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fumaronitrile, 764-42-1; squaric acid, 2892-51-5; allyl alcohol, 107-18-6; K<sub>4</sub>[Pt<sub>2</sub>(pop)<sub>4</sub>], 79716-40-8.

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## Mechanism of Oxygenation of cis-Stilbene by Iron Bleomycin

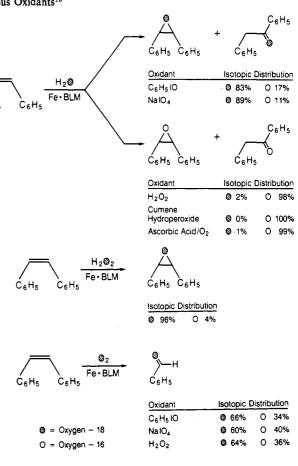
Sir:

The bleomycins (BLM's) are a family of antitumor antibiotics isolated from Streptomyces verticillus.<sup>1</sup> They mediate DNA strand scission in the presence of  $O_2$  and certain metal ions;<sup>2</sup> this transformation is believed to constitute the basis for their antitumor activity.<sup>3</sup> Metallobleomycins may also be activated for DNA strand scission by the use of oxidants such as iodosobenzene.<sup>4</sup> In addition to mediating destruction of DNA, activated metallobleomycins have been shown to effect the oxidation of several low molecular weight olefinic substrates.4a,4c,5

Although the products<sup>1a,6</sup> and chemical mechanism<sup>1a,6,7</sup> of DNA cleavage by bleomycin have been studied in some detail, as have the products resulting from BLM-mediated oxidation of small molecules, 4a,4c,5 the only published oxygen-labeling experiments identified the aqueous medium as the source of oxygen in cisstilbene oxide formed in the  $C_6H_5IO$ -supported oxidation of *cis*-stilbene by FeBLM.<sup>5b</sup> Presently, we report the results of a more thorough study in which we employed oxygen labeling to help define the mechanism of small-molecule oxidation by BLM. The results of this study suggest that the source of oxygen transferred in the formation of cis-stilbene oxide is a metal-oxo species, the oxygen in which is derived from the oxidant employed

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Scheme I. Isotopic Distribution of <sup>18</sup>O and <sup>16</sup>O in Products Resulting from FeBLM-Dependent Oxidation of cis-Stilbene in the Presence of Various Oxidants<sup>10</sup>



for metallobleomycin activation.

When carried out under aerobic conditions, the FeBLM-mediated oxidation of cis-stilbene afforded a number of products, including cis-stilbene oxide and benzaldehyde.4,5,8 Previous work suggested that these two products resulted from two different oxidation pathways: one of these was suggested to be an oxygen-transfer reaction leading to cis-stilbene oxide; the other, an electron-abstraction pathway yielding the stilbene cation radical, which reacted with  $O_2$  to form benzaldehyde.<sup>8</sup> The oxygen-18labeling data presented in Scheme I are consistent with this hypothesis.<sup>9</sup> For each of the oxidants tested, the benzaldehyde oxygen was derived primarily from  $O_2$ .<sup>10</sup> In contrast, the source of oxygen in cis-stilbene oxide depended on the oxidant used to support the reaction. In all cases, the labeling of deoxybenzoin

- Soc. 1986, 108, 7839. In a typical H<sub>2</sub><sup>18</sup>O-labeling experiment, 22  $\mu$ L of an aqueous solution (9) containing preformed 9.1 mM Fe<sup>III</sup>BLM was added to an (aerobic) solution that had been formed by admixture of 150  $\mu$ L of methanol containing 100 mM cis-stillene and 285  $\mu$ L of 4:1 CH<sub>3</sub>OH-H<sub>2</sub><sup>18</sup>O (50% <sup>18</sup>O content in H<sub>2</sub>O). The reaction was initiated by the addition of 50  $\mu$ L of 4:1 CH<sub>3</sub>OH-H<sub>2</sub><sup>16</sup>O containing 40 mM oxidant. The reaction mixture was maintained at 25 °C for 1 h and then treated with 2 mL of 0.5% aqueous NaCl. The reaction mixture was extracted with 1 mL of CH<sub>2</sub>Cl<sub>2</sub>, and the organic extract was concentrated and analyzed on a 15 m DB-17 column in a Finnigan-MAT 4610 gas chromatographmass spectrometer equipped with an INCOS data system. Reactions conducted under  ${}^{18}O_2$  ( ${}^{18}O$  content 98%) were performed similarly, with the exception that the reaction mixtures were degassed by alternate cycles of vacuum/argon purging in capped 1-mL reacti-vials (Wheaton Scientific) prior to addition of the oxidant. Following addition of <sup>18</sup>O<sub>2</sub> via gastight syringe to a final pressure of approximately 1 atm, a degassed solution of oxidant was added to initiate the reaction. (10) In addition to the exchange of the  $C_6H_5CHO$  oxygen with  $H_2O$  under
- The reaction conditions (23%), some  $C_{eH_2}CHO$  formation was also observed to occur in a BLM- and Fe-independent fashion. Therefore, the <sup>18</sup>O-labeling efficiency of C<sub>6</sub>H<sub>5</sub>CHO by the oxidative pathway envisioned<sup>8</sup> is significantly understated in Scheme I.

