

A Sterically Hindered N,N,O Tripod Ligand and Its Zinc Complex Chemistry

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The new ligand bis(2-picolyl)(2-hydroxy-3,5-di-*tert*-butylbenzyl)amine (HL) was prepared from bis(2-picolyl)amine and 2,4-di-*tert*-butyl-6-(chloromethyl)phenol. It acts as a tetradentate N,N,O tripod ligand ensuring 5-fold coordination in all its zinc complexes $L \cdot Zn-X$. The central complex of the series was $[L \cdot Zn(OH_2)]ClO_4$ (**1**) obtained from zinc perchlorate. Together with the more labile complex $L \cdot Zn-C_2H_5$ (**2**), obtained from diethyl zinc, it was used as a starting material for ligand substitutions. In the presence of bases, **1** was converted to $L \cdot Zn-OH$ (**3**), $[L \cdot Zn(py)]ClO_4$ (**4**), and $[(L \cdot Zn)_3(\mu_3-CO_3)]ClO_4$ (**5**). Metathetical reactions produced the neutral complexes $L \cdot Zn-X$ with $X = Br$ (**6**), OAc (**7**), OC_6H_5 (**8**), SC_6H_5 (**9**), $OP(O)(OPh)_2$ (**10**), *p*-nitrophenolate (**11**), 1-methyluracilate (**12**), *o*-formylphenolate (**13**), and *o*-hydroxymethylphenolate (**14**). Structure determinations of **1**, **5**, **7**, **10**, **11**, **13**, and **14** confirmed the strictly monodentate attachment of all units X in $L \cdot Zn-X$. The hydrolytic cleavage of tris(*p*-nitrophenyl) phosphate by **1** was investigated preparatively and kinetically. $L \cdot Zn-OH$ was found to be the hydrolytically active nucleophile. The second-order rate constant for the cleavage reaction was found to be slightly lower than the values for related systems, reflecting the steric hindrance in the *tert*-butyl-substituted ligand **L**.

The zinc model complex chemistry with respect to catalytic or bioinorganic systems has reached a considerable level of sophistication today. Encapsulating ligands have become the ligands of choice for mimicking the donor environment of the metal and to restrain the number of available “active” coordination sites to one or two. Among these the tripodal ligands are the most popular ones. The majority of research groups active in a preparative zinc model complex chemistry have put efforts in the design or use of tripodal ligands, as evidenced by recent publications.^{1–18} We have contributed to this mostly with

pyrazolylborate ligands¹⁹ but also with tripods possessing central carbon,²⁰ nitrogen,²¹ phosphorus, or arsenic atoms²² and cyclohexane rings.²³

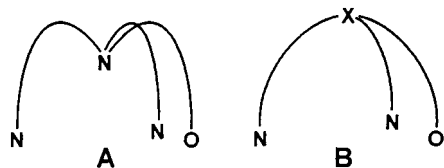
While the well-established tripod ligands possess three identical arms (more correctly feet), sophistication has been achieved by making them unsymmetrical or heteroleptic. Induced by Nature’s predominating mixed N/S or N/O donor sets (provided by cysteinate, histidine, tyrosinate, glutamate, and aspartate), mixed N/S or N/O tripod ligands have become popular. Recent examples of N_2S or NS_2 tripods with central carbon,²⁴ boron,^{5,25,26} or nitrogen atoms¹⁸ underline this, including examples from our research group.²⁷

This paper deals with tripods containing a N_2O donor set. The motivation for their design and use comes from the fact that important hydrolytic enzymes such as carboxypeptidase A,²⁸ thermolysine,²⁹ alkaline phosphatase,³⁰ L-fucose 1-phosphate

- (1) Kimblin, C.; Bridgewater, B. M.; Churchill, D. G.; Parkin, G. *J. Chem. Soc., Chem. Commun.* **1999**, 2301.
- (2) Adams, H.; Bailey, N. A.; Fenton, D. E.; He, Q.-Y. *J. Chem. Soc., Dalton Trans.* **1997**, 1533.
- (3) Hammes, B. S.; Carrano, C. J. *Inorg. Chem.* **1999**, *38*, 4593.
- (4) Cronin, L.; Greener, B.; Foxon, S. P.; Heath, S. L.; Walton, P. H. *Inorg. Chem.* **1997**, *36*, 2594.
- (5) Chiou, S.-J.; Innocent J.; Riordan, C. G. *Inorg. Chem.* **2000**, *39*, 4347.
- (6) Mann, K. L. V.; Jeffery, J. C.; McCleverty, J. A.; Ward, M. D. *J. Chem. Soc., Dalton Trans.* **1998**, 3029.
- (7) Kläui, W.; Berghahn, M.; Rheinwald, G.; Lang, H. *Angew. Chem.* **2000**, *112*, 2590; *Angew. Chem., Int. Ed. Engl.* **2000**, *39*, 2464.
- (8) Reglinski, J.; Garner, M.; Cassidy, I. D.; Slavin, P. A.; Spicer, M. D.; Armstrong, D. R. *J. Chem. Soc., Dalton Trans.* **1999**, 2119.
- (9) Darensbourg, D. J.; Holtcamp, M. W.; Longridge, E. M.; Khandelwal, B.; Klausmeyer, K. K.; Reibenspies, J. H. *J. Am. Chem. Soc.* **1995**, *117*, 318.
- (10) Weber, M.; Kuppert, D.; Hegetschweiler, K.; Gramlich, V. *Inorg. Chem.* **1999**, *38*, 859.
- (11) Murthy, N. N.; Karlin, K. D. *J. Chem. Soc., Chem. Commun.* **1993**, 1236.
- (12) Hihichi, S.; Tanaka, M.; Moro-oka, Y.; Kitajima, N. *J. Chem. Soc., Chem. Commun.* **1992**, 814.
- (13) Le Cloux, D. D.; Tolman, W. B. *J. Am. Chem. Soc.* **1993**, *115*, 1153.
- (14) Itho, T.; Fujii, Y.; Tada, T.; Yoshikawa, Y.; Hisada, H. *Bull. Chem. Soc. Jpn.* **1996**, *69*, 1265.
- (15) Meyer, F.; Rutsch, P. *J. Chem. Soc., Chem. Commun.* **1998**, 1037.
- (16) Xu, X.-D.; Lajmi, A. R.; Canary, J. W. *J. Chem. Soc., Chem. Commun.* **1998**, 2701.
- (17) Ray, M.; Hammes, B. S.; Yap, G. P. A.; Rheingold, A. L.; Borovik, A. S. *Inorg. Chem.* **1998**, *37*, 1527.

