

The Effect of Bicarbonate on Oxidation by Peroxynitrite: Implication for Its Biological Activity

Sara Goldstein* and Gidon Czapski

Department of Physical Chemistry, The Hebrew University of Jerusalem, Jerusalem 91904, Israel

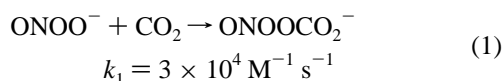
Received April 25, 1997[⊗]

The effect of bicarbonate on the oxidation rates and yields of the reaction of peroxynitrite with Ni^{II}(cyclam), ferrocyanide, and ABTS was studied using the stopped-flow technique. In the absence of bicarbonate, Ni^{II}(cyclam) is oxidized directly by peroxynitrous acid whereas ferrocyanide and ABTS are oxidized by intermediates that are formed during the self-decomposition of peroxynitrous acid. In the presence of sufficient concentrations of bicarbonate, the rate-determining step of these oxidation processes is the reaction of ONOO⁻ with CO₂. The latter reaction results in the formation of an adduct ONOOCO₂⁻, which is incapable of oxidizing directly these substrates. It is shown that about 33% of this adduct decomposes into highly oxidizing species, most probably into •NO₂ and CO₃^{•-} via the homolysis of the peroxy O–O bond. Under physiological conditions, where the most abundant species is ONOO⁻ and the concentration of CO₂ exceeds 1 mM, the half-life of peroxynitrite is reduced, and consequently its diffusion distance is shortened. Therefore, indirect oxidation by peroxynitrite through intermediates that are formed during its self-decomposition will not take place *in vivo*, whereas direct oxidation by peroxynitrite may take place provided that the rate of this reaction competes efficiently with that of peroxynitrite with CO₂. We conclude that the toxicity of peroxynitrite *in vivo* is most probably governed by ONOOCO₂⁻, which has a reactivity different from that of peroxynitrite.

Introduction

Peroxynitrous acid is an unstable and weak acid (pK_a = 6.8) that isomerizes to nitric acid (1.3 s⁻¹ at 25 °C), whereas the anion is stable.¹ Peroxynitrite (ONOOH/ONOO⁻) oxidizes a large variety of substrates including many biomolecules through complex mechanisms.^{1,2} The oxidation may take place directly through the reaction of peroxynitrite with the substrate or through intermediates formed during the decomposition of peroxynitrous acid.²

Lymar and Hurst³ have shown that ONOO⁻ reacts rapidly with CO₂ to form the unstable nitrosoperoxycarbonate (reaction 1). They have suggested that reaction 1 must be the predomi-



nant pathway for peroxynitrite disappearance in biological systems as physiological concentrations of bicarbonate in intracellular and interstitial fluids are approximately 12 and 30 mM, respectively.⁴

In the absence of other reactants, nitrate was found to be the major product of the reaction of CO₂ with peroxynitrite.⁵ The adduct may act as a damaging species either by reacting directly with a substrate or through an intermediate formed during its decomposition. Recent studies have demonstrated that ONOOCO₂⁻ participates in oxidation and nitration processes: (i) Bicarbonate significantly enhanced peroxynitrite-mediated

nitration of aromatics,^{6–8} DNA,⁹ and ethyl acetoacetate.¹⁰ (ii) It partially inhibited the oxidation by peroxynitrite of thiols,⁷ dimethyl sulfoxide,⁷ oxyhemoglobin,⁷ and (cytochrome c)²⁺,⁷ and it had hardly any effect on the oxidation of ABTS (2,2'-azinobis(3-ethyl-1,2-dihydrobenzothiazoline-6-sulfonate))⁵ and DNA.⁹ (iii) It inhibited completely peroxynitrite-mediated hydroxylation of benzoate⁷ and peroxynitrite-induced strand breakage in plasmid pBR322 DNA⁹ and thymine–propenal formation from thymidine.⁹

In this study, the effect of bicarbonate on the kinetics and yields of the oxidation of Ni^{II}(cyclam), ferrocyanide, and ABTS was investigated in line with the above observations and the earlier studies on the oxidation of these substrates by peroxynitrite.^{1,11–13}

Experimental Section

Chemicals. All chemicals used were of analytical grade and were used as received. Solutions were prepared with deionized water that was distilled and purified using a Milli-Q water purification system.

Peroxynitrite was synthesized in a quenched-flow system by the reaction of nitrite with acidified hydrogen peroxide. A syringe pump (Model SP 230IW) was used to inject 0.6 M nitrite and 0.606 M H₂O₂ in 0.7 M HClO₄ through four tangential inlets of the first mixing chamber, and the combined solutions flowed through a short connector

* To whom all correspondence should be addressed. Tel: 972-2-6586478. Fax: 972-2-6586925. E-mail: SARAG@HUJI.VMS.AC.IL.

