Inorganic Chemistry

N-H and N-C Bond Activation of Pyrimidinic Nucleobases and Nucleosides Promoted by an Osmium Polyhydride

Miguel A. Esteruelas,*^{,†} Jorge García-Raboso,[†] Montserrat Oliván,[†] and Enrique Oñate[†]

[†]Departamento de Química Inorgánica-Instituto de Síntesis Química y Catálisis Homogénea (ISQCH), Universidad de Zaragoza-CSIC, 50009 Zaragoza, Spain

Supporting Information

ABSTRACT: Complex $OsH_6(P^iPr_3)_2$ (1) reacts with 1methylthymine and 1-methyluracil to give $OsH_3(P^iPr_3)_2$ (nucleobase') (2, 3) containing the deprotonated nucleobases (nucleobase') κ^2 -N,O coordinated by the nitrogen atom at position 3 and the oxygen bonded to the carbon atom of the ring at position 4. Similarly, the reactions of 1 with thymidine, 5-methyluridine, deoxyuridine, and uridine lead to $OsH_3(P^iPr_3)_2$ (nucleoside') (4–7) with the deprotonated nucleoside (nucleoside') κ^2 -N,O coordinated by the nitrogen atom at position 3 and the oxygen bonded to the carbon atom



at position 4 of the nucleobases. Treatment of complexes 5 and 7, containing nucleosides derived from ribose, with $OsH_2Cl_2(P^iPr_3)_2(8)$ in the presence of Et₃N affords dinuclear species $OsH_3(P^iPr_3)_2(nucleobase')$ -(ribose)($P^iPr_3)_2H_2Os(9, 10)$ formed by two different metal fragments. Complex 1 also promotes the cleavage of the N–C bond of 2–7 to give the dinuclear species $\{OsH_3(P^iPr_3)_2\}_2(nucleobase')$ (11, 12) with the nucleobase skeleton (nucleobase') κ^2 -N,O coordinated to both metal fragments. These compounds can be also prepared by reaction of 1 with 0.5 equiv of thymine and uracil. The use of 1:1 hexahydride:nucleobase molar ratios gives rise to the preferred formation of the mononuclear complexes $OsH_3(P^iPr_3)_2(nucleobase'')$ (13, 14; nucleobase''' = monodeprotonated thymine or uracil). The X-ray structures of complexes 6, 11, and 14 are also reported.

INTRODUCTION

Cisplatin, and the second generation alternatives carboplatin and oxaliplatin, are still the most widely used chemotherapeutic agents for cancer.¹ However, some toxicological side-effects of cisplatin and the drug resistance developed by some tumors have stimulated research toward the synthesis of complexes with another transition metals, in particular some group 8 compounds.² Thus, a few reports of anticancer active halfsandwich osmium d⁶-complexes have recently appeared.³

DNA is an important potential biological target for many metal-based anticancer agents. Distortions of DNA structure often correlate with anticancer activity. Hence, it is of great important to understand DNA binding properties of transitionmetal species.⁴ It is generally accepted that the solvolytic activation of *cis*-Pt^{II}(NH₃)₂X₂-drugs involves the displacement of leaving groups X by water molecules, which are subsequently replaced by the DNA donor heteroatoms.¹ In contrast to square-planar platinum d⁸-species, octahedral osmium d⁶complexes are relatively inert toward substitution reactions due to the dependence of the crystal field activation energy on Δ_0 .⁵ Thus, in order to develop new models of osmium anticancer drugs, we reasoned that osmium d²-species containing some Brønsted base as a ligand could be a promising alternative, since the fair acidity of the hydrogen atoms bonded to the nucleobase heteroatoms. In this context, it should be noted that a nitride-osmium(VI) complex has recently shown anticancer activity *in vivo*.⁶

The saturated hexahydride d^2 -complex OsH₆(PⁱPr₃)₂ activates C–H bonds of a broad number of organic molecules⁷ and, by protonation with weak Brønsted acids, releases molecular hydrogen to afford osmium-hydride d⁴-species, which contain the corresponding conjugated Brønsted base as a ligand.⁸ Thymine as a constituent of DNA and uracil as a constituent of RNA can both form transition-metal compounds resulting from deprotonation and subsequent coordination of the nitrogen atom at position 3 and the oxygen atoms bonded to the carbons at positions 2 and 4, depending on the coordination metal.^{1b,9} Recently, deprotonated 1-methylthymine acting as a chelating ligand for the *cis*-Pd(C₆F₃H₂)₂ and *cis*-Pd(C₆F₅)₂ moieties through the nitrogen atom at position 3 and the oxygen atom bonded to the carbon at position 4 has been also reported.¹⁰