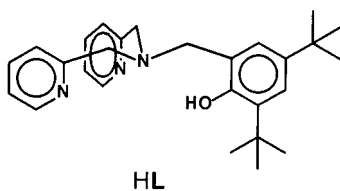
- (18) Berreau, L. M.; Allred, R. A.; Makowska-Grzyska, M. M.; Arif, A. M. *J. Chem. Soc., Chem. Commun.* **2000**, 1423.
- (19) Vahrenkamp, H. *Acc. Chem. Res.* **1999**, *32*, 589.
- (20) Brandt, W.; Wirbser, J.; Powell, A. K.; Vahrenkamp, H. *Z. Naturforsch.* **1991**, *46b*, 440. Titze, C.; Hermann, J.; Vahrenkamp, H. *Chem. Ber.* **1995**, *128*, 1095.
- (21) Gregorzik, R.; Hartmann, U.; Vahrenkamp, H. *Chem. Ber.* **1994**, *127*, 2117. Hartmann, U.; Gregorzik, R.; Vahrenkamp, H. *Chem. Ber.* **1994**, *127*, 2123. Burth, R.; Vahrenkamp, H. *Z. Anorg. Allg. Chem.* **1998**, *624*, 381.
- (22) Gregorzik, R.; Wirbser, J.; Vahrenkamp, H. *Chem. Ber.* **1992**, *125*, 1575.
- (23) Brand, U.; Vahrenkamp, H. *Inorg. Chim. Acta* **1992**, *198–200*, 663.
- (24) Hammes, B. S.; Carrano, C. J. *J. Chem. Soc., Chem. Commun.* **2000**, 1635.
- (25) Kimblin, C.; Hascall, T.; Parkin, G. *Inorg. Chem.* **1997**, *36*, 5680.
- (26) Chiou, S.-J.; Ge, P.-H.; Riordan, C. G.; Liable-Sands, L. M.; Rheingold, A. L. *J. Chem. Soc., Chem. Commun.* **1999**, 159.
- (27) Burth, R.; Stange, A.; Schäfer, M.; Vahrenkamp, H. *Eur. J. Inorg. Chem.* **1998**, 1759. Alsfasser, R.; Vahrenkamp, H. *Inorg. Chim. Acta* **1993**, *209*, 19.
- (28) Rees, D. C.; Lewis, M.; Lipscomb, W. N. *J. Mol. Biol.* **1983**, *168*, 367.
- (29) Holmes, M. A.; Matthews, B. W. *J. Mol. Biol.* **1982**, *160*, 623.

aldolase,³¹ or the peptidase astacin³² contain the catalytically active zinc ion in a N_xO_y donor environment. Major contributions to the ligand design and zinc complex chemistry of N,N,O tripods based on a central N atom (type A) were made by Fenton.^{33,34} But also Carrano,³⁵ Parkin,³⁶ and others^{37,38} published new ways of obtaining and applying such tripods, including those with a central noncoordinating atom X (type B).



We made our entry into the field of N,N,O tripods with the reaction chemistry of dipicolylglycinate complexes of zinc³⁹ and a report on the first chiral N,N,O tripod,⁴⁰ followed by some studies of the zinc complex chemistry of dipicolylalanate and dipicolyl-2-oxybenzylate.⁴¹ Like our competitors^{3,14,33} we could accumulate some evidence that the reactive species in water-containing solutions of these complexes is the monoqua cation [(ligand)Zn(OH₂)]⁺. Yet so far neither of us had been able to prepare or isolate this elusive species or its even more interesting deprotonated form (ligand)Zn–OH.

We therefore resorted to a technique that had been so successful for us in (pyrazolylborato)zinc chemistry,¹⁹ the introduction of voluminous substituents on the ligand L, which favor lower coordination numbers of zinc and induce a hydrophobic environment of the metal. Both effects should enhance the stability of a L·Zn(OH₂) or L·Zn–OH complex. This paper reports the synthesis of the so designed ligand bis(2-picoyl)(2-hydroxy-3,5-di-*tert*-butylbenzyl)amine (HL) and the reaction chemistry of its zinc aqua complex [L·Zn(H₂O)]⁺. The latter is the first zinc monoqua complex of a N,N,O tripod. It has allowed to prove that the hydrolytically active species in its reactions is actually the hydroxide L·Zn–OH.



Results and Discussion

Ligand HL. The ligand synthesis was straightforward, combining HL from the easily available components bis(2-

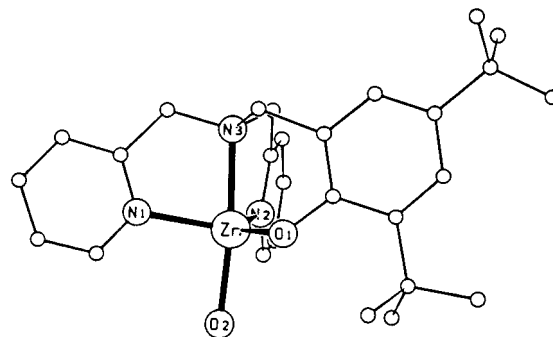


Figure 1. Molecular structure of **1**. (One cationic complex is shown; in one of them the water ligand is hydrogen-bonded to two water molecules, and in the other it is bonded to one water and one THF molecule). Important bond lengths (Å) and angles (deg) for both independent cations: Zn–O1 1.945(6)/1.924(6), Zn–N1 2.099(7)/2.073(8), Zn–N2 2.061(7)/2.059(8), Zn–N3 2.178(7)/2.219(7), Zn–O2 2.059(6)/2.079(7); O2–Zn–O1 99.8(3)/94.0(3), O2–Zn–N1 91.8(3)/95.4(3), O2–Zn–N2 98.3(3)/97.6(3), O2–Zn–N3 165.0(3)/173.2(3).

picoyl)amine⁴² and 2,4-di-*tert*-butyl-6-(chloromethyl)phenol.⁴⁰ Chromatographic purification yielded 70% HL as a yellowish solid which is soluble in organic solvents but insoluble in water. HL is easily identified by its NMR spectrum and its characteristic pyridine IR band at 1591 cm⁻¹.

Starting Complexes. It had been shown that the simplest way of generating a Zn–OH₂ or Zn–OH function in the pocket of an encapsulating tripodal ligand is the reaction between the ligand and the hydrated zinc salt of a noncoordinating anion. This had worked for us for the tris(pyrazolyl)borate–Zn–OH and tris(benzimidazolymethyl)amine–Zn–OH₂ complexes^{19,21} but failed for others and ourselves when applied to ligands analogous to HL.^{3,14,33,34,39–41} It worked now for HL due to the presence of the *tert*-butyl substituents. Complex **1** resulted from HL, KOH, and Zn(ClO₄)₂·6H₂O in methanol. Its characteristic spectroscopic features are a proton resonance for the aqua ligand at 3.34 ppm in CDCl₃ and the typically shifted pyridine IR band at 1610 cm⁻¹.