[⊗] Abstract published in *Advance ACS Abstracts*, October 1, 1997.

- (1) Pryor, W. A.; Squadrito, G. L. *Am. J. Physiol.* **1995**, *268*, L699.
- (2) Goldstein, S.; Squadrito, G. L.; Pryor, W. A.; Czapski, G. *Free Radical Biol. Med.* **1996**, *21*, 965 and references cited therein.
- (3) Lymar, S. V.; Hurst, J. K. *J. Am. Chem. Soc.* **1995**, *117*, 8867.
- (4) Carola, R.; Harely, J. P.; Noback, C. R. *Human Anatomy & Physiology*, McGraw Hill: New York, 1990.
- (5) Uppo, R. M.; Squadrito, G. L.; Pryor, W. A. *Arch. Biochem. Biophys.* **1996**, *327*, 335.

- (6) Lymar, S. V.; Jiang, Q.; Hurst, J. K. *Biochemistry* **1996**, *35*, 7855.
- (7) Denicola, A.; Freeman, B. A.; Trujillo, M.; Radi, R. *Arch. Biochem. Biophys.* **1996**, *333*, 49.
- (8) Gow, A.; Duran, D.; Thom, S. R.; Ischiropoulos, H. *Arch. Biochem. Biophys.* **1996**, *333*, 42.
- (9) Yermilov, V.; Yoshie, Y.; Rubio, J.; Ohshima, H. *FEBS Lett.* **1996**, *399*, 67.
- (10) Uppo, R. M.; Pryor, W. A. *Biochem. Biophys. Res. Commun.* **1996**, *229*, 1996.
- (11) Goldstein, S.; Czapski, G. *Inorg. Chem.* **1995**, *34*, 4041.
- (12) Yang, G.; Candy, T. E. G.; Boaro, M.; Wilkin, H. E.; Jones, P.; Nazhat, N. B.; Saadalla-Nazhat, R. A.; Blake, D. *Free Radical Biol. Med.* **1992**, *12*, 327.
- (13) Crow, J. P.; Spruell, C.; Chen, J.; Gunn, C.; Ischiropoulos, H.; Tsai, M.; Smith, C. D.; Radi, R.; Koppenol, W. H.; Beckman, S. J. *Free Radical Biol. Med.* **1994**, *16*, 331.

to the second mixing chamber where 3 M NaOH was pushed at the same flow rate through four inlets to quench the reaction. By varying the flow rates of the solutions and/or the delay volume between the two mixing chambers, we were able to control the time elapsed between starting the reaction and quenching it.¹⁴ The yield of peroxyntirite was about 85% as determined by measuring the absorbance at 302 nm using $\epsilon = 1670 \text{ M}^{-1} \text{ cm}^{-1}$.¹⁵ These solutions contained about 1.5% nitrite, 2.5% H_2O_2 , and 13.5% nitrate.¹⁴ Ni(cyclam)(ClO₄)₂ was synthesized according to published procedures.¹⁶

Kinetic Measurements. Kinetic measurements were carried out using a Bio SX-17MV sequential stopped-flow apparatus from Applied Photophysics with a mixing time of less than 2 ms and a 1-cm-long mixing cell. The reactions of peroxyntirite with Ni^{III}(cyclam), ferrocyanide, and ABTS were followed at 360 nm ($\epsilon_{360} = 5100 \text{ M}^{-1} \text{ cm}^{-1}$),¹¹ 420 nm ($\epsilon_{420} = 1000 \text{ M}^{-1} \text{ cm}^{-1}$) and 660 nm ($\epsilon_{660} = 12000 \text{ M}^{-1} \text{ cm}^{-1}$),¹⁷ respectively, by mixing equal volumes of alkaline peroxyntirite with Ni^{II}(cyclam), ferrocyanide, or ABTS in 0.2 M phosphate or 0.2 M acetate buffers with and without bicarbonate. Bicarbonate was added as a sodium salt to the buffer solutions 2–5 min before mixing to allow the equilibration of the various carbonated species. The concentration of CO₂ in the buffer solutions was determined by measuring the pH of these solutions and assuming $\text{p}K = 6.2$ ($I = 0.25 \text{ M}$) for the reaction $\text{CO}_2 + \text{H}_2\text{O} \rightleftharpoons \text{HCO}_3^- + \text{H}^+$.¹⁸ Under all experimental conditions, the reaction of peroxyntirite with CO₂ was faster than CO₂ equilibration with HCO₃⁻.^{18,19} Therefore, the concentration of CO₂ immediately after mixing was half of that at equilibrium prior to mixing. Peroxyntirite concentration was measured before mixing by transferring a portion from the syringe into a cuvette and measuring the absorbance at 302 nm. To determine the effect of CO₂ contamination, the buffer solutions without bicarbonate were purged with argon for at least 30 min. The pH of the mixture was measured at the outlet of the flow system. Each run was repeated at least five times. The formation of the absorbance was fitted to first-order reaction kinetics, using a nonlinear regression program. The kinetic measurements were carried out at 21.7 ± 0.1 °C.