In the search for models of osmium anticancer drugs, we have studied the reactivity of the hexahydride d²-complex $OsH_6(P^iPr_3)_2^{11}$ toward pyrimidinic N-methyl nucleobases and nucleosides. In this paper we report N–H and N–C bond activations of these compounds, in a sequential manner.

Received:
 March 27, 2012

 Published:
 May 9, 2012

Inorganic Chemistry

RESULTS AND DISCUSSION

N–H Bond Activation of Nucleobases. Treatment of toluene solutions of $OsH_6(P^iPr_3)_2$ (1) with 1.0 equiv of 1-methylthymine and 1-methyluracil for 3 h, under reflux, leads to complexes 2 and 3 according to Scheme 1. They result from the

Scheme 1



deprotonation of the nitrogen atom at position 3 and the chelate coordination of the generated anion, by the deprotonated nitrogen and the oxygen bonded to the carbon at position 4.

Complexes 2 and 3 were isolated as white solids in 70% and 61% yields, respectively. As expected, for three inequivalent hydride ligands, the ¹H NMR spectra in toluene- d_8 at 203 K show three hydride resonances at about -10.6 (H_A), -13.5 (H_B), and -14.9 (H_C). These resonances are temperature dependent (Figure 1). The coalescence between the H_B and H_C



Figure 1. High field of the ¹H{³¹P} NMR spectra (300 MHz, toluene d_8) of complex **2** as a function of temperature.

resonances occurs between 233 and 243 K, whereas a single hydride signal is observed at temperatures higher than 273 K. This is consistent with the operation of two thermally activated site exchange processes, in agreement with the behavior of related OsH_3 -derivatives.^{8b,c} The exchange mechanism implies Os-H stretching, H-H shortening, and subsequent rotation of the resulting dihydrogen ligand. Since the activation barrier for both exchanges is similar, between 10 and 11 kcal·mol⁻¹, the transition states containing the dihydrogen ligand trans disposed to the nitrogen or oxygen atoms of the deprotonated nucleobase appear to be equally favored. The ³¹P{¹H} NMR spectra at room temperature contain a singlet at about 34 ppm, according to the presence of equivalent phosphines in the complexes. In the ${}^{13}C{}^{1}H$ NMR spectra, the resonance corresponding to the coordinated carbonyl group appears at about 177 ppm whereas that due to the free one is observed at about 154 ppm, highfield shifted by about 23 ppm.

Thymidine, 5-methyluridine, deoxyuridine, and uridine show a behavior similar to 1-methylthymine and 1-methyluracil. Treatment of toluene solutions of 1 with 1.0 equiv of the nucleosides for 3 h, under reflux, affords the related derivatives 4-7 (Scheme 2), containing the corresponding deprotonated nucleoside that is chelated by the nitrogen atom at position 3 and the oxygen bonded to the carbon at position 4 of the nucleobase.



Complexes 4-7 were isolated as white (4 and 5) and yellow (6 and 7) solids in 48-60% yield. Complex 6 was characterized by X-ray diffraction analysis.¹² The structure (Figure 2) proves



Figure 2. Molecular diagram of complex 6. Selected bond lengths (Å) and angles (°): Os-P(1) = 2.3439(7), Os-P(2) = 2.3461(7), Os-O(1) = 2.2361(19), Os-N(1) = 2.210(2), O(1)-C(1) = 1.267(3), O(2)-C(2) = 1.229(3); P(1)-Os-P(2) = 167.87(2), N(1)-Os-O(1) = 59.01(7).