While **1** served the purpose of functionalization by replacement of the aqua ligand in polar media (see below), another starting complex for derivatizations by proteolytic cleavage in nonpolar media was found in **2**. As experienced for a similar system before,⁴⁰ **2** was obtained from HL and diethylzinc in hydrocarbon solvents. Its lability and its high solubility in nonpolar media prevented its purification, but it was easily identified by its proton resonances for the Zn–C₂H₅ unit. For derivatizations **2** was prepared and used in situ.

Proof for the existence and structure of **1** was obtained by a X-ray analysis; see Figure 1. **1** crystallizes with two formula units/asymmetric unit which differ in the hydrogen-bonding patterns of the ligated water molecules but not in the gross molecular features. The coordination geometry is roughly trigonal-bipyramidal with a typical distortion toward a pseudotetrahedral ligation (long Zn–N bonds to the apical nitrogen atom and O(axial)–Zn–O,N(equatorial) angles above 90°).

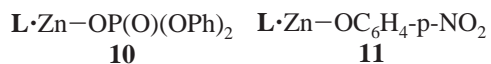
There are some trigonal-bipyramidal Zn–OH₂ complexes

- (30) Kim, E. E.; Wyckof, H. W. *J. Mol. Biol.* **1991**, *218*, 449.
 (31) Dreyer, M. K.; Schulz, G. E. *J. Mol. Biol.* **1993**, *231*, 549.
 (32) Bode, W.; Goumis-Rüth, F.; Huber, R.; Zwilling, R.; Stöcker, W. *Nature* **1992**, *358*, 164.
 (33) Adams, H.; Bailey, N. A.; Fenton, D. E.; He, Q.-Y. *J. Chem. Soc., Dalton Trans.* **1996**, 2857.
 (34) Rodriguez de Barbarin, C. O.; Bailey, N. A.; Fenton, D. E.; He, Q.-Y. *J. Chem. Soc., Dalton Trans.* **1997**, 161.
 (35) Higgs, T. C.; Spartalian, K.; O'Connor, C. J.; Matzanke, B. F.; Carrano, C. J. *Inorg. Chem.* **1998**, *37*, 2263.
 (36) Ghosh, P.; Parkin, G. J. *Chem. Soc., Dalton Trans.* **1998**, 2281.
 (37) Otero, A.; Fernandez-Baeza, J.; Tejada, J.; Antiñolo, A.; Carillo-Hermosilla, F.; Diez-Barra, E.; Lara-Sanchez, A.; Fernandez-Lopez, M. *J. Chem. Soc., Dalton Trans.* **2000**, 2367.
 (38) Mao, Z.-W.; Yu, K.-B.; Chen, D.; Han, S.-Y.; Sui, Y.-X.; Tang, W.-X. *Inorg. Chem.* **1993**, *32*, 3104.
 (39) Abufarag, A.; Vahrenkamp, H. *Inorg. Chem.* **1995**, *34*, 2207.
 (40) Abufarag, A.; Vahrenkamp, H. *Inorg. Chem.* **1995**, *34*, 3279.
 (41) Trösch, A.; Vahrenkamp, H. *Eur. J. Inorg. Chem.* **1998**, 827.

- (42) Romary, J. K.; Zachariassen, R. D.; Bargar, J. D.; Schiesser, H. *J. Chem. Soc.* **1968**, 2884.

scopic features. The coligands X in these species $L \cdot Zn-X$ are monodentate. Only the acetate ligand in **7** has the alternative to be bidentate. Therefore, **7** was chosen for a structure determination which is documented in the Supporting Information. It confirmed the strictly monodentate acetate attachment ($Zn-O-C = 138^\circ$). Thus the four simple complexes **6-9** can be assigned a structure like **1** with the coligand X on an apical position of a distorted trigonal-bipyramidal $ZnON_3X$ ligand set.

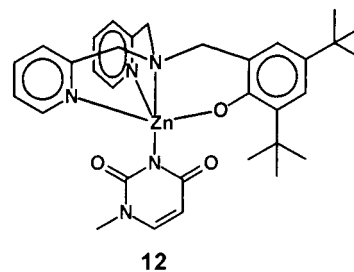
The second group of coligands chosen contained diphenyl phosphate and *p*-nitrophenolate which are substrate and product analogues of zinc enzyme catalyzed phosphate ester hydrolyses. At the same time they were meant to provide a structural background for the mechanistic investigation of the phosphate ester cleavage by **3** described below. Complexes **10** and **11** were obtained from **1** by addition of base and the reagent (diphenylphosphoric acid or *p*-nitrophenol). Both reaction courses (deprotonation of **1** before addition of the reagent or deprotonation of the reagent before addition to **1**) are feasible. Alternatively, **10** and **11** are accessible from the ethyl complex **2** and the reagent.



Solubility and spectroscopic properties confirm the mononuclear and molecular nature of **10** and **11**. One significant spectroscopic feature of **11** is the position of the UV absorption band due to the *p*-nitrophenyl group ($\lambda_{max} = 385$ nm) which allows one to distinguish **11** from free *p*-nitrophenolate ($\lambda_{max} = 400$ nm), which is important for the mechanistic investigation described below. Both **10** and **11** were subjected to crystal structure determinations which are documented in the Supporting Information. They confirmed the distorted trigonal-bipyramidal ZnN_3O_2 coordination with axial angles of 167° in **10** and 160° in **11** and with $Zn-O(\text{substrate})$ bond lengths of 1.99 \AA in **10** and 1.98 \AA in **11**. The other molecular details of **10** and **11** correspond closely to those reported by us³⁹⁻⁴¹ for similar phosphate complexes and by Fenton⁴⁸ and us⁴⁰ for related phenolate complexes, and the ligand arrangements in **11** and **13/14** (see below) are also very similar.

