Coordination Chemistry of Rh(III) Porphyrins: Complexes with Hydrazine, Disulfide, and Diselenide Bridging Ligands

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Rh(III) porphyrin complexes with bridging hydrazine and substituted hydrazine ligands were characterized in solution by ¹H NMR spectroscopy and in the solid state by X-ray diffraction. Addition of further ligand to these species afforded 1:1 complexes in which methylhydrazine and *N*,*N*-dimethylhydrazine preferentially bound to the Rh center through the substituted nitrogen atom, as evidenced by ¹H NMR chemical shifts. An alkylated Rh(III) porphyrin was isolated as a decomposition product of the reaction of *N*,*N*-dimethylhydrazine with Rh(III) porphyrin in the presence of light and oxygen. Me₂Se₂ and Me₂S₂ formed bridging and nonbridging complexes with Rh(III) porphyrin, analogous to that observed with *N*,*N*'-dimethylhydrazine.

Introduction

The crystal structure of a nitrogenase enzyme cofactor¹ revealed an Fe-Mo sulfide cluster believed to be responsible for the biological fixation of nitrogen. These findings renewed interest in the coordination chemistry of dinitrogen with transition metals with a view to both understanding the natural system and developing artificial nitrogen fixation processes. Of particular importance is the reduction of coordinated nitrogen to hydrazine and ammonia or its incorporation into organic molecules under mild conditions. Recent advances in this field have included the formation of ammonia by reaction of a tungsten-dinitrogen complex with an acidic $Ru-\eta^2-H_2$ complex² and reaction of a Zr₂ (μ - η^2 : η^2 -N₂) species with H₂ to form an N-H bond.³ The complexes of hydrazines with metalloporphyrins have generated interest as models for intermediates in the reduction of dinitrogen. Collman et al. have reported the preparation and redox chemistry of covalently linked Ru(II) porphyrin dimers in which hydrazine was bound between the already cofacial porphyrins.^{4–6} We have previously described hydrazine binding to steroid-capped Zn(II) porphyrins,7 and more recently, others have reported syntheses of Ru(IV) and Ti(IV) porphyrin complexes with hydrazido ligands.^{8,9} Continuing our studies of rhodium porphyrins,^{10,11} in this

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paper, we describe the preparation of Rh(III) porphyrin complexes with hydrazine ligands and demonstrate the formation of analogous species with disulfide and diselenide ligands in solution.

Experimental Section

Instrumentation. NMR spectra were recorded on Bruker DRX 500 (500.1 MHz for ¹H, 125.7 MHz for ¹³C), AM 400 or DRX 400 (400.1 MHz for ¹H, 100.6 MHz for ¹³C), and DPX 250 or AC 250 (250.1 MHz for ¹H, 62.9 MHz for ¹³C) instruments. Chemical shifts are quoted in parts per million with respect to tetramethylsilane (TMS). Infrared spectra were recorded on a Perkin-Elmer Paragon 1000 FTIR spectrometer at 4 cm⁻¹ resolution or better. UV–visible spectra were recorded on a Hewlett-Packard 8452A diode array spectrophotometer.

X-ray Crystallography. Diffraction data were collected in Cambridge on a Rigaku R-Axis IIc or Nonius Kappa CCD device, or for small crystals, were collected at the Daresbury SRS (UK), Station 9.8, using a Bruker AXS SMART CCD area detector.^{12–14} Structures were solved with either SHELXS-97 or SIR92.^{15–17} Refinement was carried out with SHELXL-97.¹⁸ Extensive disorder was often observed in the solubilizing hexyl chains, typical of this kind of structure.¹⁹ The carbon