the deprotonation of the nitrogen atom at position 3 (N(1))and the chelate coordination of the nucleobase by the latter and the oxygen atom bonded to the carbon atom at position 4 (O(1)). Thus, the geometry around the osmium atom can be rationalized as a distorted pentagonal bipyramidal with the phosphine ligands occupying axial positions (P(1)-Os-P(2) = $167.87(2)^\circ)$. The metal coordination sphere is completed by the deprotonated nucleoside, which acts with a bite angle N(1)–Os–O(1) of 59.01(7)°, and the hydride ligands. The Os–N(1) and Os–O(1) bond lengths are 2.210(2) and 2.2361(19) Å, respectively. As expected the separation between the coordinated oxygen atom O(1) and C(1), 1.267(3) Å, is slightly longer than the separation between the free oxygen atom O(2) and C(2), 1.229(3) Å. An extended view of the structure (Figure 3) reveals that the molecules of the complex



Figure 3. View of the interactions via hydrogen bonding in the structure of complex 6 [symmetry codes: (I) x + 1/2, -y + 1/2, -z + 1; (II) x - 1/2, -y + 1/2, -z + 1; (III) x - 1, y, z; (IV) x + 1, y, z].

form polymers in the *ab* plane by means of intermolecular $O(5)-H(5)\cdots O(4)$ hydrogen bonds between sugars of adjacent molecules, in addition to $O(4)-H(4)\cdots O(2)$ hydrogen bonds between the O(4)-H(4) substituent of the fivemembered ring of the sugar of one molecule and the free O(2) oxygen atom of an neighboring molecule. In agreement with this, the separations $H(5)\cdots O(4)$ and $H(4)\cdots O(2)$ of 1.876(2) and 1.880(2) Å, respectively, are shorter than the sum of the van der Waals radii of hydrogen and oxygen $(r_{vdw}(H) = 1.20$ Å, $r_{vdw}(O) = 1.52$ Å).¹³ Furthermore, the $O(4)\cdots O(5)$ and $O(4)\cdots O(2)$ separations are 2.729(3) and 2.705(3) Å, and the angles O(4)-H(4)-O(2) and O(5)-H(5)-O(4) are almost linear at $166.7(2)^{\circ}$ and $179.4(2)^{\circ}$, respectively.

The ¹H, ³¹P{¹H}, and ¹³C{¹H} NMR spectra of 4–7 are consistent with the structure shown in Figure 2 and agree well with those of 2 and 3. The ¹H NMR spectra in toluene- d_8 at 213 K show three hydride resonances at about –10.5, –13.6, and –15.5 ppm, whereas at room temperature only one hydride signal is observed. The activation barriers for the thermally activated site exchange processes are also, in the four cases, between 10 and 11 kcal·mol⁻¹. The ³¹P{¹H} NMR spectra at 298 K show a singlet at about 34 ppm. In the ¹³C{¹H} NMR spectra the resonance due to the coordinated carbonyl group is observed at about 177 ppm, whereas that corresponding to the free carbonyl group appears at about 154 ppm.

The protons of the OH functional groups of the ribose fivemembered ring of **5** and 7 can be replaced by the $OsH_2(P^iPr_3)_2$ metal fragment. Thus, the treatment of toluene solutions of these compounds with 1.0 equiv of the known complex $OsH_2Cl_2(P^iPr_3)_2$ (8) in the presence of 4.0 equiv of Et_3N , at room temperature, for 20 min leads to the dinuclear derivatives 9 and 10 and $[Et_3NH]Cl$. These complexes were isolated as yellow solids in 59% and 64% yield, according to Scheme 3.

Scheme 3



Their dinuclear character is strongly supported by the highresolution electrospray mass spectra, in acetonitrile, which show the molecular peaks at 1283.5936 (9) and 1269.5683 (10) with the isotope pattern as expected, as shown in Figure 4 for 9.

The ¹H and ³¹P{¹H} NMR spectra of 9 and 10 are consistent with the presence of two different metal fragments in these compounds. One of them is a seven-coordinate $OsH_3(P^iPr_3)_2(nucleobase)$ moiety similar to those of 2-7.



Figure 4. (a) Selected region of the ESI-HRMS spectrum of complex 9 showing the isotopic distribution of $[M + H]^+$ (highest peak at 1283.5936). (b) Simulated pattern for the molecular formula $C_{46}H_{101}N_2O_6Os_2P_4$ $[M + H]^+$.

