

## Proton NMR Spectroscopy as a Probe of Dinuclear Copper(II) Centers

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Enzymes containing dinuclear Cu(II) centers play important roles in nature and, consequently, characterization of their structure and function is a problem of outstanding importance.<sup>1,2</sup> A fundamental and, as yet, largely unexplored issue is the determination of the structural and magnetic properties of dinuclear Cu(II) centers using NMR spectroscopy. <sup>1</sup>H NMR is a natural technique to probe these systems because only protons proximate to the paramagnetic center are affected.<sup>3,4</sup> However, the slow electronic relaxation typical of Cu(II) ions makes this type of study extremely difficult to execute, and correspondingly few examples exist in the literature.<sup>5–12</sup> In an effort to gain insight into the structures of dinuclear Cu(II) metalloprotein active sites and model complexes, we have applied one- and two-dimensional <sup>1</sup>H NMR techniques to an antiferromagnetically coupled ( $\mu$ -phenoxo)( $\mu$ -hydroxo)dicopper(II) complex. Clear COSY cross-signals are observed between hyperfine shifted signals allowing the complete assignment of the <sup>1</sup>H NMR spectrum. These data, coupled with X-ray crystallographic results, indicate that a paramagnetic dipolar relaxation mechanism is the dominant proton relaxation pathway.

The previously reported ( $\mu$ -phenoxo)( $\mu$ -hydroxo)dicopper(II) complex [Cu<sub>2</sub>(BPMP)(OH)](ClO<sub>4</sub>)<sub>3</sub> (**1**) was synthesized and crystallographically characterized.<sup>13–15</sup> Complex **1** exhibits several sharp, isotropically shifted <sup>1</sup>H NMR signals in acetonitrile solution at 55 °C in the 150 to –50 ppm chemical shift range (Figure 1,<sup>25</sup> Table 1). All of the isotropically shifted signals sharpen and shift toward the diamagnetic region as the temperature is increased. While magnetic data have not been

reported for **1**, –2*J* values for several related ( $\mu$ -phenoxo)( $\mu$ -hydroxo)dicopper(II) complexes have been reported.<sup>16–19</sup> All of these complexes exhibit moderate to strong antiferromagnetic coupling between the Cu(II) centers with –2*J* values greater than 100 cm<sup>–1</sup>. Using the Evans susceptibility method,<sup>20,21</sup> the room temperature magnetic moment ( $\mu_{\text{eff}}/\text{Cu}$ ) of **1** was found to be 1.27  $\mu_{\text{B}}$  which gives the number of unpaired electrons (*n*/Cu) as 0.62. These data indicate that the Cu(II) ions in **1** are moderately antiferromagnetically coupled. Since the magnetic moment of the complex is small, relatively sharp isotropically shifted NMR signals are observed.<sup>12</sup>

Several of the isotropically shifted <sup>1</sup>H NMR signals observed for **1** can be initially assigned by inspection of their peak areas. Signals D (23.6 ppm), E (22.2 ppm), F (14.6 ppm), G (9.93 ppm), and H (5.12 ppm) integrate to 4:4:2:4:3 protons, respectively (Table 1). These data taken together with the crystallographic results suggest that signals D, E, and G arise from pyridyl protons while signals F and H are due to the *m*-phenol and the *p*-methylphenol protons, respectively. Definitive assignment of each of these signals comes from two-dimensional NMR techniques. A magnitude COSY spectrum of **1** was recorded at 25 °C and clearly shows cross-signals between resonances D and G and also between resonances E and G (Figure 2). These signals can be unequivocally assigned to the pyridine  $\beta$ -H (E),  $\beta'$ -H (D), and  $\gamma$ -H (G) protons, respectively.

Assignment of the remaining signals of **1** comes from *T*<sub>1</sub> values<sup>22</sup> and comparison of the spectrum of **1** with that of a related complex [Cu<sub>2</sub>(CH<sub>3</sub>HXTA)(OH)]<sup>2–</sup> (**2**) where the methylpyridyl ligands have been replaced by acetate moieties.<sup>23,24</sup> The <sup>1</sup>H NMR spectrum of **2** shows five isotropically shifted signals at 55 °C in D<sub>2</sub>O and pH 10 (Figure 1<sup>25</sup>). Signals D (16.8 ppm; 7 ms) and E (9.1 ppm; 25 ms) integrate to two and three protons, respectively, and are thus assigned to the *m*-phenol and *p*-methylphenol protons, respectively. Signals B (69 ppm; ~1 ms) and C (62 ppm; ~1 ms) are assigned to the diastereotopic  $\beta$ -CH<sub>2</sub> acetate protons based upon their relative integrations, *T*<sub>1</sub> values, and the fact that they are selectively deuterated at 90 °C and pH 10. The only remaining protons in **2** unassigned are the  $\beta$ -CH<sub>2</sub> protons of the phenol–methylamine linkage. These protons can therefore be assigned to signal A (143 ppm). Comparison of the chemical shift, *T*<sub>1</sub> values, and