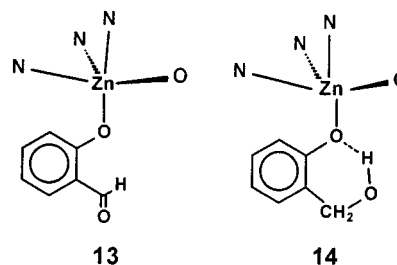
The third group of coligands contained those which are potentially bidentate and relevant in the context of nonhydrolytic zinc enzymes. We had observed that nucleobases, which are synthesized and metabolized inter alia by zinc enzymes, are bound to zinc preferably via their deprotonated NH functions and can use their additional donor functions for chelation.^{49,50} We tested this here for 1-methyluracil as the simplest analogue of uridine and the uridine nucleotides. Its reaction with **2** yielded the uracilate **12**. The spectroscopic data for **12** support the formulation given without coordination of the $C=O$ functions to zinc. The main indicator is the $\nu(CO)$ absorption in the IR at 1644 cm^{-1} which exactly corresponds to that of the pyrazolylborate complex $Tp^{Cum,Me}Zn-1\text{-methyluracilate}$ which we have structurally characterized.⁵¹

o-Formylphenolate (the anion of salicylic aldehyde) and *o*-hydroxymethylphenolate were applied with the clear expectation that they might act as bidentate ligands. Their formyl and hydroxymethyl groups represent the oxidized and reduced forms of the substrates (aldehyde/alcohol) which are interconverted



by the zinc-containing alcoholdehydrogenase enzymes. We had observed before^{52,53} that the enzyme-substrate interactions of these enzymes can be modeled by zinc complexes in which the aldehydic and alcoholic functions are attached to zinc in a chelating fashion. As the second donor of the chelating alcohol or aldehyde ligands we had used pyridine nitrogen⁵² or phenolate oxygen,⁵³ and even in the pocket within encapsulating pyrazolylborate ligands could the increase of the coordination number of zinc be realized. With this in mind, complexes $L \cdot Zn-OC_6H_4-o-CHO$ (**13**) and $L \cdot Zn-O-C_6H_4-o-CH_2OH$ (**14**) were synthesized from **1** and the potassium salts of *o*-formylphenol and *o*-(hydroxymethyl)phenol.

The relevant spectroscopic data for **13** and **14** [$\nu(CO\text{-aldehyde})$ and $\delta(CHO)$ as well as $\delta(CH_2OH)$] do not differ significantly from those of the free substrates and hence speak against a zinc coordination by the CHO or CH_2OH groups. Structure determinations confirmed this conclusion. Both **13** and **14** (details in the Supporting Information) have a zinc coordination which is virtually identical to that of **11** and related complexes.^{40,48} The aldehyde donor in **13** is turned away from the zinc ion. The alcoholic function in **14** is bent inward but not enough to have a bonding interaction with zinc ($Zn \cdots O = 4.27 \text{ \AA}$). Instead it is connected to the phenolate oxygen by a hydrogen bond ($O \cdots O = 2.66 \text{ \AA}$), as displayed below. The major conclusion from these findings is that the steric situation of ligand **L** is such that it unambiguously enforces 5-fold coordination in its zinc complexes.



Phosphate Ester Cleavage. The abundance of zinc-containing phosphatases and the biological importance of phosphate transfer have induced numerous mechanistic studies of zinc complex mediated phosphate ester cleavages.^{14,33,54-56} We have contributed to this with preparative and detailed kinetic investigations involving (pyrazolylborato)zinc complexes.⁵⁷⁻⁵⁹ Our

(48) Rodriguez de Barbarin, C. O.; Bailey, N. A.; Fenton, D. E.; He, Q.-Y. *Inorg. Chim. Acta* **1994**, *219*, 205.

(49) Ruf, M.; Weis, K.; Vahrenkamp, H. *Inorg. Chem.* **1997**, *36*, 2130.

(50) Koppenhöfer, A.; Hartmann, U.; Vahrenkamp, H. *Chem. Ber.* **1995**, *128*, 779.

(51) Badura, D.; Vahrenkamp, H. Unpublished results.

(52) Müller, B.; Schneider, A.; Tesmer, M.; Vahrenkamp, H. *Inorg. Chem.* **1999**, *38*, 1900.

(53) Walz, R.; Ruf, M.; Vahrenkamp, H. *Eur. J. Inorg. Chem.* **2001**, 139.

(54) Gellmann, S. H.; Petter, R.; Breslow, R. *J. Am. Chem. Soc.* **1986**, *108*, 2388.

(55) Koike, T.; Kimura, E. *J. Am. Chem. Soc.* **1991**, *113*, 8935.

(56) Jurek, P.; Martell, A. E. *Inorg. Chim. Acta* **1999**, 287, 47.

(57) Weis, K.; Rombach, M.; Ruf, M.; Vahrenkamp, H. *Eur. J. Inorg. Chem.* **1998**, 263.

(58) Weis, K.; Vahrenkamp, H. *Eur. J. Inorg. Chem.* **1998**, 271.

(59) Rombach, M.; Maurer, C.; Weis, K.; Keller, E.; Vahrenkamp, H. *Chem. Eur. J.* **1999**, *5*, 1013.

preparative work proved that the Zn–OH unit is the hydrolytically active nucleophile. In our kinetic work we observed for the first time the strongly negative activation entropies indicative of the four-center association of the Zn–OH and P=O functions in the rate-determining step. The availability of the Zn–OH complex **3** in this work provided a chance to verify the previous conclusions for the L·Zn complexes. Work by Fenton had already provided indirect evidence for the involvement of a Zn–OH complex with a tripodal ligand similar to **L**.³³

As usual for these studies, tris(*p*-nitrophenyl) phosphate (TNP) was chosen for the cleavage reactions which we performed in chloroform because several of the uncharged molecular species involved are insoluble in water. We observed that the aqua complex **1** does not react with TNP but the hydroxo complex **3** does, thereby underlining the essentiality of the Zn–OH function. The reaction of 2 equiv of **3** with 1 equiv of TNP produces a mixture of the phosphate complex L·Zn–OPO(OC₆H₄-*p*-NO₂)₂ (**15**) and the nitrophenolate complex **11**. The individual components of this mixture were prepared from **3** and bis(*p*-nitrophenyl) phosphate (**15**) or *p*-nitrophenol (**11**; see above), respectively. **15** could not be obtained in a pure form but was easily identified by its ³¹P NMR signal at –15.4 ppm. The individual preparations from **3** showed that the formation of **15** is about 2 orders of magnitude slower than the formation of **11**. The nucleophilic strength of **3** is not high enough to effect hydrolytic cleavage of bis(*p*-nitrophenyl) phosphate.

All these observations were favorable for the kinetic study under pseudo-first-order conditions, i.e., with a large excess of **3** for the cleavage of TNP. It can be assumed that the cleavage initially produces **15** and free *p*-nitrophenol. The latter is then converted by a fast reaction with excessive **3** into **11**. The concentration of **11** can be monitored by UV spectroscopy using its unobscured absorption band at 385 nm. It is a measure of the concentration of TNP, and from the UV extinctions *I* the pseudo-first-order rate constant can be derived using the equation

$$\ln[1 - (I/I_{\infty})] = -k_{\text{obs}}t$$

The values of *k*_{obs} were determined this way for six different concentrations of **3** in the presence of 0.1 mM TNP. The resulting plot is given in the Supporting Information. The linearity of the plot points to a clean second-order reaction, as expected. The slope yields a second-order rate constant *k*'' of 0.27 s⁻¹ M⁻¹. The least-squares line does not pass through the origin. This means that hydrolysis occurs also in the absence of **3**, e.g. by traces of water in the reaction system. As mentioned above, the aqua complex **1** is not hydrolytically active. The intercept of the ordinate is small enough to ensure that complex **3** is the essential hydrolytically active species.