The other is a 16-valence electrons osmium(IV) $OsH_2(P^iPr_3)_2(ribose)$ moiety with a structure in the solid state, characteristic for unsaturated $OsH_2P_2L_2$ d⁴-species, which can be described as a C_2 -square antiprism with two missing vertexes.^{8a,14} Thus, as is shown in Figure 5 for 9, the ¹H NMR



Figure 5. ¹H NMR spectrum ($C_6D_{6^j}$ 500 MHz) of complex 9. (A) Methyl resonances of the PⁱPr₃ ligands. (B) Hydride resonances.

spectra at room temperature contain doublets of virtual triplets corresponding to the P'Pr3 methyl groups of the sevencoordinate metal fragment and a double doublet due to the P'Pr₃ methyl groups of the six-coordinate moiety. In the high field region of the spectra the OsH₃ unit of these compounds displays a broad resonance at about -13 ppm, whereas the OsH₂ unit gives rise to a triplet with a H–P coupling constant of 41 Hz at about -18 ppm. In agreement with 2-7, the resonance at about -13 ppm is converted into three signals, at about -10.5, -13.6, and -14.9 ppm, at 233 K. However, at this temperature, the resonance at about -18 ppm generates a complex pattern as expected for the presence of two slowly interconverting dihydride-osmium(IV) isomers, one having C_2 symmetry and the other with no symmetry.¹⁵ The ³¹P{¹H} NMR spectra contain two singlets between 35 and 33 ppm. In addition, it should be mentioned that the replacement of the COH-protons of the ribose subtituents of 5 and 7 by the $OsH_2(P^iPr_3)_2$ metal fragment has a marked influence in the chemical shifts of the ¹³C{¹H} NMR ribose-CO-Os resonances of 9 and 10 (92.6 and 90.9 (9); 93.2 and 90.9 (10) ppm), which appear downfield shifted by about 20 ppm with regard to the respective COH resonances of 5 and 7 (75.6 and 70.6 (5); 76.0 and 70.6 (7) ppm).

C–N Bond Activation of the Coordinated Nucleobase. The reactions providing straightforward examples of C–N single bond activation are rare in comparison with the known C–H bond activation processes.¹⁶ In spite of this, the hexahydride complex 1 does not only promote the cleavage of the N–H bond of 1-methylthymine and 1-methyluracil but also the activation of the respective N–CH₃ bond. Thus, the treatment of toluene solutions of 2 and 3 with 1.0 equiv of 1 for 8.5 and 2 h, respectively, under reflux leads to the dinuclear complexes 11 and 12 (Scheme 4), as a result of the demethylation of the deprotonated nucleobases and the coordination of the carbon atom at position 2 to a new OsH₃(PⁱPr₃)₂ metal fragment. These compounds were isolated as white solids in 65% (11) and 70% (12) yield.

The hexahydride complex 1 also promotes the rupture of the N-sugar bond of 4-7 to afford the dinuclear compounds 11 and 12. However, the reactions are not clean and complex mixtures are generated. The formation of these mixtures is

Scheme 4



consistent with the versatile reactivity of the osmium polyhydrides,¹⁷ in particular that of 1,^{7b-e,18} with alcohols, aldehydes, and ketones. As expected, the mixtures resulting from the reactions of the ribose derivatives 5 and 7 with 1 also contain the dinuclear complexes 9 and 10.

Complex 11 was crystallized from the mixture generated from the reaction of 4 with 1 and characterized by X-ray diffraction analysis. Figure 6 shows a view of the molecule. The



Figure 6. Molecular diagram of complex 11. Selected bond lengths (Å) and angles (°): Os(1)-P(1) = 2.3366(8), Os(1)-P(2) = 2.3407(9), Os(2)-P(3) = 2.3356(9), Os(2)-P(4) = 2.3209(9), Os(1)-O(1) = 2.227(2), Os(1)-N(1) = 2.220(3), Os(2)-O(2) = 2.260(2), Os(2)-N(2) = 2.163(3), C(1)-O(1) = 1.283(4), C(2)-O(2) = 1.297(4); P(1)-Os(1)-P(2) = 167.46(3), P(3)-Os(2)-P(4) = 169.06(3), N(1)-Os(1)-O(1) = 59.77(10), N(2)-Os(2)-O(2) = 59.60(10).