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Table 1. Crystallographic Data for 2, 4, 5, 7, and	graphic Data for 2, 4, 5, 7, ar	1d 9
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	2	4	5	7	9
empirical formula	$C_{120}H_{156}I_2N_{10}Rh_2$	C ₆₀ H ₇₉ IN ₅ Rh	$C_{123}H_{162}Cl_2I_2N_{10}Rh_2$	$C_{123}H_{162}Cl_2I_2N_{10}Rh_2$	$C_{122}H_{160}Cl_2I_2N_{10}Rh_2$
fw	2198.17	1100.09	2311.15	2311.15	2297.12
cryst size, mm	$0.09\times0.04\times0.02$	$0.07\times0.06\times0.02$	$0.16 \times 0.14 \times 0.03$	$0.20 \times 0.04 \times 0.04$	$0.12\times0.06\times0.06$
space group	$P\overline{1}$	$P2_{1}/c$	$P\overline{1}$	$P\overline{1}$	$P\overline{1}$
a, Å	16.2361(13)	16.3158(13)	15.1170(4)	15.231(2)	15.1665(9)
b, Å	18.6695(15)	26.431(2)	18.7980(8)	18.438(3)	18.7297(11)
<i>c</i> , Å	20.6876(16)	12.7523(10)	21.9870(9)	22.377(3)	21.7848(12)
α, deg	72.554(2)	90	68.4040(14)	65.943(5)	67.709(5)
β , deg	87.348(2)	100.830(2)	81.457(2)	83.689(5)	81.345(5)
γ, deg	67.114(2)	90	79.604(2)	81.358(5)	79.304(5)
$V, Å^3$	5493.8(8)	5401.5(8)	5691.0(4)	5664.8(14)	5604.5(6)
Ζ	2	4	2	2	2
$\rho_{\rm calc},{\rm mg}/{\rm m}^3$	1.329	1.353	1.349	1.355	1.361
R	0.0895	0.0632	0.0863	0.1544	0.0702
wR2	0.2170	0.1177	0.1988	0.3393	0.1891
GOF on F^2	1.065	1.073	1.032	1.143	0.997

atoms in these disordered groups were refined with isotropic temperature factors and restrained to give chains with a reasonable geometry. Even with synchrotron radiation, crystals were found to be weakly diffracting, resulting in a relatively high R1 value.

¹H NMR Titrations. CDCl₃ for use in titrations was stored over anhydrous K_2CO_3 in a refrigerator. Typically, a solution of Rh(III) porphyrin (1)·MeOH (4 mg) in CDCl₃ (600 μ L) was titrated with 5- μ L aliquots of a solution of ligand in CDCl₃ at a concentration of 72 mM. ¹H spectra (400 MHz) were acquired at room temperature. Me₂Se₂ was filtered through neutral alumina before use.

UV–Visible Titrations. A 3.0-mL aliquot of a 4 μ M solution of 1·MeOH in CH₂Cl₂/MeOH (200 μ L MeOH/100 mL of solvent) was titrated at 25 °C with 10- μ L aliquots of a 75 μ M solution of ligand in the same solvent mixture. The cuvette was magnetically stirred for ~30 s prior to recording each spectrum. Spectra were processed by factor analysis followed by least-squares fitting using the program Specfit.²⁰

Materials. 1 was prepared as previously described¹⁰ and recrystallized from CHCl₃ layered with MeOH. Other chemicals were obtained from Aldrich and used without further purification. Merck 60 mesh silica gel was used for column chromatography.



Synthesis. 1₂•**NH**₂**NH**₂ (2). Hydrazine monohydrate (0.5 μ L, 10 μ mol) in water (10 mL) was added to a mixture of a solution of 1 (10 mg, 9.2 μ mol) in CH₂Cl₂ (1 mL) and water (5 mL). After 150 min at room temperature, an additional portion of CH₂Cl₂ (20 mL) was added, and the organic layer was separated, washed with water (2 × 30 mL), and dried (MgSO₄). Evaporation of the solvent afforded the product as an orange solid (5 mg, 49%).

For structure determination, crystals of **2** were obtained from a solution of **2** and **3** (prepared by adding > 0.5 equiv of N₂H₄·H₂O to **1**) in toluene layered with MeOH (see Table 1).