Of the previous investigations of phosphate ester hydrolysis with zinc complexes few were done in solvents other than water. As these reactions are generally much faster in water, our rate constant can only be compared with those obtained in similar solvents. We found *k*'' values between 0.45 and 1.55 s⁻¹ M⁻¹ for TNP cleavage by pyrazolylborate–Zn–OH complexes.⁵⁹ Fenton found *k*'' values between 0.90 and 2.21 s⁻¹ M⁻¹ for the cleavage by Zn–OH complexes of ligands similar to **L**.³³ The *k*'' value of 0.27 s⁻¹ M⁻¹ observed here for L·Zn–OH has the same order of magnitude but is considerably smaller than the others. We invoke steric hindrance by **L** to explain this. Just like **L** prevents all its stable zinc complexes from being octahedral, it should make it more difficult than the other ligands to bring in the phosphate substrate and form the four-center Zn–OH/P=O aggregate which is a reaction intermediate or the transition state of the cleavage reaction.

Conclusions

The chemistry of the new ligand **L** has fulfilled the expectations put into it. It is easy to synthesize, it makes stable zinc complexes of great variety, the complexes L·Zn–X are easy to handle due to their molecular nature, they are strictly and reliably five-coordinate, and the functionality of the Zn–X unit can be exploited in the intact L·Zn environment. All these properties provide the L·Zn moiety with the same advantages for five-coordinate zinc that the pyrazolylborate–Zn moiety has for four-coordinate zinc.

The ZnN₃O coordination pattern provided by **L** corresponds to the ligation of zinc in several hydrolytic enzymes. In addition to this structural analogy there exists the functional analogy in the form of the stable complex [L·Zn(OH₂)]⁺ (**1**). **1** is the first isolated monoaquazinc complex for the class of tripodal ligands such as **L**. Its relevance could be demonstrated by the preparation and use of the complex L·Zn–OH (**3**), resulting from deprotonation of **1**. **3** is a general base, being able to deprotonate species HX which then form complexes L·Zn–X. It is also strong enough a base to bind CO₂ from the air forming the trinuclear carbonato complex [(L·Zn)₃(μ₃-CO₃)]⁺ (**5**).

The functional analogy of **1** or **3** with the hydrolytic zinc enzymes could be verified by phosphate ester cleavage. The kinetic data of this reaction show that, although the bulky ligand **L** limits access to the hydrolytically active Zn–OH center, the reactivity of the complex is only slightly lower than that of related systems. Complexes [L·Zn(OH₂)]⁺ and L·Zn(OH) thereby suggest themselves for further biomimetic studies concerning the hydrolysis of phosphates, esters, peptides, and CO₂.

Experimental Section

General Information. The general working techniques were as described previously.⁶⁰ Reactions involving ZnEt₂ were performed in dehydrated solvents and in a nitrogen atmosphere. Starting materials were obtained commercially. Bis(picoly)amine⁴² and 2,4-di-*tert*-butyl-6-(chloromethyl)phenol⁴⁰ were prepared according to the published procedures. The term hexanes is used for petroleum ether boiling between 60 and 70 °C. In those cases where several preparations yielded the same complex only the best procedure is described.

Ligand HL. A solution of bis(picoly)amine (4.00 g, 20.1 mmol) and triethylamine (7.26 g, 71.8 mmol) in dioxane (30 mL) was treated with a solution of 2,4-di-*tert*-butyl-6-(chloromethyl)phenol (5.12 g, 20.1 mmol) in dioxane (20 mL). After stirring of the mixture for 1 day, the triethylammonium chloride was filtered off and the filtrate evaporated to dryness. Chromatography with ethanol/ethyl acetate (7:3) over a 6 × 30 cm silica gel column using UV detection allowed us to separate the product **HL** with a *R*_f value of 0.78. A 5.89 g (70%) amount of **HL** remained as a yellowish solid, mp 96 °C. IR (KBr): 3210 (m, br) (OH), 1591 (s) (C=N). ¹H NMR (CDCl₃): 1.19 [s, 9H, *t*-Bu], 1.38 [s, 9H, *t*-Bu], 3.73 [s, 2H, CH₂–phenol], 3.80 [s, 4H, CH₂–py], 6.80–7.60 [m, 8H, aromatic], 8.48 [d, *J* = 4.9 Hz, 2H, H_α], 10.58 [s, 1H, OH].

Anal. Calcd for C₂₇H₃₅N₃O (*M*_r = 417.6): C, 77.66; H, 8.45; N, 10.06. Found: C, 77.12; H, 8.49; N, 9.44.

Complex 1. A solution of **HL** (300 mg, 0.72 mmol) and KOH (40 mg, 0.72 mmol) in methanol (20 mL) was treated with a solution of Zn(ClO₄)₂·6H₂O (268 mg, 0.72 mmol) in methanol (10 mL). After stirring of the mixture for 30 min, the KClO₄ precipitate was filtered off and the filtrate evaporated to dryness. Crystallization from benzene yielded 259 mg (60%) of **1** as colorless crystals, mp 250 °C. IR (KBr): 3434 (m, br) (OH), 1610 (s) (C=N), 1105 (vs) (ClO₄). ¹H NMR (CDCl₃): 1.22 [s, 9H, *t*-Bu], 1.30 [s, 9H, *t*-Bu], 3.34 [s, 2H, H₂O], 3.82 [s, 2H, CH₂–phenol], 3.97 [d, *J* = 16.3 Hz, 2H, CH₂–py], 4.12

(60) Förster, M.; Burth, R.; Powell, A. K.; Eiche, T.; Vahrenkamp, H. *Chem. Ber.* **1993**, *126*, 2643.

[d, $J = 16.3$ Hz, 2H, CH₂-py], 6.81 [d, $J = 2.6$ Hz, 1H, H_c], 7.10 [d, $J = 2.6$ Hz, 1H, H_a], 7.32–7.90 [m, 6H, aromatic], 8.95 [d, $J = 4.5$ Hz, 2H, H_a].

Anal. Calcd for C₂₇H₃₆ClN₃O₆Zn ($M_r = 599.4$): C, 54.10; H, 6.05; N, 7.01. Found: C, 54.29; H, 6.11; N, 6.32.