structure proves the rupture of the N-sugar bond of 4 and the unprecedented coordination of the nucleobase skeleton, as a bridge ligand coordinated κ^2 -N,O to both osmium atoms. Thus, the geometry around each metal center can be rationalized as a distorted pentagonal bipyramid with the phosphines occupying axial positions (P(1)-Os-(1)-P(2) = 167.46(3)°, P(3)-Os(2)-P(4) = 169.06(3)°). The metal coordination spheres are completed by the donor atoms of the nucleobase skeleton, which act with bite angles N(1)-Os(1)-O(1) and N(2)-

Os(2)-O(2) of 59.77(10)° and 59.60(10)°, respectively, and the hydride ligands. The Os(1)-N(1) bond length of 2.220(3) Å is about 0.06 Å longer than the Os(2)-N(2) separation of 2.163(3) Å, while the Os(1)-O(1) distance of 2.227(2) Å is about 0.03 Å shorter than the Os(2)-O(2) bond length of 2.260(2) Å. The four distances compare well with the respective bond length of **6**. The C(1)-O(1) distance of 1.283(4) Å is statistically identical with the C(2)-O(2) bond length of 1.297(4) Å.

The ¹H NMR spectra of **11** and **12** in toluene- d_8 at 203 K are consistent with the structure shown in Figure 6. As shown in Figure 7 for **11**, they contain six high field resonances between



Figure 7. High field of the ¹H{³¹P} NMR spectra (400 MHz, toluene d_8) of complex **11** as a function of temperature.

-10 and -16 ppm, corresponding to the six inequivalent hydride ligands. At room temperature the different OsH₃ units share the same chemical shift. As a consequence of this and in agreement with the operation of four thermally activated site exchange processes, the spectra at 303 K show only one hydride signal at about -14 ppm. The activation barrier for all exchanges is similar to those of 2-10. The ${}^{31}P{}^{1}H{}$ NMR spectra at room temperature contain two singlets at 36.9 and 30.9 (11) and 36.7 and 30.9 (12) ppm. In the ${}^{13}C{}^{1}H{}$ NMR spectra the resonances due to the coordinated carbonyl groups appear at 176.9 and 165.9 (11) and 178.8 and 167.4 (12) ppm.

Complexes 11 and 12 can be also prepared by treatment of toluene solutions of 1 with 0.5 equiv of thymine and uracil, respectively, for 3 h, under reflux. Their formation involves the double deprotonation of the nucleobase. The use of 1:1 hexahydride:nucleobase molar ratios leads to 0.2:1 mixtures of the dinuclear compounds 11 and 12 and the mononuclear derivatives 13 and 14 (Scheme 5), as a result of the preferred monodeprotonation of the nucleobases. Complexes 13 and 14, which were isolated as white solids in 66% and 61% yield, contain a deprotonated nucleobase κ^2 -N,O-coordinated by the ONO-nitrogen atom and the oxygen bonded to the carbon adjacent to the CR unit (R = CH₃, H).

The uracil derivate complex 14 was characterized by X-ray diffraction analysis. The structure has four chemically equivalent but crystallographically independent molecules in the asymmetric unit. Figure 8 shows a drawing of one of them. The geometry around the osmium atom is as the observed ones in 6 and 11; i.e., a distorted pentagonal bipyramid with axial phosphines $(P(1)-Os-P(2) = 171.42(6)^{\circ}-167.64(9)^{\circ})$. The



Figure 8. Molecular diagram of complex 14. Selected bond lengths (Å) and angles (°): Os(1)-P(1) 2.3330(1), 2.3414(18), 2.329(2), 2.298(2); Os(1)-P(2) 2.3442(17), 2.3526(18), 2.326(2), 2.310(2); Os(1)-O(1) 2.266(4), 2.231(4), 2.212(4), 2.253(5); Os(1)-N(1) 2.175(5), 2.195(5), 2.180(6), 2.168(5); P(1)-Os(1)-P(2) 170.34(6), 171.42(6), 169.27(7), 167.64(9); O(1)-Os(1)-N(2) 58.85(17), 59.41(16), 59.60(18), 59.28(18).

metal coordination sphere is completed by the bidentate nucleobase $(N(1)-Os(1)-O(1) = 59.60(18)^{\circ}-58.85(17)^{\circ})$ and the hydrides. The Os(1)-O(1) (2.231(4)-2.212(4) Å) and Os(1)-N(1) = 2.195(5)-2.168(5) Å) bond lengths compare well with those of **6** and **11**.