Results and Discussion

The Ni^{II}(cyclam) System. Previously,¹¹ we demonstrated that ONOOH rapidly oxidizes Ni^{II}(cyclam) to Ni^{III}(cyclam) in the presence of phosphate ions in a reaction that is first order in both reactants. The observed rate constant was determined to be $6.5 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$, and it decreased with the increase in pH as ONOO⁻ does not oxidize the metal complex.¹¹ The relatively rapid oxidation of Ni^{II}(cyclam) by peroxyntirite implies that the contamination of the buffer solutions with carbonate had no effect on the kinetics and yields in this system as the reaction of peroxyntirite with the metal complex is faster than reaction 1 under all of our previous experimental conditions ($[\text{Ni}^{\text{II}}(\text{cyclam})] = 0.5\text{--}3 \text{ mM}$, pH 5.5–8).¹¹ The stoichiometry of the reaction was found to be 1.69 ± 0.19 at pH 6.0–6.9, decreasing at higher pH's due to the decrease in $[\text{H}_2\text{PO}_4^-]$, which is essential for stabilizing the trivalent complex.¹¹

The effect of bicarbonate was studied at pH 6.3, 7, and 7.75 (0.1 M phosphate buffer) in the presence of 0.125–0.25 mM

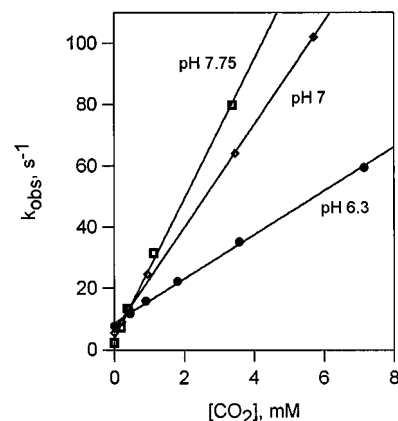


Figure 1. The observed rate constant of the formation of Ni^{III}(cyclam) as a function of $[\text{CO}_2]$ at various pH's. The final mixtures contained 0.125–0.25 mM Ni^{II}(cyclam), 20–45 μM peroxyntirite and 0.1 M phosphate buffer.

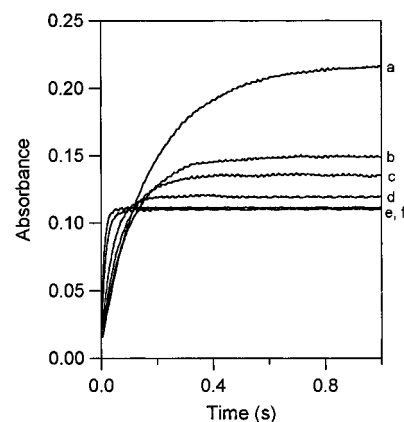


Figure 2. Kinetic traces recorded at 360 nm when 44 μM peroxyntirite in 0.01 M NaOH was mixed with 0.25 mM Ni^{II}(cyclam) in 0.2 M phosphate buffer at pH 6.8 in the absence and presence of various concentrations of bicarbonate. The pH measured at the outlet of the flow system was 7.0 ± 0.05 . The final concentrations of CO₂ were (a) none, (b) 0.208 mM, (c) 0.416 mM, (d) 0.946 mM, (e) 3.45 mM, (f) 5.71 mM (f is faster than e).

Ni^{II}(cyclam), 20–45 μM peroxyntirite, and an excess of CO₂ over peroxyntirite. The rate of the formation of Ni^{III}(cyclam) increased linearly with $[\text{CO}_2]$ (Figure 1). The observed second-order rate constants obtained from the slopes of the lines given in Figure 1 are $k = 7.19 \times 10^3$, 1.68×10^4 , and $2.28 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$ at pH 6.3, 7, and 7.75, respectively. The increase in k with the pH confirms the earlier conclusion that CO₂ reacts mainly with ONOO⁻,³ and therefore, $k_1 = k(K_a + [\text{H}^+])/K_a$. The best value obtained for k_1 from these three slopes was $(2.50 \pm 0.02) \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$ for $\text{p}K_a = 6.7$ (for $\text{p}K_a = 6.8$, $k_1 = (2.8 \pm 0.2) \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$).