Complex 2. A solution of ZnEt₂ (1 M, 0.72 mmol, 0.72 mL) in *n*-hexane was added to a solution of HL (300 mg, 0.72 mmol) in toluene/*n*-hexane (5:1) and stirred for 1 h. Filtration and removal of the solvent from the filtrate left behind 316 mg (86%) of impure **2** as a yellowish solid which decomposes within hours. ¹H NMR (C₆D₆): 0.76 [q, $J = 8.2$ Hz, 2H, Zn-CH₂], 1.37 [s, 9H, *t*-Bu], 1.71 [t, $J = 8.2$ Hz, 3H, ethyl-CH₃], 1.85 [s, 9H, *t*-Bu], 3.45 [s, 2H, CH₂-phenol], 3.74 [d, $J = 15.0$ Hz, 2H, CH₂-py], 4.04 [d, $J = 15.0$ Hz, 2H, CH₂-py], 6.40–7.40 [m, 8H, aromatic], 8.19 [d, $J = 4.6$ Hz, 2H, H_a].

Complex 3. A mixture of **1** (100 mg, 0.17 mmol) and triethylamine (17 mg, 0.17 mmol) in chloroform (5 mL) was stirred for 10 min. Removal of the solvent in vacuo left behind a mixture of triethylammonium perchlorate and **3** as a colorless solid. ¹H NMR (CDCl₃): 1.10 [t, $J = 7.2$ Hz, 9H, CH₃-ethyl], 1.26 [s, 9H, *t*-Bu], 1.35 [s, 9H, *t*-Bu], 2.66 [q, $J = 7.2$ Hz, 6H, N-CH₂], 3.83 [s, 2H, CH₂-phenol], 4.12 [d, $J = 16.6$ Hz, 2H, CH₂-py], 4.23 [d, $J = 16.6$ Hz, 2H, CH₂-py], 6.87 [d, $J = 2.4$ Hz, 1H, H_c], 7.17–7.83 [m, 7H, aromatic], 8.85 [d, $J = 4.8$ Hz, 2H, H_a].

Complex 4. A solution of pyridine (40 mg, 0.50 mmol) in methanol (5 mL) was added to a solution of **1** (300 mg, 0.50 mmol) in methanol (15 mL). After the solution was stirred for 1 h, the solvent was removed in vacuo and the residue crystallized from ethanol, yielding 116 mg (35%) of **4** as a colorless solid, mp 196 °C. IR (KBr): 1608 (s) (C=N), 1098 (vs) (ClO₄). ¹H NMR (CDCl₃): 1.15 [s, 9H, *t*-Bu], 1.30 [s, 9H, *t*-Bu], 3.74 [s, 2H, CH₂-phenol], 4.12 [d, $J = 16.0$ Hz, 2H, CH₂-py], 4.40 [d, $J = 16.0$ Hz, 2H, CH₂-py], 6.78 [d, $J = 2.4$ Hz, 1H, H_c], 7.03 [d, $J = 2.4$ Hz, 1H, H_a], 7.19–8.02 [m, 11H, aromatic], 9.15 [d, $J = 4.6$ Hz, 2H, H_a].

Anal. Calcd for C₃₂H₃₉ClN₄O₅Zn ($M_r = 660.5$): C, 58.19; H, 5.95; N, 8.48; Zn, 9.90. Found: C, 60.21; H, 6.23; N, 8.18; Zn, 9.47.

Complex 5. A solution of **1** (500 mg, 0.83 mmol) in methanol (20 mL) was treated with a solution of NaOH (33 mg, 0.83 mmol) in methanol (10 mL), stirred for 1 h, and left standing in an open beaker for 2 days. A 763 mg (57%) amount of **5** precipitated as colorless crystals, mp 195 °C, which were filtered off and washed with hexanes. IR (KBr): 1708 (w) (C=O_{as}), 1605 (s) (C=N), 1262 (s) (C=O_{sym}), 1101 (vs) (ClO₄). ¹H NMR (CDCl₃): 1.32 [s, 9H, *t*-Bu], 1.55 [s, 9H, *t*-Bu], 3.74 [s, 2H, CH₂-phenol], 3.99 [s, 4H, CH₂-py], 6.76 [d, $J = 2.5$ Hz, 1H, H_c], 7.04 [d, $J = 2.5$ Hz, 1H, H_a], 7.13–7.80 [m, 6H, aromatic], 9.26 [d, $J = 5.2$ Hz, 2H, H_a].

Anal. Calcd for C₈₂H₁₀₂ClN₆O₁₆Zn₃·3H₂O ($M_r = 1605.4 + 54.0$): C, 58.71; H, 6.61; N, 7.52. Found: C, 58.50; H, 6.30; N, 6.96.

Complex 6. A solution of ZnBr₂ (162 mg, 0.72 mmol) in methanol (10 mL) was added to a solution of HL (300 mg, 0.72 mmol) and NaOH (29 mg, 0.72 mmol) in methanol (20 mL). After the solution was stirred for 2 h, diethyl ether (50 mL) was added, precipitating **6**. Recrystallization from methanol yielded 289 mg (64%) of **6** as colorless crystals, mp 268 °C. IR (KBr): 1607 (s) (C=N). ¹H NMR (CDCl₃): 1.16 [s, 9H, *t*-Bu], 1.35 [s, 9H, *t*-Bu], 3.39 [s, 6H, methanol], 3.69 [s, 2H, CH₂-phenol], 3.80 [d, $J = 15.9$ Hz, 2H, CH₂-py], 4.04 [d, $J = 15.9$ Hz, 2H, CH₂-py], 6.71 [d, $J = 2.5$ Hz, 1H, H_c], 7.04 [d, $J = 2.5$ Hz, 1H, H_a], 7.16–7.79 [m, 6H, aromatic], 9.48 [d, $J = 5.0$ Hz, 2H, H_a].

Anal. Calcd for C₂₇H₃₄BrN₃OZn·2CH₃OH ($M_r = 561.9 + 64.1$): C, 55.65; H, 6.76; N, 6.71. Found: C, 55.43; H, 6.74; N, 6.72.

Complex 7. A solution of glacial acetic acid (40 mg, 0.67 mmol) and NaOH (27 mg, 0.67 mmol) in methanol (15 mL) was added to a solution of **1** (400 mg, 0.67 mmol) in methanol (20 mL) and stirred for 1 h. The solvent was removed in vacuo and the residue taken up in 5 mL of dichloromethane. Layering with hexanes yielded, within 3 days, 180 mg (43%) of **7** as colorless crystals, mp 221 °C. IR (KBr): 1609 (vs) (C=O and C=N). ¹H NMR (CDCl₃): 1.08 [s, 9H, *t*-Bu], 1.28 [s, 9H, *t*-Bu], 2.02 [s, 3H, acetate], 3.58 [s, 2H, CH₂-phenol], 4.06 [s, 4H, CH₂-py], 5.32 [s, 2H, CH₂Cl₂], 6.62 [d, $J = 2.6$ Hz, 1H, H_c], 6.83 [d, $J = 2.6$ Hz, 1H, H_a], 7.00–7.66 [m, 6H, aromatic], 8.72 [d, $J = 5.0$ Hz, 2H, H_a].