¹H NMR (400 MHz, CDCl₃): δ 9.37 (s, 4H, meso), 7.84 (m, 8H, Ar), 7.55 (m, 8H, Ar), 7.25 (m, Ar), 3.72 (m, 16H, hex H¹), 2.24 (s, 24H, CH₃), 2.04 (m, 16H, hex H²), 1.84 (m, 16H, hex H³), 1.63 (m, 16H, hex H⁴), 1.52 (m, 16H, hex H⁵), 1.05 (t, *J* = 7 Hz, 24H, hex H⁶), -11.03 (s, 4H, NH₂). ¹³C NMR (100.6 MHz, CDCl₃): δ 143.5, 141.6, 138.6, 138.0, 137.8, 133.0, 132.9, 128.5, 127.6, 127.3, 118.3, 96.8, 33.4, 31.9, 30.5, 27.0, 22.9, 15.1, 14.2. UV-vis (CH₂Cl₂) λ_{max} , nm:

360, 408, 528, 558. IR (CCl₄) ν_{max} (cm⁻¹): 3287 (w), 3231 (w), 3147 (w), 2957 (s), 2930 (s), 2858 (m), 1466 (m).



1·NH₃ Complex (4). A solution of **1** (5 mg, 4.6 μ mol) in CH₂Cl₂ (1 mL) was treated with vapor from aqueous NH₃ solution for 30 min at room temperature. The solvent was evaporated by blowing with N₂, and the product was obtained quantitatively after being dried in vacuo. Crystals for structure determination were obtained from a toluene solution layered with MeOH.

¹H NMR (400 MHz, CDCl₃): δ 10.23 (s, 2H, meso), 8.14 (d, J =7 Hz, 2H, Ar), 7.98 (d, J = 7 Hz, 2H, Ar), 7.78–7.68 (m, 6H, Ar), 4.00 (m, 4H, hex H¹), 3.89 (m, 4H, hex H¹), 2.45 (s, 12H, CH₃), 2.22 (m, 8H, hex H²), 1.80 (m, 8H, hex H³), 1.55 (m, 8H, hex H⁴), 1.43 (m, 8H, hex H⁵), 0.95 (t, J = 7 Hz, 12H, hex H⁶), -5.77 (s, 3H, NH₃). ¹³C NMR (100.6 MHz, CDCl₃): δ 144.1, 142.8, 140.5, 140.1, 138.6, 134.0, 132.8, 128.3, 127.8, 127.2, 119.3, 98.8, 33.4, 32.0, 30.3, 27.1, 22.8, 15.6, 14.2. UV-vis (CH₂Cl₂) λ_{max} (nm): 360, 420, 532, 562. MALDI MS m/z: [M - NH₃]⁺ 1084. IR (CCl₄) ν_{max} (cm⁻¹): 3381 (w), 2957 (s), 2930 (s), 2858 (m). Anal. Calcd for C₆₀H₇₉IN₅Rh: C, 65.51; H, 7.24; N, 6.37. Found: C, 65.70; H, 7.43; N, 6.03.

1₂•NHMeNHMe (5). A solution of 1•NHMeNHMe (6) (10 mg, 8.7 μ mol) in CH₂Cl₂/hexane (1:1) was passed through a silica column. The solvent was evaporated to afford the product as an orange solid (7.1 mg, 73%). Crystals for structure determination were obtained from a CH₂Cl₂ solution layered with MeOH.

¹H NMR (400 MHz, CDCl₃): δ 9.64 (s, 4H, meso), 7.79 (m, 12H, Ar), 7.53 (m, 8H, Ar), 4.17 (m, 4H, hex H¹), 4.05 (m, 4H, hex H¹),

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3.79 (m, 8H, hex H¹), 2.32 (s, 12H, CH₃), 2.29 (s, 12H, CH₃), 1.96 (m, 4H, hex H²), 1.85 (m, 12H, hex H²), 1.65 (m, 16H, hex H³), 1.47–1.33 (m, 32H, hex H^{4.5}), 0.93, 0.90 (2 × t, J = 7 Hz, 24H, hex H⁶), -7.32 (d, J = 6 Hz, 6H, NHCH₃), -9.40 (t, J = 6 Hz, 2H, NHCH₃).