The observed rate constant of the formation of Ni^{III}(cyclam) increased with bicarbonate concentration, while the yield of Ni^{III}(cyclam) decreased until it reached a plateau value (Figure 2). The yield of Ni^{III}(cyclam) at the plateau region was found to be $[\text{Ni}^{\text{III}}(\text{cyclam})]/[\text{peroxyntirite}]_0 = 0.64 \pm 0.03$ at pH 6.3 and 0.72 ± 0.04 at pH 7.0 as compared to 1.7 ± 0.2 in the absence of bicarbonate at both pH's.

The Ferrocyanide System. Previously,¹¹ we showed that the oxidation of ferrocyanide by peroxyntirite takes place via intermediates formed during the self-decomposition of peroxyntirous acid. We determined the stoichiometry of the reaction by rapidly vortexed equal volumes of peroxyntirite and ferrocyanide solutions. The stoichiometry of the reaction was about 0.8, independent of pH 5.5–9.4 and ferrocyanide concentrations

(14) Saha, A.; Goldstein, S.; Cabelli, D.; Czapski, G. *Free Radicals Biol. Med.*, in press.

(15) Halfpenny, E.; Robinson, P. L. *J. Chem. Soc. A* **1952**, 928.

(16) Bosnich, B.; Tobe, M. L.; Webb, G. A. *Inorg. Chem.* **1965**, *4*, 1109.

(17) (a) Goldstein, S.; Czapski, G. *J. Am. Chem. Soc.* **1995**, *117*, 12078.

(b) Upon irradiation of aerated solutions containing 0.5 M nitrate, 0.05 M nitrite, and 0.1–3 mM ABTS at pH 6.4 (2 mM phosphate buffer), the solvated electrons and hydroxyl radicals are converted into $\cdot\text{NO}_2$, which subsequently reacts with ABTS to form ABTS⁺. The spectrum of ABTS⁺ was recorded in the presence of the various concentrations of ABTS. The absorption at 660 nm is almost unaffected by [ABTS] ($\epsilon_{660} = 12000 \pm 1000 \text{ M}^{-1} \text{ cm}^{-1}$), whereas at [ABTS] > 0.5 mM, the maximum absorption at 414 nm is reduced and shifted toward 420 nm. In the presence of 0.1 mM ABTS, $\epsilon_{414}/\epsilon_{660} = 2.8$, and in the presence of 1–3 mM ABTS, $\epsilon_{420}/\epsilon_{660} = 2.3$ and $\epsilon_{404}/\epsilon_{660} = 2.1$.

(18) Alberty, R. A. *J. Phys. Chem.* **1995**, *99*, 11028.

(19) Kern, D. M. *J. Chem. Educ.* **1960**, *37*, 14.

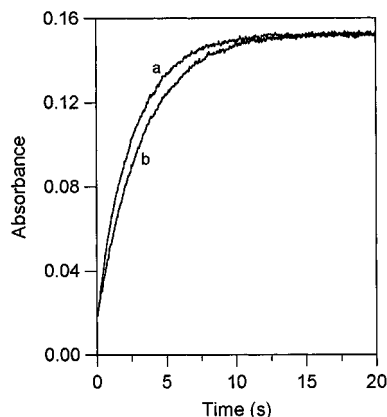


Figure 3. Kinetic traces recorded at 420 nm when 0.46 mM peroxynitrite in 0.01 M NaOH was mixed with 20 mM ferrocyanide in 0.2 M phosphate buffer: (a) air-saturated buffer solution, $k = 0.38 \text{ s}^{-1}$; (b) the buffer solution was purged with argon for 1 h, $k = 0.31 \text{ s}^{-1}$.

(10–50 mM).¹¹ Therefore, we concluded that about 60% of ONOOH isomerizes to nitric acid and about 40% is converted into a highly reactive intermediate via the indirect oxidation pathway.^{2,11} This intermediate directly oxidizes ferrocyanide, most probably through the formation of $\cdot\text{NO}_2$, which subsequently oxidizes ferrocyanide.^{2,11}

However, if the buffer solutions were contaminated with atmospheric CO_2 , which catalyzes peroxynitrite decomposition,²⁰ the measured rates and yields might be higher than in the absence of CO_2 . Therefore, these experiments were repeated and the results obtained in air-saturated buffer solutions were compared to those obtained when the buffers were purged with argon.