Anal. Calcd for C₂₉H₃₇N₃O₃Zn·CH₂Cl₂ ($M_r = 541.0 + 84.9$): C, 57.57; H, 6.28; N, 6.71. Found: C, 57.24; H, 6.13; N, 6.95.

Complex 8. To a solution of HL (300 mg, 0.72 mmol) in toluene (10 mL) were added 0.72 mmol (0.72 mL of a 1 M solution in *n*-hexane) of diethylzinc and a solution of phenol (68 mg, 0.72 mmol) in toluene (5 mL). After the mixture was stirring for a few minutes, the product precipitated. Recrystallization from methanol yielded 341 mg (78%) of **8** as a colorless solid, mp 225 °C. IR (KBr): 1606 (s) (C=N). ¹H NMR (CDCl₃): 1.23 [s, 9H, *t*-Bu], 1.36 [s, 9H, *t*-Bu], 3.39 [s, 3H, methanol], 3.80 [s, 2H, CH₂-phenol], 3.88 [d, $J = 15.8$ Hz, 2H, CH₂-py], 4.11 [d, $J = 15.8$ Hz, 2H, CH₂-py], 6.81 [m, 1H, phenolate], 7.12 [d, $J = 2.6$ Hz, 1H, H_c], 7.18 [d, $J = 2.6$ Hz, 1H, H_a], 7.20–7.78 [m, 10H, aromatic], 9.06 [d, $J = 5.0$ Hz, 2H, H_a].

Anal. Calcd for C₃₃H₃₉N₃O₂Zn·CH₃OH ($M_r = 575.1 + 32.0$): C, 67.26; H, 7.14; N, 6.92. Found: C, 67.72; H, 6.90; N, 6.70.

Complex 9. This was made as was **8** from HL (300 mg, 0.72 mmol), with 0.72 mmol of diethylzinc and thiophenol (79 mg, 0.72 mmol). Yield: 294 mg (69%) of **9** as a colorless solid, mp 179 °C. IR (KBr): 1604 (s) (C=N). ¹H NMR (CDCl₃): 1.13 [s, 9H, *t*-Bu], 1.29 [s, 9H, *t*-Bu], 3.65 [s, 2H, CH₂-phenol], 3.90 [d, $J = 15.7$ Hz, 2H, CH₂-py], 4.06 [d, $J = 15.7$ Hz, 2H, CH₂-py], 6.68–7.70 [m, 13H, aromatic], 8.97 [d, $J = 4.9$ Hz, 2H, H_a].

Anal. Calcd for C₃₃H₃₉N₃OSZn ($M_r = 591.2$): C, 67.05; H, 6.65; N, 7.11. Found: C, 66.56; H, 6.70; N, 6.86.

Complex 10. A solution of diphenyl phosphate (168 mg, 0.67 mmol) and NaOH (27 mg, 0.67 mmol) in methanol (15 mL) was added to a solution of **1** (400 mg, 0.67 mmol) in methanol (20 mL). After the solution was stirred for 1 h, the solvent was removed in vacuo and the residue crystallized from acetone, yielding 159 mg (30%) of **10** as colorless crystals, mp 217 °C. IR (KBr): 1607 (s) (C=N), 1281 (s), 1211 (s) (P=O). ¹H NMR (CDCl₃): 1.22 [s, 9H, *t*-Bu], 1.43 [s, 9H, *t*-Bu], 2.09 [s, 6H, acetone], 3.74 [s, 2H, CH₂-phenol], 3.81 [d, $J = 16.0$ Hz, 2H, CH₂-py], 4.08 [d, $J = 16.0$ Hz, 2H, CH₂-py], 6.77 [d, $J = 2.4$ Hz, 1H, H_c], 7.04–7.82 [m, 17H, aromatic], 9.08 [d, $J = 5.2$ Hz, 2H, H_a].

Anal. Calcd for C₃₉H₄₄N₃O₅PZn·(CH₃)₂CO ($M_r = 731.2 + 58.0$): C, 63.92; H, 6.39; N, 5.32. Found: C, 64.51; H, 6.18; N, 5.33.

Complex 11. A solution of *p*-nitrophenol (93 mg, 0.68 mmol) in chloroform (5 mL) was added to a solution of **1** (400 mg, 0.67 mmol) and triethylamine (68 mg, 0.67 mmol) in chloroform (15 mL). After the solution was stirred for 5 min, the solvent was removed in vacuo and the residue crystallized from acetone, yielding 224 mg (54%) of **11** as yellow crystals, mp 238 °C. IR (KBr): 1609 (s) (C=N), 1287 (vs) (N=O). ¹H NMR (CDCl₃): 1.17 [s, 9H, *t*-Bu], 1.27 [s, 9H, *t*-Bu], 3.76 [s, 2H, CH₂-phenol], 3.83 [d, $J = 16.0$ Hz, 2H, CH₂-py], 4.07 [d, $J = 16.0$ Hz, 2H, CH₂-py], 6.74 [d, $J = 2.5$ Hz, 1H, H_c], 7.08 [d, $J = 2.5$ Hz, 1H, H_a], 7.19–8.09 [m, 10H, aromatic], 8.83 [d, $J = 5.0$ Hz, 2H, H_a].

Anal. Calcd for C₃₃H₃₈N₄O₄Zn ($M_r = 620.1$): C, 63.92; H, 6.18; N, 9.04. Found: C, 63.70; H, 6.23; N, 8.87.

Complex 12. To a solution of HL (300 mg, 0.72 mmol) in acetonitrile (10 mL) were added 0.72 mmol of diethylzinc (0.72 mL of a 1 M solution in *n*-hexane). After 5 min of stirring and subsequent heating to 50 °C, a solution of 1-methyluracil (91 mg, 0.72 mmol) in acetonitrile (10 mL) was added. After being stirred for 1 h at 50 °C, the mixture was allowed to cool to room temperature upon which the product was precipitated. Recrystallization from methanol/water (6:1) yielded 142 mg (31%) of **12** as a colorless solid, mp 120 °C. IR (KBr): 1644 (vs) (C=O), 1605 (s) (C=N). ¹H NMR (DMSO-*d*₆): 1.09 [s, 9H, *t*-Bu], 1.30 [s, 9H, *t*-Bu], 3.16 [s, 3H, CH₃-N], 3.47 [s, 2H, CH₂-phenol], 4.22 [d, $J = 15.5$ Hz, 2H, CH₂-py], 4.38 [d, $J = 15.5$ Hz, 2H, CH₂-py], 5.30 [d, $J = 7.4$ Hz, 1H, uracil], 6.64 [d, $J = 2.5$ Hz, 1H, H_c], 6.66 [d, $J = 2.5$ Hz, 1H, H_a], 7.27–7.83 [m, 7H, aromatic + uracil], 8.32 [d, $J = 4.7$ Hz, 2H, H_a].