1·NHMeNHMe (6). To a solution of **1·**MeOH (12.2 mg, 11 μ mol) in CH₂Cl₂ (20 mL) was added *N*,*N'*-dimethylhydrazine dihydrochloride (17.3 mg, 0.13 mmol) in water (20 mL). NaOH (10%, 2 drops) was added, and the mixture was shaken. The organic phase was separated, dried (MgSO₄), and evaporated to an orange solid (10.6 mg, 85%).

¹H NMR (400 MHz, CDCl₃): δ 10.28 (s, 2H, meso), 8.10 (m, 2H, Ar), 8.04 (m, 2H, Ar), 7.76 (m, 6H, Ar), 4.03 (m, 4H, hex H¹), 3.91 (m, 4H, hex H¹), 2 × 2.48 (s, 12H, CH₃), 2.21 (m, 8H, hex H²), 1.77 (m, 8H, hex H³), 1.52 (m, 8H, hex H⁴), 1.41 (m, 8H, hex H⁵), 2 × 0.92 (t, J = 7 Hz, hex H⁶), -0.66 (s, 3H, NH*CH*₃), -3.27 (d, J = 6 Hz, 3H, RhNH*CH*₃), -4.09 (s, 1H, N*H*CH₃), -4.90 (m, 1H, RhN*H*CH₃). ¹³C NMR (100.6 MHz, CDCl₃): δ 2 × 144.3, 142.7, 140.5, 140.4, 140.1, 140.0, 138.8, 138.7, 133.8, 133.0, 128.3, 127.7, 127.3, 119.5, 98.9, 34.3, 33.8, 2 × 33.3, 32.0, 2 × 30.1, 2 × 27.0, 22.7, 15.5, 14.1.

1₂·NH₂NMe₂ (7). 7 was prepared by titration of a solution of **1**·MeOH (4.0 mg, 3.6 μ mol) with *N*,*N*-dimethylhydrazine in CDCl₃. Material for crystal growth was prepared by passing a solution of **1** (5.2 mg, 4.8 μ mol) and *N*,*N*-dimethylhydrazine (1 μ L, 13 μ mol) in CH₂Cl₂ (0.5 mL) and hexane (0.5 mL) through a silica column. Porphyrin was washed from the column with CH₂Cl₂/hexane (1:1), and the solvent was evaporated. X-ray quality crystals were obtained from a CH₂Cl₂ solution layered with MeOH.

¹H NMR (400 MHz, CDCl₃): δ 9.77 (s, 2H, meso), 0.37 (s, 2H, meso), 7.80 (m, 12H, Ar), 7.62 (m, 6H, Ar), 7.37 (d, J = 7 Hz, 2H, Ar), 4.04 (m, 8H, hex H¹), 3.76 (m, 8H, hex H¹), 2.38 (s, 12H, CH₃), 2.29 (s, 12H, CH₃), 1.85 (m, 16H, hex H²), 1.63 (m, 16H, hex H³), 1.50–1.30 (m, 32H, hex H^{4.5}), 0.93 (t, J = 7 Hz, 12H, hex H⁶), 0.88 (t, J = 7 Hz, 12H, hex H⁶), -7.83 (s, 6H, NCH₃), -10.24 (s, 2H, NH₂).

1₂·NHMeNH₂ (9). 9 was prepared by titration of a solution of **1**· MeOH (4.2 mg, 3.8 μ mol) with methylhydrazine in CDCl₃. After the titration, the sample was eluted through a silica column with hexane/CH₂Cl₂ (1:1) and evaporated to an orange solid. This was dissolved in CH₂Cl₂ and layered with MeOH to obtain crystals for structure determination.