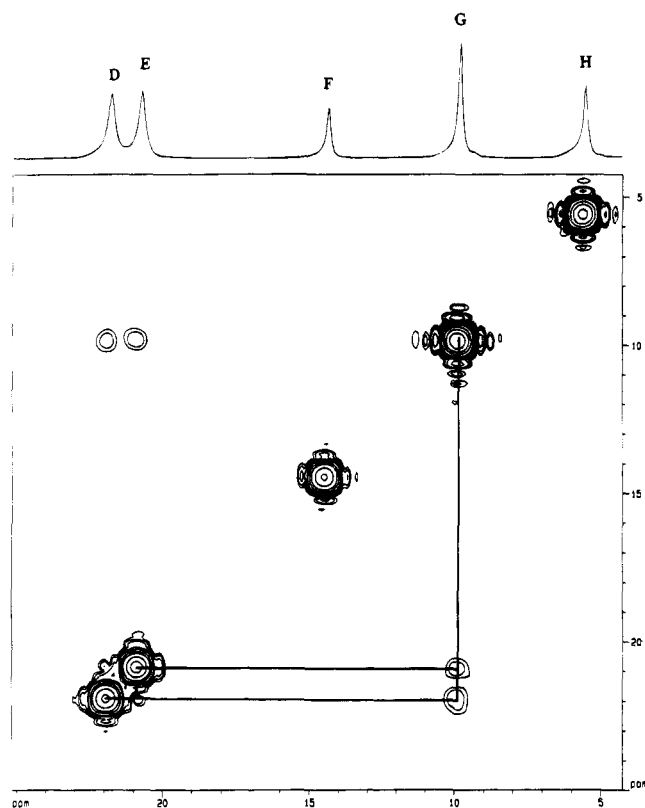
- (1) Sorrell, T. N. *Tetrahedron* **1989**, *45*, 3–68.
- (2) Karlin, K. D.; Tyeklar, Z. *Bioinorganic Chemistry of Copper*; Chapman & Hill: New York, 1993.
- (3) Bertini, I.; Luchinat, C. *NMR of Paramagnetic Molecules in Biological Systems*; Benjamin & Cummings: Menlo Park, CA, 1986.
- (4) *NMR Methodology for Paramagnetic Proteins*; La Mar, G. N., de Ropp, J. S., Eds.; Plenum Press: New York, 1993; Vol. 12, pp 1–78.
- (5) Bertini, I.; Luchinat, C. In *Physical Methods for Chemists*, 2nd ed.; Drago, R. S., Ed.; Harcourt Brace Jovanovich: Orlando, FL, 1992; pp 500–556.
- (6) Bertini, I.; Turano, P.; Vila, A. J. *Chem. Rev.* **1993**, *93*, 2833–2932.
- (7) Zelonka, R. A.; Baird, M. C. *Inorg. Chem.* **1972**, *11*, 134–137.
- (8) Wang, S.; Pang, Z.; Zheng, J.-C.; Wagner, M. J. *Inorg. Chem.* **1993**, *32*, 5975–5980.
- (9) Maekawa, M.; Kitagawa, S.; Munakata, M.; Masuda, H. *Inorg. Chem.* **1989**, *28*, 1904–1909.
- (10) Dei, A.; Gatteschi, D.; Piergentili, E. *Inorg. Chem.* **1979**, *18*, 89–93.
- (11) Kitajima, N.; Fujisawa, K.; Fujimoto, C.; Moro-oka, Y.; Hashimoto, S.; Kitagawa, T.; Toriumi, K.; Tatsumi, K.; Nakamura, A. *J. Am. Chem. Soc.* **1992**, *114*, 1277–1291.
- (12) Byers, W.; Williams, R. J. P. *J. Chem. Soc., Dalton Trans.* **1972**, 555–560.
- (13) Maloney, J. J.; Glogowski, M.; Rohrbach, D. F.; Urbach, F. L. *Inorg. Chim. Acta* **1987**, *127*, L33–L35.
- (14) Abbreviations: BPMP = 2,6-bis[[bis(2-pyridylmethyl)amino]methyl]-4-methylphenol; CH<sub>3</sub>HXTA = *N,N'*-(2-hydroxy-5-methyl-1,3-xylylene)bis(*N*-carboxymethylglycine).
- (15) X-ray analysis for [Cu<sub>2</sub>(BPMP)(OH)](ClO<sub>4</sub>)<sub>3</sub> (**1**) (C<sub>33</sub>H<sub>34</sub>N<sub>6</sub>O<sub>10</sub>Cu<sub>2</sub>Cl<sub>2</sub>): monoclinic space group *P2<sub>1</sub>/c*, *a* = 11.843 (3) Å, *b* = 22.126 (7) Å, *c* = 14.356 (6) Å, and  $\beta$  = 105.58 (2)°, with *V* = 3623.8 (16) Å<sup>3</sup>,  $\rho_{\text{calcd}}$  = 1.600 g cm<sup>–3</sup>, and *Z* = 4. A total of 4825 unique reflections out of  $2\theta = 50^\circ$  Mo K $\alpha$  were collected at –100 °C on a Siemens P4 diffractometer equipped with an LT-2a low-temperature device. The structure was solved by Patterson methods and refined anisotropically to *R* = 5.71%, *R<sub>w</sub>* = 6.89% using SHELXTL-PLUS programs (Siemens).