Anal. Calcd for C₃₂H₃₉N₅O₃Zn·1.5H₂O ($M_r = 607.1 + 27.0$): C, 60.61; H, 6.68; N, 11.04. Found: C, 60.75; H, 6.48; N, 11.04.

Complex 13. A solution of potassium 2-formylphenolate (89 mg, 0.67 mmol) in methanol (15 mL) was added to a solution of **1** (400 mg, 0.67 mmol) in methanol (20 mL). After the mixture was stirred for 1 h, the KClO₄ precipitate was filtered off and the filtrate evaporated to dryness. The residue was taken up in 5 mL of THF and layered

with hexanes. After 3 days, 199 mg (44%) of **13** had separated as yellowish crystals, mp 128 °C. IR (KBr): 1635 (s) (C=O), 1606 (vs) (C=N). ¹H NMR (CDCl₃): 1.13 [s, 9H, *t*-Bu], 1.25 [s, 9H, *t*-Bu], 1.85 [broad, 4H, THF], 3.75 [broad, 4H, THF], 3.78 [s, 2H, CH₂-phenol], 3.93 [d, *J* = 15.6 Hz, 2H, CH₂-py], 4.08 [d, *J* = 15.6 Hz, 2H, CH₂-py], 6.68 [d, *J* = 2.4 Hz, 1H, H_c], 6.93 [d, *J* = 2.4 Hz, 1H, H_a], 7.09–7.69 [m, 10H, aromatic], 8.78 [d, *J* = 4.0 Hz, 2H, H_α], 10.01 [s, 1H, CHO].

Anal. Calcd for C₃₄H₃₉N₃O₃Zn·C₄H₈O (*M*_r = 603.1 + 72.1): C, 67.60; H, 7.02; N, 6.22; Zn, 9.68. Found: C, 66.42; H, 7.05; N, 6.04; Zn 9.43.

Complex 14. Like **13** from **1** (400 mg, 0.67 mmol) and potassium 2-(hydroxymethyl)phenolate (109 mg, 0.67 mmol). The residue after filtration was picked up in 5 mL of acetone and exposed to an atmosphere of diethyl ether. After 4 days, 302 mg (68%) of **14** had separated as yellowish crystals, mp 207 °C. IR (KBr): 1605 (s) (C=N). ¹H NMR (CDCl₃): 1.23 [s, 9H, *t*-Bu], 1.38 [s, 9H, *t*-Bu], 2.09 [s, 6H, acetone], 3.82 [s, 2H, CH₂-phenol], 3.91 [d, *J* = 16.0 Hz, 2H, CH₂-py], 4.14 [d, *J* = 16.0 Hz, 2H, CH₂-py], 4.86 [s, 2H, CH₂OH], 6.56–7.79 [m, 12H, aromatic], 8.93 [d, *J* = 5.2 Hz, 2H, H_α].

Anal. Calcd for C₃₄H₄₁N₃O₃Zn·(CH₃)₂CO (*M*_r = 605.1 + 58.1): C, 67.01; H, 7.14; N, 6.34. Found: C, 67.09; H, 7.12; N, 6.24.

Reaction of 3 with TNP. A solution of 0.17 mmol of **3** in 15 mL of chloroform was prepared as described above. Tris(*p*-nitrophenyl) phosphate (39 mg, 0.085 mmol) was added with stirring and the resulting solution monitored by ³¹P NMR and UV. After 2 h the ³¹P NMR resonance of TNP (−20.0 ppm) had completely disappeared and been replaced by the resonance of **15** (−15.4 ppm). The solution had turned yellow, and the UV spectrum confirmed the formation of **11** by the unperturbed absorption band at 385 nm. When an equimolar amount of bis(*p*-nitrophenyl) phosphate (58 mg, 0.17 mmol) was added to the solution of **3**, the UV spectrum showed no sign of the formation of *p*-nitrophenol or **11**, and the NMR spectrum showed the quantitative formation of **15**.

Kinetic Data. Measurements were performed on a JASCO V-570 UV spectrometer by continually recording the 385 nm absorption of **11**. Chloroform (UV quality) was used as a solvent. Samples of TNP were taken from a 2.00 mM stock solution. Stock solutions of **3** (0.100 M) were prepared prior to use as described above from **1** and triethylamine. The measuring chamber and the solutions were thermostated to 25.0 °C for 30 min prior to the measurements and during the kinetic runs. Reagents were mixed in the quartz cuvettes. The concentration of TNP was 0.100 mM for all measurements; that of **3**

was adjusted to 5.00, 10.00, 15.00, 20.00, and 25.00 mM, corresponding to a 50-, 100-, 150-, 200-, and 250-fold excess.

The absorption intensities were recorded for 5*t*_{1/2}. The *I* value at this time was taken as *I*_∞. Reaction times for 90% conversion were 2–8 h. Up to a conversion of 75% the reactions were cleanly of first order with a correlation coefficient greater than 0.997. For the computations the measurements up to 2*t*_{1/2} were included. The reproducibility of the *I* values was within 10%. Each data point in Figure 8 represents the average of three measurements.

Structure Determinations. Crystals of **5**, **10**, **11**, **13**, and **14** were obtained directly from the preparations, those of **1** and **9** by layering THF solutions with hexanes. They were immersed in fluorinated polyether oil for the measurements on a Nonius CAD4 diffractometer with graphite-monochromatized Mo K α radiation (λ = 0.7107 Å) at 180 K (room temperature for **13** and **14**). No absorption corrections were applied. The structures were solved with direct methods and refined anisotropically with the SHELX program suite.⁶¹ Hydrogen atoms were included with fixed distances and isotropic temperature factors 1.5 times those of their attached atoms. Parameters were refined against *F*². Complexes **1**, **5**, and **7** show a rotational disorder of one of their *tert*-butyl groups; complex **10** has one of its phosphate phenyl groups disordered over two positions. Drawings were produced with SCHAKAL.⁶² The crystallographic data are listed in the Supporting Information.

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Supporting Information Available: A table with all crystallographic details, fully labeled ORTEP plots for all seven structure determinations, a plot of the rate constants of the phosphate cleavage, and seven crystallographic files, in CIF format. This information is available free of charge via the Internet at <http://pubs.acs.org>.

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(61) Sheldrick, G. M. *SHELXS-86 and SHELXL-93*; Universität Göttingen: Göttingen, Germany, 1986 and 1993.

(62) Keller, E. *SCHAKAL for Windows*; Universität Freiburg: Freiburg, Germany, 1999.