¹H NMR (400 MHz, CDCl₃): δ 9.47 (s, 2H, meso), 9.43 (s, 2H, meso), 7.93 (d, J = 7 Hz, 2H, Ar), 7.82 (m, 8H, Ar), 7.52 (m, 8H, Ar), 7.33 (d, J = 4 Hz, 2H, Ar), 4.03 (m, 4H, hex H¹), 3.89 (m, 4H, hex H¹), 3.75 (m, 4H, hex H¹), 3.66 (m, 4H, hex H¹), 2.33 (s, 6H, CH₃), 2 × 2.27 (2 × s, 18H, CH₃), 2.17–1.34 (m, 64H, hex H^{2–5}), 0.96 (m, 24H, hex H⁶), -7.79 (d, J = 6 Hz, 3H, NHCH₃), -10.00 (m, 2H, NHMeNHH), -10.34 (m, 1H, NHH).

Results and Discussion

1 was prepared as a methanol complex according to the literature.¹⁰ A solution of 1·MeOH in CDCl₃ was titrated with hydrazine monohydrate, and the product distribution was monitored by ¹H NMR spectroscopy. Hydrazine displaced the coordinated methanol and initially formed a 2:1 complex (2) with a bridging hydrazine ligand as evidenced by a highly shielded resonance at -11.03 ppm and an upfield shifted porphyrin meso resonance at 9.37 ppm. Collman et al. observed a similarly upfield shifted resonance at -10.15 ppm, corresponding to hydrazine bound within a Ru(II) cofacial diporphyrin.⁶ Addition of >0.5 equiv of hydrazine resulted in the breakup of the dimeric complex to afford the 1:1 complex 3, which displayed a pair of shielded NH₂ resonances at -3.77and -3.28 ppm and a porphyrin meso resonance at 10.25 ppm. 2 and 3 were in slow exchange on the NMR chemical shift time scale, although equilibration occurred too rapidly to readily monitor the kinetics of this process by ¹H NMR. The more upfield NH₂ resonance, assigned to RhNH₂NH₂, remained sharp throughout the titration, whereas the peak at -3.28 ppm, assigned to RhNH₂NH₂, progressively broadened. Broadening



Figure 1. Molecular structure of **2**. Hydrogen atoms and porphyrin β substituents have been omitted for clarity.

can be attributed to exchange of the latter protons catalyzed by traces of acidic impurity. In support of this hypothesis, exposure of the sample to trifluoroacetic acid vapor further broadened the RhNH₂NH₂ resonance without affecting the width of the RhNH₂NH₂ peak.

Passage of a solution of 2 and 3 through a silica gel column eluted with CH₂Cl₂/hexane (1:1) retained hydrazine on the silica and converted 3 to 2. In this way, a mixture with initial meso resonances in the intensity ratio of 1:0.77 (3:2) was converted to a mixture in which this ratio was 1:9.6. Starting from 10 mg of 1·MeOH, a total of 8.0 mg of material was recovered after this treatment, confirming that there is a true conversion of 3 to 2 on the silica gel and that this filtration does not simply remove 3 to leave a mixture enriched in 2. An alternative route to 2 was extraction of an aqueous solution of hydrazine with a solution of 1 in CH₂Cl₂ followed by a standard aqueous workup. The infrared spectrum of 2 in CCl₄ solution showed a band at 3231 cm^{-1} and a further pair of very weak bands in the same spectral region, which may be assigned to the NH stretches of the hydrazine.²¹

Crystallization of a mixture of **3** and **2** from a toluene solution layered with methanol resulted in preferential crystallization of **2**, most likely because of the poor solubility of this complex in methanol. A single-crystal X-ray diffraction study revealed that in the solid state the hydrazine adopts a trans geometry with a Rh–N–N–Rh torsion angle of 178.5(1)° (Figure 1). The N–N bond length of 1.47(1) Å is in the range typically reported for bridging hydrazine complexes.²² The two inequivalent Rh–N_{axial} bond lengths are 2.030(9) and 2.089(9) Å. The porphyrins are oriented with almost parallel planes but with the axes joining the opposite meso positions twisted slightly, apparently to avoid steric clash of the peripheral substituents. The deviation of the porphyrins from planarity is irregular and not easily classified into one of the commonly observed distortion modes.²³

For comparison with 2 and 3, the complex of 1 with ammonia, 4, was prepared by exposure of a CH₂Cl₂ solution of 1 to

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Figure 2. Molecular structure of 5. Hydrogen atoms have been omitted.