Heimbrook, D. C.; Mulholland, R. L., Jr.; Hecht, S. M. J. Am. Chem. (8)

**Table I.** <sup>18</sup>O Content of  $(C_6H_5)_3P$ =O Formed by  $(C_6H_5)_3P$ Oxidation following  $C_6H_5IO/H_2O$  Exchange<sup>a</sup>

incubation time, min	solvent system	
	5% H <sub>2</sub> <sup>18</sup> O <sup>b</sup> - 95% CH <sub>3</sub> OH	40% H <sub>2</sub> <sup>18</sup> O <sup>c</sup> - 60% CH <sub>3</sub> OH
0	0	0
$0.5^{d}$	90	
15	91	82
45	92	82
75	89	82

<sup>*a*</sup>Also included in the incubation was  $(CH_3C_6H_4)_3P=^{16}O$  to assess possible exchange of triarylphosphine oxide with water; no exchange was observed. All experiments were carried out at 25 °C, except as noted. <sup>*b*</sup>97 atom % <sup>18</sup>O. <sup>*c*</sup>35 atom % <sup>18</sup>O. <sup>*d*</sup>Carried out at -10 °C.

was qualitatively and quantitatively the same as for *cis*-stilbene oxide (cf. ref 8). In the  $H_2O_2$  and cumene hydroperoxide supported reactions, for example, no <sup>18</sup>O incorporation was observed from either water or  $O_2$ , suggesting strongly that the source of epoxide oxygen was the oxidant itself. In fact, when the epoxidation was repeated with  $H_2$ <sup>18</sup> $O_2$ ,<sup>11</sup> the derived *cis*-stilbene oxide was found to contain predominantly <sup>18</sup>O (Scheme I). The source of the epoxide oxygen was also established directly where Fe<sup>III</sup>BLM was activated in the presence of ascorbate + <sup>18</sup> $O_2$ ; the formed *cis*-stilbene oxide contained  $\simeq 90\%$  <sup>18</sup>O in the epoxide oxygen atom. In the C<sub>6</sub>H<sub>5</sub>IO- and NaIO<sub>4</sub>-supported reactions, however, the epoxide oxygen was derived primarily from water.

One possible reason for the presence of  $H_2O$ -derived oxygen in *cis*-stilbene oxide in the C<sub>6</sub>H<sub>5</sub>IO and NaIO<sub>4</sub>-supported reactions could be preequilibration of the oxidant oxygens with water prior to FeBLM activation. Although C<sub>6</sub>H<sub>5</sub>IO has been reported not to exchange with water, <sup>5b,12</sup> few of the reports contain any experimental detail; one of those that does has subsequently been questioned, <sup>12e</sup> and other studies have reported oxygen exchange. <sup>13</sup> Accordingly, we sought to measure oxygen exchange directly, using CH<sub>3</sub><sup>16</sup>OH-H<sub>2</sub><sup>18</sup>O + C<sub>6</sub>H<sub>5</sub>I<sup>16</sup>O under conditions comparable to those employed in the BLM-mediated oxygen-transfer experiments reported here.<sup>8</sup> The exchange reactions were quenched by the addition of triphenylphosphine; mass spectrometric analysis of triphenylphosphine oxide provided a measure of oxygen exchange. As shown in Table I, under the conditions of our experiment,

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exchange with H<sub>2</sub>O was complete within 30 s.<sup>14</sup> Also studied by using the same technique was the exchange of periodate with H<sub>2</sub><sup>18</sup>O; consistent with an earlier report,<sup>15</sup> exchange was found to occur rapidly. Clearly, the incorporation of <sup>18</sup>O into *cis*-stilbene oxide from H<sub>2</sub><sup>18</sup>O will depend on the relative rates of oxygen exchange into the oxidant vs formation of the metal–oxo species. The accumulated data suggest that the observed incorporation of <sup>18</sup>O into *cis*-stilbene oxide from H<sub>2</sub><sup>18</sup>O during the C<sub>6</sub>H<sub>5</sub>IO- and NaIO<sub>4</sub>-supported FeBLM reactions is consistent with the direct transfer of oxygen to *cis*-stilbene from a high-valent metal–oxo intermediate. Accordingly, the present <sup>18</sup>O-labeling study supports the mechanistic scheme proposed earlier,<sup>8</sup> in which FeBLM was suggested to mediate the transformation of *cis*-stilbene both by electron-abstraction and oxygen-transfer pathways.

It has been noted previously that the products formed from small substrate molecules with FeBLM are strikingly similar to those obtained with cytochrome P-450 and related model systems.<sup>5b</sup> The <sup>18</sup>O-labeling patterns observed here further extend this analogy. For example, the 5-exo-hydroxylation of camphor by *Pseudomonas putida* cytochrome P-450 was accomplished by <sup>18</sup>O incorporation from H<sub>2</sub><sup>18</sup>O when C<sub>6</sub>H<sub>5</sub>IO was the exogenous oxidant, but not when activation was carried out with NADH/O<sub>2</sub>, *m*-chloroperbenzoic acid, or cumene hydroperoxide.<sup>12c</sup> Essentially the same results were obtained for the hydroxylation of cyclohexane by liver microsomal cytochrome P-450.<sup>12d</sup> In total, these results reinforce earlier suggestions that FeBLM-mediated oxidation of small molecules shares many common characteristics with cytochrome P-450.<sup>16</sup>

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