The oxidation of 0.3–50 mM ferrocyanide by 0.24–0.36 mM peroxynitrite was studied at pH 4.8–8.6 in the presence of 0.1 M acetate or phosphate buffers. We found that the rate of the formation of ferricyanide decreased only around neutral pH when the buffer solutions were purged with argon, while the oxidation yields were unaffected (Figure 3). Around neutral pH, reaction 1 competes with the self-decomposition of peroxynitrite due to contamination of the solutions with atmospheric CO_2 . In alkaline solutions the concentration of CO_2 is very low ($\text{p}K = 6.2$), and can be neglected. In acidic solutions reaction 1 will not compete with the rate of self-decomposition of peroxynitrite because the concentration of ONOO^- is markedly reduced ($\text{p}K_a = 6.8$).

The oxidation rates and yields were reported previously to be independent of ferrocyanide concentrations.¹¹ In this study we found that only below 10 mM ferrocyanide were the yields independent of ferrocyanide concentration. Above 10 mM, there was a slight increase in the yields and rates, suggesting that, in the presence of relatively high concentrations of ferrocyanide, there is mixed direct and indirect oxidation of ferrocyanide by peroxynitrite.² The second-order rate constant of the reaction of peroxynitrite with ferrocyanide was determined to be 5.3, 2.5, and $1.5 \text{ M}^{-1} \text{ s}^{-1}$ at pH 5.2, 6.3, and 7, respectively, suggesting that peroxynitrous acid directly oxidizes ferrocyanide. The direct oxidation of ferrocyanide by peroxynitrite was not observed previously.¹¹ In our earlier study we used the Hi-Tech Scientific Instrument model PQ/SF-53 stopped-flow apparatus, which yielded less accurate results,¹¹ and the increase in the rates and yields was at that time within the limits of the experimental error.

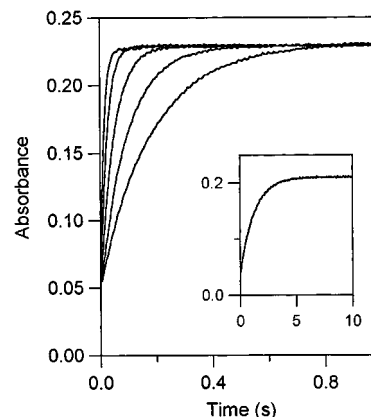


Figure 4. Kinetic traces recorded at 420 nm when 0.57 mM peroxynitrite in 0.01 M NaOH was mixed with 20 mM ferrocyanide in 0.2 M phosphate buffer at pH 6 in the absence and presence of various concentrations of bicarbonate. The pH measured at the outlet of the flow system was 6.3 ± 0.1 . The final concentrations of CO_2 were 0.81, 1.62, 3.24, 6.48, and 12.96 mM.

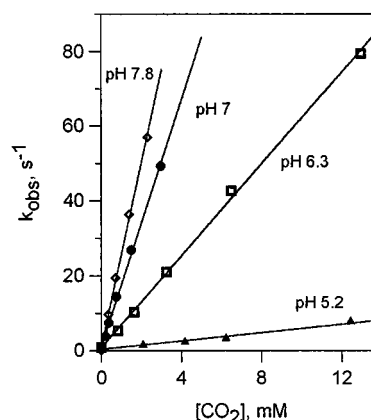


Figure 5. The observed rate constant of the formation of ferricyanide as a function of $[\text{CO}_2]$ at various pH's. The final mixtures contained 10 mM ferrocyanide, 0.27–0.32 mM peroxynitrite, and 0.1 M phosphate or acetate buffers.

The effect of bicarbonate on the oxidation rates and yields was studied of 1–50 mM ferrocyanide at pH 5.2–8.6. In the presence of an excess of CO_2 over peroxynitrite, the rate of the formation of ferricyanide increased with the increase in $[\text{CO}_2]$, while the oxidation yields were unaffected (Figure 4). The observed rate constants for the formation of ferricyanide were linearly dependent on $[\text{CO}_2]$ and highly dependent on pH (Figure 5). As in the case of $\text{Ni}^{\text{II}}(\text{cyclam})$, the increase in the slope of the lines in Figure 5 with the pH is due to the reaction of CO_2 with ONOO^- .³ From the slopes of these lines the best value of k_1 was determined to be $(2.9 \pm 0.1) \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$ for $\text{p}K_a = 6.9$ (for $\text{p}K_a = 6.8$, $k_1 = (2.6 \pm 0.3) \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$).

The yield of ferricyanide was determined to be $[\text{Fe}(\text{CN})_6^{3-}]/[\text{peroxynitrite}]_0 = 0.66 \pm 0.06$, independent of pH, ferrocyanide, and bicarbonate concentrations (Table 1).