Table 2. Selected Bond Lengths (Å), Angles (deg), and Torsions for 2, 4, and 5 $\,$

	2	4	5
Rh-N _{axial}	2.030(9), 2.089(9)	2.113(5)	2.195(7), 2.199(7)
Rh-I _{axial}	2.622(1), 2.620(1)	2.6447(6)	2.6261(8), 2.6261(9)
Iaxial-Rh-Naxial	177.7(3), 179.5(3)	178.0(1)	176.4(2), 175.9(2)
Naxial-Naxial	1.47(1)		1.479(9)
Rh-Naxial-Naxial	120.1(6), 115.8(6)		118.2(5), 117.3(5)
Rh-Naxial-Naxial-Rh	178.5(5)		151.4(3)
C-Naxial-Naxial-C			58.2(9)
Rh-N _{axial} -C			114.5(5), 114.6(5)

ammonia vapor. The ¹H NMR spectrum of **4** displays a singlet at -5.77 ppm, arising from the ammonia ligand. We have been unable to observe any coupling between the NH₃ protons and ¹⁰³Rh. The X-ray structure of **4** revealed a Rh–N_{axial} bond length of 2.113(5) Å, comparable with the corresponding bond lengths of **2** (see Table 2).

Treatment of 1 in dichloromethane with an aqueous solution of *N*,*N*'-dimethylhydrazine dihydrochloride basified by addition of NaOH afforded the 1:1 complex 6. Both the ¹³C and ¹H NMR spectra of this compound are complicated by the formation of a chiral center at the bound nitrogen atom of the dimethylhydrazine. This removes mirror symmetry from the porphyrin, leaving only average C_2 symmetry, assuming free rotation about the Rh-Naxial bond. Splittings are observed in the hexyl and β -methyl resonances but not the meso and aryl resonances. Coupling is observed between the proton of the coordinated nitrogen atom at -4.90 ppm and its methyl group at -3.27ppm with J = 6 Hz. The value of ${}^{3}J_{\text{HNCH}}$ in N,N'-dimethylhydrazine has been reported as 6.1 Hz.²⁴ The other methyl group of the ligand appears as a singlet at -0.66 ppm while the NH resonance at -4.09 ppm is broad, presumably because of proton exchange. Passage of this sample through silica gel and recrystallization from a chloroform/methanol mixture afforded the 2:1 complex 5. Although this could exist as a mixture of diastereomers, ¹H NMR spectroscopy suggested that only one was present or that interconversion was rapid on the NMR time scale. A single meso resonance and doubled hexyl and β -methyl resonances were observed, consistent with the presence of a chiral center at the nitrogen atom of the bound ligand but with rapid rotation about the Rh-Naxial bond. The methyl groups of the ligand at -7.32 ppm are split into a doublet with ${}^{3}J_{\text{HNCH}} =$ 6 Hz, and a corresponding splitting is observed in the NH resonance at -9.40 ppm. In the solid-state structure (Figure 2), the ligand is ordered, and the chirality of each of the nitrogen atoms in a molecule is the same, but overall, the crystal is



Figure 3. Upfield region of the 500 MHz ¹H NMR spectrum of 9 showing the NH resonances.

centrosymmetric with a molecule of each enantiomer in the unit cell. The porphyrin planes are not parallel but are tilted by $28.31(5)^{\circ}$ to accommodate the methyl groups of the ligand. The axes joining the opposite substituted meso positions of each porphyrin are rotated by approximately 90° with respect to each other to avoid steric clash. The Rh–N_{axial} bond lengths of 2.195(7) and 2.199(7) Å are comparable to the value of 2.216(8) Å reported for the Rh–N distance of the [RhCl4(PPh₃)(NHMeNHMe)]⁻ anion.²⁵