- (16) Murry, K. S. In *Biological and Inorganic Chemistry of Copper*; Karlin, K. D., Zubieta, J., Eds.; Adenine: Guilderland, NY, 1986; Vol. II.
- (17) Karlin, K. D.; Farooq, A.; Hayes, J. C.; Brett, I. C.; Rowe, T. M.; Sinn, E.; Zubieta, J. *Inorg. Chem.* **1987**, *26*, 1271–1280.
- (18) Sorrell, T. N.; Jameson, D. L.; O'Connor, C. J. *Inorg. Chem.* **1984**, *23*, 190–195.
- (19) Oberhausen, K. J.; Richardson, J. F.; Buchanan, R. M.; McCusker, J. K.; Hendrickson, D. N.; Latour, J.-M. *Inorg. Chem.* **1991**, *30*, 1357–1365.
- (20) Evans, D. F. *J. Chem. Soc.* **1959**, 2003–2005.
- (21) Phillips, W. D.; Poe, M. *Methods Enzymol.* **1972**, *24*, 304–317.
- (22) *T*<sub>1</sub> values were obtained using an inversion–recovery pulse sequence (180°– $\tau$ –90°). Plots of  $\ln(I_0 - I_t)$  vs  $\tau$  for each signal provided a straight line over all  $\tau$  values investigated.
- (23) Schwarzenbach, G.; Anderegg, G.; Sallmann, R. *Helv. Chim. Acta* **1952**, *35*, 1785–1792.
- (24) Holz, R. C.; Brink, J. M.; Gobena, F. T. *Inorg. Chem.*, submitted for publication.
- (25) See paragraph at end of paper regarding supplementary material.

**Table 1.**  $^1\text{H}$  NMR (300 MHz) Parameters for  $[\text{Cu}_2(\text{BPMP})(\text{OH})](\text{ClO}_4)_2$  at 55 °C in  $\text{CD}_3\text{CN}$ 

signal	assignt	chem shift (ppm)	line width [fwhm <sup>a</sup> (Hz)]	$T_1$ (ms) <sup>b</sup>	integration <sup>c</sup>	av <sup>d</sup> Cu–H (Å)	calcd <sup>e</sup> Cu–H (Å)
A	Ph–CH <sub>2</sub> –N	130	~4000	5	~4	3.42	4.4
A'	Py $\alpha$ -H	78	1500	3	~4	3.15	4.0
B	N–CH <sub>2</sub> –Py	69	900	4	4	3.68	4.3
C	N–CH <sub>2</sub> –Py	56	475	2	4	3.19	3.8
D	Py $\beta'$ -H	22.4	77	24	4	4.90	5.7
E	Py $\beta$ -H	21.3	66	24	4	5.09	5.7
F	Ph $\beta$ -H	14.3	40	22	2	5.66	5.6
G	Py $\gamma$ -H	9.45	32	56	4	5.75	6.5
H	Ph–CH <sub>3</sub>	5.06	23	103	3	7.31	
I	OH	-33	850	2	~1	2.54	3.4

<sup>a</sup> Full width at half-maximum. <sup>b</sup>  $T_1$  values were obtained using the inversion-recovery method.<sup>22</sup> <sup>c</sup> Integrations are based on the area of signals H and F. <sup>d</sup> Crystallographically determined average distances. All Cu–H average distances are taken as the arithmetic average of equivalent protons to each copper(II) ion. <sup>e</sup> Distances calculated from  $r_i = r_{\text{ref}}(T_{1i}/T_{1\text{ref}})^{1/6}$  assuming that signal H ( $r_{\text{ref}}$ ) is purely dipolar in nature.



