; H, 7.36; P, 10.2; I, 21.0.

trans-[(*t*-C₄H₉)₂PCH₃]₂Ir(CO)Br (5) was prepared from 2 with use of lithium bromide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 620 (FAB); calcd *M_r*, 620.6. Anal. Calcd for C₁₉H₄₂BrOP₂Ir: C, 36.8; H, 6.82; P, 10.0; Br, 12.9. Found: C, 36.0; H, 6.82; P, 9.42; Br, 12.0.

trans-[(*t*-C₄H₉)₂PCH₃]₂Ir(CO)I (6) was prepared from 2 with use of sodium iodide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 668 (FAB); calcd *M_r*, 667.6. Anal. Calcd for C₁₉H₄₂IOP₂Ir: C, 34.2; H, 6.34; P, 9.28; I, 19.0. Found: C, 34.2; H, 6.44; P, 9.07; I, 18.7.

trans-[(*t*-C₄H₉P)₂PH]₂Rh(CO)Br (9) was prepared from 7 with use of lithium bromide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 502 (DEI); calcd *M_r*, 503.2. Anal. Calcd for C₁₇H₃₈BrOP₂Rh: C, 40.6; H, 7.61; P, 12.3; Br, 15.9. Found: C, 40.7; H, 7.70; P, 11.7; Br, 14.5. A persistent trace of starting material (7) is observed in the ³¹P{¹H} NMR spectrum and the sample did give an analysis for some chlorine (1.25%).

trans-[(*t*-C₄H₉)₂PH]₂Ir(CO)Br (10) was prepared from 8 with use of lithium bromide and the procedure described for 3. NMR data: Table I. Mass spectral data: *m/e* 592 (DEI); calcd *M_r*, 592.5. Anal. Calcd for C₁₇H₃₈BrOP₂Ir: C, 34.5; H, 6.46; P, 10.5; Br, 13.5. Found: C, 35.6; H, 6.80; P, 9.29; Br, 12.3. A persistent trace of starting material (8) is observed in the ³¹P{¹H} NMR spectrum and the sample did give an analysis for some chlorine (0.95%).

trans-[(*t*-C₄H₉)₂PH]₂Rh(CO)I (11) was prepared from 7 with use of sodium iodide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 550 (DEI); calcd *M_r*, 550.2. Anal. Calcd for C₁₇H₃₈IOP₂Rh: C, 37.11; H, 6.96; P, 11.3; I, 23.1. Found: C, 37.14; H, 7.03; P, 11.0; I, 22.8.

trans-[(*t*-C₄H₉)₂PH]₂Ir(CO)I (12) was prepared from 8 with use of sodium iodide and the procedure described for 3 above. NMR data: Table I. Mass spectral data: *m/e* 640 (DEI); calcd *M_r*, 639.5. Anal. Calcd for C₁₇H₃₈IOP₂Ir: C, 31.93; H, 5.99; P, 9.69; I, 19.84. Found: C, 31.97; H, 6.01; P, 9.59; I, 19.5.

Acknowledgment. We are grateful to the National Science Foundation for financial support (Grant Nos. CHE-8024931 and CHE-8306876) and to the University of Vermont Academic Computing Center for outstanding computational support. We are also grateful to Steve Werness and Dana DeJohn of Warner-Lambert/Parke-Davis Pharmaceuticals for running the mass spectra.

Registry No. 1, 34365-67-8; 2, 34365-68-9; 3, 101403-82-1; 4, 101403-83-2; 5, 101403-84-3; 6, 101403-85-4; 7, 33246-87-6; 8, 33246-91-2; 9, 33246-88-7; 10, 33246-92-3; 11, 33246-89-8; 12, 33246-93-4.

Supplementary Material Available: ³¹P{¹H} and ¹³C{¹H} DNMR spectra of complexes 2-8 and decompositions of slow-exchange ³¹P{¹H} NMR spectra for complexes 2, 3, and 5-8 (Figures 4-9 and 11-18) (14 pages). Ordering information is given on any current masthead page.