The ABTS System. The oxidation of ABTS by peroxynitrite takes place through a complex mechanism, where ONOOH indirectly oxidizes ABTS to ABTS^+ through intermediates which are formed during its decomposition.^{1,12,13} However, the yield of ABTS^+ decreased with the increase in pH,^{1,12,13} whereas in the case of ferrocyanide it is independent of pH (Table 1). The yield of ABTS^+ was determined to be $[\text{ABTS}^+]/[\text{peroxynitrite}]_0 = 0.63,^{12} 0.45,^{11}$ and 0.24⁵ in the presence of $[\text{ABTS}] \leq 4 \text{ mM}$ and at $\text{pH} \leq 7$.

We found that, under limiting concentrations of peroxynitrite, the yield of ABTS^+ was 0.42 ± 0.03 and 0.51 ± 0.03 at pH

(20) Pryor, W. A.; Lemerrier, J.-N.; Zhang, H.; Uppo, R. M.; Squadrito, G. L. *Free Radical Biol. Med.* **1997**, *23*, 331.

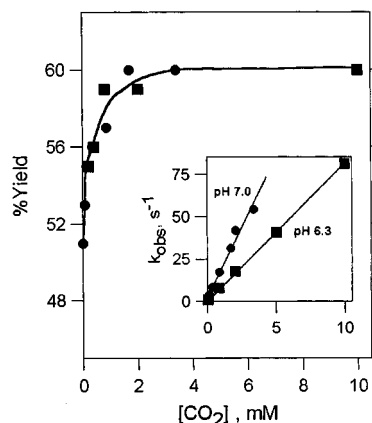


Figure 6. The yield of ABTS^+ ($[\text{ABTS}^+]/[\text{peroxynitrite}]_0$) as a function of $[\text{CO}_2]$ at pH 6.3 (■) and 7.0 (●). The final mixtures contained 2 mM ABTS, 72–75 μM peroxynitrite and 0.1 M phosphate buffer. Inset: The observed rate constant of the formation of ABTS^+ as a function of $[\text{CO}_2]$ under the same conditions as stated above.

Table 1. The Yield of Ferricyanide Obtained through the Reaction of Peroxynitrite with Ferrocyanide under Various Conditions

$[\text{Fe}(\text{CN})_6^{4-}]$, mM	$[\text{peroxynitrite}]$, mM	pH	$[\text{CO}_2]$, mM	$[\text{Fe}(\text{CN})_6^{3-}]/[\text{peroxynitrite}]_0$
5	0.30	5.2	air	0.68
10	0.30	5.2	12.4	0.68
10	0.30	5.2	2.1	0.69
25	0.24	5.5	10.3	0.70
10	0.24	5.5	10.3	0.68
1	0.24	5.5	10.3	0.69
50	0.24	6.3	2.9	0.69
10	0.29	6.3	12.9	0.64
10	0.24	6.3	2.9	0.66
1	0.24	6.3	2.9	0.66
5	0.23	7	air	0.63
50	0.29	7	1.58	0.60
10	0.29	7	1.48	0.63
10	0.30	7	0.37	0.66
1	0.29	7	1.58	0.61
5	0.29	7.8	purged	0.66
10	0.30	7.8	2.3	0.59
10	0.30	7.8	0.69	0.66
20	0.36	8.6	1.09	0.62

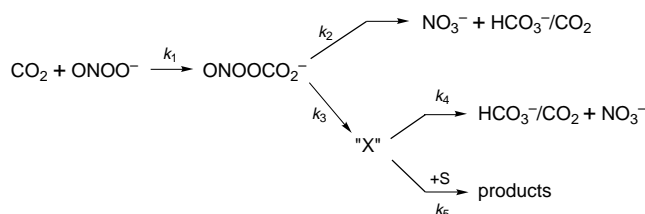
6.2 and 7.0, respectively, independent of 0.1–2 mM ABTS. The oxidation yields and rates increased with the increase in $[\text{ABTS}]$, in agreement with previous observations,¹² suggesting that under these conditions a mixed direct and indirect oxidation takes place.

The effect of bicarbonate on the oxidation yields and rates was studied at pH 6.3 and 7.0 in the presence of 0.05–10 mM ABTS. When $[\text{CO}_2] > [\text{peroxynitrite}]$, the rate of the formation of ABTS^+ was linearly dependent on $[\text{CO}_2]$ and pH dependent (Figure 6). From the slope of the lines in Figure 6 we calculated $k_1 = (2.7 \pm 0.2 \times 10^4 \text{ M}^{-1} \text{ s}^{-1})$ using $\text{p}K_a = 6.8$ for peroxynitrite. The yield of ABTS^+ increased with $[\text{CO}_2]$ reaching a plateau value where $[\text{ABTS}^+]/[\text{peroxynitrite}]_0 = 0.60 \pm 0.03$ (Figure 6). The maximum yield was independent of pH and 0.05–10 mM ABTS.