**Figure 2.** Magnitude  $^1\text{H}$  COSY spectrum of **1** obtained at 400 MHz (Bruker ARX-400) at 25 °C in  $\text{CD}_3\text{CN}$  solution. This spectrum was obtained with an acquisition time of 15 ms and 256 data points in the F1 dimension and 512 data points in the F2 dimension. A sine bell squared weighting function and zero-filling to 1024 data points were applied prior to Fourier transformation in both dimensions.

relative integrations of signals B (69 ppm) and C (62 ppm) of complex **2** with signals B (76 ppm) and C (61 ppm) of **1** are consistent with the assignment of these signals to the diastereotopic  $\beta$ -CH<sub>2</sub> protons of the pyridylmethyl moiety. Comparison of signal A (143 ppm) of **2** with signal A (130 ppm) of **1** is consistent with this signal resulting from the  $\beta$ -CH<sub>2</sub> protons of the phenol–methylamine linkage.

Signal A' (78 ppm) and I (-35 ppm) are the only remaining unassigned signals in the  $^1\text{H}$  NMR spectrum of **1**. The only protons in **1** not assigned are the pyridine  $\alpha$ -H protons and the  $\mu$ -hydroxo OH proton. Signal I can be assigned to the  $\mu$ -hydroxo OH proton since the addition of a small amount of  $\text{D}_2\text{O}$  causes this signal to disappear.<sup>11</sup> Moreover, the  $T_1$  value is  $\sim 1$  ms which is consistent with the short crystallographically

determined Cu–H distance (2.54 Å). From X-ray diffraction results for **1** the Cu  $d_{z^2}$  orbital, which contains the unpaired electron, is directed along the Cu–O  $\mu$ -hydroxo bond. Therefore, a spin polarization mechanism would cause the  $\mu$ -hydroxo proton to be shielded and thus shifted upfield, consistent with its assignment to signal I. Signal A' can be assigned to the pyridine  $\alpha$ -H protons by default, and this assignment is consistent with  $T_1$  values, chemical shift, and relative integrations.

Full assignment of the  $^1\text{H}$  NMR spectrum of **1** combined with the crystallographic results and  $T_1$  values allows the dominant proton relaxation pathway to be determined. Assuming a paramagnetic dipolar relaxation mechanism for antiferromagnetically coupled dinuclear Cu(II) complexes, the Cu–H distance ( $r$ ) should be proportional to  $T_1^{1/6}$  (Table 1).<sup>4</sup> Using the equation  $r_i = r_{\text{ref}}(T_{1i}/T_{1\text{ref}})^{1/6}$  where  $r_i$  is the Cu–H<sub>*i*</sub> distance,  $r_{\text{ref}}$  is the Cu–H<sub>*ref*</sub> distance,  $T_{1i}$  is the relaxation time of proton *i*, and  $T_{1\text{ref}}$  is the relaxation time of the reference proton, distances of each proton from the Cu(II) center can be estimated. If  $r_{\text{ref}}$  is taken as the arithmetic average of equivalent protons to each Cu(II) ion for the *p*-methylphenol group (7.31 Å), the remaining distances of all of the protons in **1** can be calculated (Table 1). All of the calculated Cu–H distances are within ca. 20% of the Cu–H distances derived from the X-ray structure of **1** (Table 1). These data indicate that a paramagnetic dipolar relaxation mechanism dominates in antiferromagnetically coupled trigonal bipyramidal dicopper(II) complexes.

In conclusion, isotropically shifted  $^1\text{H}$  NMR signals can be easily obtained for antiferromagnetically coupled dicopper(II) complexes, and both one- and two-dimensional  $^1\text{H}$  NMR techniques can be performed. We are currently probing the effect of the strength of the magnetic coupling on line widths and  $T_1$  values. The application of  $^1\text{H}$  NMR techniques to dicopper(II) metalloprotein active sites is also under investigation.

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**Supplementary Material Available:** A thermal ellipsoid drawing, tables detailing the X-ray data collection and refinement, bond distances, bond angles, final anisotropic thermal parameters, calculated or refined H atom coordinates, and the atomic coordinates and equivalent isotropic thermal parameters for **1**, and Figure 1, showing  $^1\text{H}$  NMR spectra of **1** and **2** (8 pages). Ordering information is given on any current masthead page.