An extended view of the structure (Figure 9) reveals that the molecules are associated, to form pairs, through intermolecular $N-H\cdots O$ hydrogen bonds between the free heteroatoms of the nucleobases. One of the pairs implies molecules of the same asymmetric unit. The other two molecules are associated with molecules of adjacent asymmetric units. The H…O separations are between 1.680(5) and 2.00(5) Å, whereas the $N-H\cdots O$ angles lie in the range 165.8(3)°-173.4(3)°. The separations $N\cdots O$ range from 2.759(8) to 2.805(7) Å.

The ¹H, ³¹P{¹H} and ¹³C{¹H} NMR spectra of 13 and 14 are consistent with the structure shown in Figure 8 and agree well with those of 2–7 and 9–12. In addition to three hydride resonances between –10 and –15.5 ppm, the ¹H NMR spectra in toluene- d_8 at 203 K contain the NH-resonances at 12.46 (13) and 12.14 (14). The ³¹P{¹H} NMR spectra show singlets at 33.9 (13) and 34.1 (14). In agreement with 2–7, the ¹³C{¹H} NMR spectra show the resonances due to the



Figure 9. View of the interactions via hydrogen bonding in the structure of complex 14 [symmetry codes: (I) x, -y + 3/2, z + 1/2; (II) x, -y + 1/2, z + 1/2].

coordinated carbonyl group, 178.0 (13) and 178.1 (14) ppm, shifted by about 20 ppm toward lower field with regard to those corresponding to the free carbonyl group, 158.0 (13) and 157.8 (14) ppm.

CONCLUSION

This study has revealed that the hexahydride $OsH_6(P^iPr_3)_2$ promotes the N-H and N-C cleavages of pyrimidinic Nmethyl nucleobases and nucleosides, in a sequential manner. The N-H rupture leads to trihydride-osmium(IV) complexes containing the deprotonated nucleobase or nucleoside κ^2 -N,O coordinated by the nitrogen atom at position 3 and the oxygen bonded to the carbon atom of the ring at position 4. The N–C bond activation processes afford dinuclear derivatives showing an unprecedented coordination of the nucleobase skeletons, as bridge ligands κ^2 -N,O to two Os^(IV)H₃(P'Pr₃)₂ metal fragments. Complexes containing nucleosides derived from ribose react with $OsH_2Cl_2(P^iPr_3)_2$ in the presence of a Brønsted base to give also unprecedented dinuclear species formed by two different metal fragments of osmium(IV): a seven-coordinate $OsH_3(P'Pr_3)_2$ (nucleobase) moiety and a six-coordinate 16valence electrons $OsH_2(P'Pr_3)_2(ribose)$ unit.

EXPERIMENTAL SECTION

General Information. All reactions were carried out with rigorous exclusion of air using Schlenk-tube techniques. Solvents (except methanol that was dried over magnesium and distilled under argon) were obtained oxygen- and water-free from an MBraun solvent purification apparatus. ¹H, ³¹P{¹H}, and ¹³C{¹H} NMR spectra were recorded on Bruker 300 ARX, Bruker Avance 300 MHz, Bruker Avance 400 MHz, and Bruker Avance 500 MHz instruments. Chemical shifts (expressed in parts per million) are referenced to residual solvent peaks (¹H, ${}^{13}C{{}^{1}H}$) or to external 85% H_3PO_4 $({}^{31}P{}^{1}H{})$. Coupling constants J and N are given in hertz. Infrared spectra were recorded on a Perkin-Elmer Spectrum 100 spectrometer as neat solids. C, H, and N analyses were carried out in a Perkin-Elmer 2400 CHNS/O analyzer. High-resolution electrospray mass spectra were acquired using a MicroTOF-Q hybrid quadrupole time-of-flight spectrometer (Bruker Daltonics, Bremen, Germany). OsH₆(PⁱPr₃)₂ (1) and $OsH_2Cl_2(P^iPr_3)_2$ (8) were prepared by published methods.