Uppo et al.⁵ reported that the yield of ABTS^+ increased from 0.24 to 0.27 upon the addition of 20 mM bicarbonate to 0.005–0.1 mM peroxynitrite and 2 mM ABTS at pH 7.0. However, they measured ABTS^+ spectrophotometrically at 404 nm using $\epsilon_{404} = 36\,800 \text{ M}^{-1} \text{ cm}^{-1}$.⁵ We have already noted that the spectrum of ABTS^+ depends on $[\text{ABTS}]$.¹⁷ The absorption at 660 nm is almost unaffected by $[\text{ABTS}]$ ($\epsilon_{660} = 12\,000 \text{ M}^{-1} \text{ cm}^{-1}$), whereas the maximum absorption at 414 nm is reduced and shifted toward 420 nm in the presence of $[\text{ABTS}] > 0.5 \text{ mM}$.¹⁷ Furthermore, Uppo et al.⁵ allowed peroxynitrite to react

with ABTS for 2 min, ignoring the slow decay of ABTS^+ .^{1,12} Nevertheless, the yield of ABTS^+ in their system increased by about 13% in the presence of bicarbonate, which correlates well with our observations at pH 7.0.

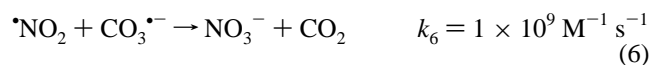
Mechanism. Under the conditions where reaction 1 competes efficiently with the oxidation of $\text{Ni}^{\text{II}}(\text{cyclam})$, ferrocyanide, and ABTS by peroxynitrite, the yields of the various products were determined to be $66 \pm 6\%$, independent of pH, $[\text{CO}_2]$, and substrate concentrations. Since two molecules of these substrates are consumed by one molecule of peroxynitrite, only $33 \pm 3\%$ of added peroxynitrite is available for these oxidation processes, and we define the oxidation yield as 33%. Our results are in agreement with those reported in the case of glutathione (GSH),²¹ nitrate,²² and $\text{Mo}(\text{CN})_8^{4-}$.²² Zhang et al.²¹ showed that the amount of GSH consumed in the presence of bicarbonate by 0.4 mM peroxynitrite was 0.23 mM. Since two GSH molecules are consumed upon reaction with peroxynitrite, we calculated that the amount of peroxynitrite available for the reaction is 29%. Hurst and Lyman²² reported that 33–34% of added peroxynitrite in the presence of bicarbonate is available for the reaction with nitrate and $\text{Mo}(\text{CN})_8^{4-}$. These results suggest that the oxidation by peroxynitrite in the presence of bicarbonate takes place through the following mechanism, which has already been suggested by Lyman et al.⁵ for tyrosine:



The oxidation yield in the presence of sufficient concentrations of the substrates ($k_5[\text{S}] \gg k_4$) is $33 \pm 3\%$, and therefore $k_2/k_3 = 2.0 \pm 0.3$.

Lyman et al.⁶ have also suggested that reaction 1 may form in parallel ONOOCO_2^- and "X". These two mechanisms are kinetically indistinguishable, and therefore neither is excluded by the data. However, it is difficult to envisage that "X" is formed directly from the reaction of ONOO^- and CO_2 , especially if "X" is the couple $\text{CO}_3^{\bullet-}$ and $\bullet\text{NO}_2$ (see below).

At present, our data are not sufficient for identification of "X". It might be an isomer of ONOOCO_2^- , which hydrolyzes slowly and hence is capable of oxidizing the substrate. Uppo et al.⁵ have suggested the species $\text{O}_2\text{NOCO}_2^-$, but theoretical calculations in the gas phase imply that it is unstable as it undergoes spontaneous homolysis to give $\bullet\text{NO}_2$ and $\text{CO}_3^{\bullet-}$.²³ "X" may represent the couple $\bullet\text{NO}_2$ and $\text{CO}_3^{\bullet-}$, which are formed via the homolytic cleavage of the peroxy O–O bond, as both radicals are relatively long-lived and are capable of oxidizing many compounds.²⁴ In the absence of any substrate they will yield nitrate and CO_2 via reaction 6.²⁵



- Zhang, H.; Squdrito, G. L.; Uppo, R. M.; Lemercier, J.-N.; Cueto, R.; Pryor, W. A. *Arch. Biochem. Biophys.* **1997**, *339*, 183.
- Hurst J. K.; Lyman, S. V. *Book of Abstracts*; The First International Conference on the Chemistry and Biology of Peroxynitrite; Ascona, Switzerland, 1997; p 26.
- Houk, K. N.; Condroski, K. R.; Pryor, W. A. *J. Am. Chem. Soc.* **1996**, *118*, 13002.
- Ross, A. B.; Mallard, W. G.; Helman, W. P.; Buxton, J. V.; Huie, R. E.; Neta, P. *NIST Standard References Database 40, Version 2.0* 1994.