Titrations of 1. MeOH in CDCl₃ with methylhydrazine and N,N-dimethylhydrazine were carried out. On addition of < 0.5equiv of these substituted hydrazines, the expected bridging complexes 7 and 9 were observed, as evidenced by highly shielded NH and methyl resonances. The resonances of methylhydrazine in 9 all appear as multiplets (Figure 3). The methyl resonance is assigned to a doublet at -7.79 ppm with ${}^{3}J_{\text{HNCH}}$ = 6 Hz. ${}^{3}J_{\text{HNCH}}$ for free methylhydrazine has been reported as 6.25 Hz.26 A COSY spectrum revealed a cross-peak between this resonance and a multiplet at -10.00 ppm that integrated to two protons. This peak is assigned to the overlapping resonances of the NHCH₃ proton and a single proton of the diastereotopic NH₂ group. The resonance of the other proton of this group resembles a triplet at -10.34 ppm. In contrast to 9, the ¹H NMR spectrum of 7 is simple with singlets at -7.83 and -10.24 ppm assigned to the N(CH₃)₂ and NH₂ resonances, respectively. Addition of >0.5 equiv of the hydrazines dissociated the dimers to afford the 1:1 complexes 8 and 10. Here, the situation is complicated by the possibility of isomeric forms in which the hydrazine coordinates through different nitrogen atoms. Although theoretical and NMR studies²⁷ have supported the belief that the substituted nitrogen of alkyl hydrazines is the more basic, in reported crystal structures of methylhydrazine and N,Ndimethylhydrazine, these ligands bound through the unsubstituted and less sterically hindered nitrogen atom.²⁸⁻³³ The ability of the Rh(III) porphyrin to act as a shift reagent for bound ligands has now permitted the elucidation of the binding mode of these substituted hydrazines. We propose that the N,Ndimethylhydrazine ligand of 8 is bound through the substituted nitrogen atom on the basis of the chemical shifts of the methyl groups (-3.31 ppm) and by comparison with the spectrum of

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6. The situation for 10 is not so clear-cut, although the observation of a doublet at -3.24 ppm with J = 6 Hz, assigned to the hydrazine methyl group, suggests that the hydrazine is bound through the substituted nitrogen atom in the major species in solution. However, a minor singlet at -0.79 ppm can be tentatively assigned to the methyl group of the isomeric complex in which the unsubstituted nitrogen is bound to the rhodium center. In support of this assignment was the observation of a cross-peak in the NOESY spectrum, acquired at 40 °C, between the peaks at -0.79 and -3.24 ppm. At room temperature, this cross-peak was extremely weak, consistent with an increasing rate of exchange of the methylhydrazine between the two binding modes as the temperature was increased. The NH protons of the minor species cannot be located with confidence, although the peak at -3.91 ppm assigned to the NHCH₃ resonance of the major isomer appears to be overlapping with another minor resonance.

Passage of samples of **8** and **10** through silica gel, eluted with dichloromethane/hexane (1:1), converted them into **7** and **9**, respectively. The solid-state structures of **7** and **9** were found to resemble that of **5**, except the ligands were disordered. As in **5**, the porphyrins are canted to accommodate the methyl groups. The angles between the best fit planes of **7** and **9** are 25.49(9) and 25.94(3)°, respectively.

Initial attempts to obtain crystals of **7** by diffusion of methanol into a solution of **8** and excess *N*,*N*-dimethylhydrazine afforded a mixture of crystals from which could be identified the organometallic derivative **11** by X-ray diffraction and ¹H NMR, in comparison with an authentic sample.¹⁰ By integration of the



meso resonances in the ¹H NMR spectrum, **11** was estimated to comprise 6% of the porphyrinic material in this sample, with the remainder largely attributed to **7**. Further experiments, in which samples of **8** and excess *N*,*N*-dimethylhydrazine in CDCl₃ were exposed to combinations of air and light, revealed that a combination of both accelerates the formation of **11** and other unidentified products. Similarly, others have reported the formation of metal alkyls from decomposition of alkyl hydrazine complexes.^{34–38} The IR spectrum of **7** showed a sharp but weak peak at 3216 cm⁻¹, whereas **9** gave a group of peaks at 3224 cm⁻¹, which are attributed to NH stretches by analogy with **2**.

As Me_2S_2 and Me_2S_2 are also known to act as bridging ligands,^{39–43} we predicted that they would form complexes with **1** analogous to **5**. An ¹H NMR titration of **1** in CDCl₃ with

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Me₂S₂ led to broadening of the porphyrin resonances and an upfield shift of the meso resonance to 9.8 ppm. Evidence for a bridged dimer was the observation of a broad peak at -5.6 ppm assigned to the methyl groups of the bridging ligand. On addition of >0.5 equiv of Me₂S₂, this peak progressively disappeared, to be replaced by peaks at 0.1 and -2.7 ppm assigned to the inequivalent methyl groups of a 1:1 complex in which a single sulfur atom binds to the rhodium center. This was accompanied by a sharpening and downfield shift of the meso resonance to 10.22 ppm. Likewise, a titration of **1** in CDCl₃ with Me₂Se₂ afforded a bridged dimer with a meso resonance at 9.62 ppm and ligand-methyl resonance at -5.21 ppm. The 1:1 complex displayed a meso resonance at 10.21 ppm and methyl groups at 0.58 and -2.30 ppm. In this case, peaks were sharper, and the meso resonances were in slow exchange on the 400 MHz ¹H NMR chemical shift time scale, indicating that the diselenide ligand has a lower exchange rate than the disulfide ligand. Additional support for these complexes was obtained from UVvisible titrations of 1 with Me₂S₂ and Me₂Se₂ in dichloromethane with methanol added as a competitive ligand. On addition of the chalcogenide, the Soret band of 1 at 412 nm was replaced by a red-shifted Soret band, observed at 422 nm in the case of Me₂S₂ and 428 nm for Me₂Se₂. In the former instance, we have been able to account for the observed spectral changes during the course of the titration using a model consisting solely of 1, Me₂S₂, and 1·Me₂S₂ with a binding constant of log(K) = 5.7. The concentration of 2:1 complex is likely to be negligible at the porphyrin concentration employed for the titration experiment. However, the data for Me₂Se₂ cannot be satisfactorily modeled without inclusion of the species $1_2 \cdot Me_2Se_2$ with a broad Soret band centered at 426 nm. In this case, we calculate log-(K) = 6.6 for the binding of a single porphyrin to Me₂Se₂, and log(K) = 5.4 for the binding of a second porphyrin unit, implying weakly anticooperative behavior.

In summary, we have prepared complexes of Rh(III) porphyrins with bridging hydrazine and substituted hydrazine ligands and characterized these in solution by ¹H NMR spectroscopy and in the solid state by X-ray diffraction. Addition of further hydrazine to these species afforded 1:1 complexes. On the basis of ¹H NMR chemical shift evidence, we propose that methylhydrazine and *N*,*N*-dimethylhydrazine preferentially bind to the Rh center through the substituted nitrogen atom. A methylated Rh(III) porphyrin was obtained as a decomposition product of the reaction of *N*,*N*-dimethylhydrazine with Rh(III) porphyrin in a process accelerated by oxygen and light. Me₂Se₂ and Me₂S₂ behaved analogously to *N*,*N*²-dimethylhydrazine and formed bridging and nonbridging complexes with Rh(III) porphyrin.

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Supporting Information Available: ¹H NMR spectra of 1-10 and titrations of 1 with Me₂S₂ and Me₂Se₂. Crystallographic data of 2, 4, 5, 7, and 9 in CIF format. Equations of least-squares planes and deviations from them. This material is available free of charge via the Internet at http://pubs.acs.org.

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