The identity of "X" may be the couple NO_2^+ and CO_3^{2-} , which are formed via the heterolytic cleavage of the peroxo O–O bond. However, the half-life of NO_2^+ in water has been determined to be 1.4 ns,²⁶ which is too short to be scavenged by the relatively low concentrations of the substrates, thus eliminating this species as the oxidizing entity.

It has been shown that nitrate is the major product of the reaction of peroxynitrite with CO_2 in the absence of any other reactants.⁵ Therefore, we have to assume that the decomposition of ONOOCO_2^- via reaction 2 yields nitrate. However, the hydrolysis of ONOOCO_2^- to nitrate and bicarbonate seems unlikely since it will yield a peroxidic species (ONOO^- or $^- \text{OOCO}_2$) as one of the immediate products.²⁷ In addition, if water can attack the adduct, any other good nucleophile should also be able to capture all ONOOCO_2^- , which so far has not been reported.²⁷ Therefore, we suggest that the decomposition of ONOOCO_2^- to form nitrate as a final product (reaction 2) takes place via the formation of an intermediate. This intermediate might be an isomer of ONOOCO_2^- , e.g., $\text{O}_2\text{NOCO}_2^-$, which hydrolyzes to form nitrate and bicarbonate or decomposes to NO_3^- and CO_2 .

Conclusions

The reaction of peroxynitrite with CO_2 yields ONOOCO_2^- , which under our experimental conditions is incapable of directly oxidizing $\text{Ni}^{\text{II}}(\text{cyclam})$, ferrocyanide, and ABTS. We have shown that only about 33% of this adduct can oxidize the various

substrates. Note that this ratio is similar to that determined earlier for the direct isomerization of peroxynitrite to nitrate and its conversion into a highly oxidizing intermediate.^{2,11} However, though these ratios are similar, they are related to different reactions. The reactivity of peroxynitrite in the presence of bicarbonate differs from that of peroxynitrite as it is governed by the formation of ONOOCO_2^- , of which $33 \pm 3\%$, at most, can participate in the oxidation processes.

The toxicity of peroxynitrite is usually attributed to its relatively long half-life which enables it to diffuse away from the site of its formation. However, under physiological conditions (pH 7.4, $\text{p}K_a(\text{ONOOH}) = 6.8$, $[\text{HCO}_3^-] = 12\text{--}30$ mM, 37°C), the most abundant species is ONOO^- which reacts with CO_2 with $k_1 = 5.8 \times 10^4 \text{ M}^{-1} \text{ s}^{-1}$.⁷ As the concentrations of CO_2 under these conditions are 0.7–1.8 mM, the half-life of peroxynitrite in the intracellular and interstitial fluids is reduced from about 1 s to less than 20 ms, and consequently its average diffusion distance is shortened to less than $6.3 \mu\text{m}$ ($L = (2Dt)^{0.5}$, $D \sim 10^{-5} \text{ cm}^2 \text{ s}^{-1}$). We conclude that indirect oxidation by peroxynitrite will not take place *in vivo* in the presence of bicarbonate. The direct oxidation by peroxynitrite will take place provided that this reaction competes efficiently with reaction 1.

In conclusion, the toxicity of peroxynitrite *in vivo* is most probably governed by ONOOCO_2^- , which has a reactivity different from that of peroxynitrite.

Acknowledgment. This research was supported by Grant 4129 from The Council For Tobacco Research and by The Israel Science Foundation. We thank Prof. D. Meyerstein for his generous gift of $\text{Ni}^{\text{II}}(\text{cyclam})(\text{ClO}_4)_2$.

IC970471V

(25) Lillie, J.; Hanrahan, R. J.; Henglein, A. *Radiat. Phys. Chem.* **1978**, *11*, 225.

(26) Moodie, R. B.; Schofield, K. Taylor, P. G.; *J. Chem. Soc., Perkin Trans. 2* **1979**, 133.

(27) We thank Drs. W. A. Pryor and G. L. Squadrito for this comment as we originally suggested reaction 2 as the hydrolysis of ONOOCO_2^- .