Reaction of OsH_6(P^{\circ}P_{3})_2 with 1-Methylthymine: Preparation of 2. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of 1-methylthymine (26.9 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a white solid. Yield: 88.4 mg (70%). Anal. Calcd for $C_{24}H_{52}N_2O_2OsP_2$: C, 44.15; H, 8.03; N, 4.29. Found: C, 44.56; H, 8.27; N, 4.06. ESI-HRMS

(*m*/*z*): calcd for $C_{24}H_{51}N_2O_2OSP_2$ [M - H]⁺ 653.0365; found 653.3036. IR (neat compound, cm⁻¹): ν (OsH) 2105 (w); ν (C==O) 1653 (s), 1629 (s); ν (C==C) 1518 (s). ¹H NMR (300 MHz, C₆D₆, 298 K): δ 6.07 (s, 1H, =CH), 2.87 (s, 3H, N-CH₃), 2.01 (m, 6H, PCH(CH₃)₂), 1.69 (s, 3H, CH₃), 1.24 (dvt, J_{H-H} = 6.6, N = 13.5, 18H, PCH(CH₃)₂), 1.22 (dvt, J_{H-H} = 6.9, N = 13.5, 18H, PCH(CH₃)₂), 1.22 (dvt, J_{H-H} = 6.9, N = 13.5, 18H, PCH(CH₃)₂), -13.08 (br, 3H, Os-H). ¹H{³¹P} NMR (300 MHz, toluene-*d*₈, 203 K, high-field region): δ -10.59 (d, J_{H-H} = 18.6, 1H, Os-H), -13.51 (dd, J_{H-H} = 18.6, J_{H-H} = 22.5, 1H, Os-H), -14.85 (d, J_{H-H} = 22.5, 1H, Os-H), -13.68 (tr, J_{C-P} = 1.5, OC-Os), 154.5 (s, CO), 142.1 (s, =CH), 105.1 (s, =CCH₃), 36.7 (s, N-CH₃), 27.3 (vt, N = 23.7, PCH(CH₃)₂), 20.5, 20.2 (both s, PCH(CH₃)₂), 10.6 (s, CH₃).

Reaction of OsH₆(PⁱPr₃)₂ with 1-Methyluracil: Preparation of 3. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of 1-methyluracil (24.7 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a white solid. Yield: 75 mg (61%). Anal. Calcd for C₂₃H₅₀N₂O₂OsP₂: C, 43.24; H, 7.89; N, 4.38. Found: C, 43.55; H, 8.07; N, 4.14. ESI-HRMS (m/ z): calcd for $C_{23}H_{49}N_2O_2OsP_2 [M - H]^+$ 639.2879; found 639.2900. IR (neat compound, cm⁻¹): ν (Os–H) 2127 (m); ν (C=O) 1654 (s), 1613 (s); ν (C=C) 1535 (s). ¹H NMR (300 MHz, C₆D₆, 298 K): δ 6.12 (d, J_{H-H} = 7.2, 1H, =CH), 5.02 (d, J_{H-H} = 7.2, 1H, =CH), 2.79 (s, 3H, N-CH₃), 2.02 (m, 6H, PCH(CH₃)₂), 1.24 (dvt, $J_{H-H} = 7.2$, N = 13.5, 18H, PCH(CH₃)₂), 1.22 (dvt, $J_{H-H} = 7.2$, N = 13.5, 18H, PCH(CH₃)₂), -13.12 (br, 3H, Os-H). ¹H{³¹P} NMR (400 MHz, toluene- d_{8} , 203 K, high-field region): δ -10.60 (d, J_{H-H} = 9.4, 1H, Os-H), -13.58 (dd, J_{H-H} = 9.4, J_{H-H} = 23.4, 1H, Os-H), -14.89 (d, J_{H-H} = 23.4, 1H, Os-H). ³¹P{¹H} NMR (121.4 MHz, C₆D₆, 298 K): δ 34.0 (s). ¹³C{¹H} NMR plus HMBC (101 MHz, C₆D₆, 298 K): δ 177.1 (s, OC-Os), 154.2 (s, CO), 145.3 (s, =CH), 97.1 (s, =CHCO), 37.0 (s, N-CH₃), 27.2 (vt, N = 23.9, PCH(CH₃)₂), 20.4 (s, $PCH(CH_3)_2$

Reaction of OsH₆(PⁱPr₃)₂ with Thymidine: Preparation of 4. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of thymidine (46.9 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a white solid. Yield: 83 mg (60%). Anal. Calcd for C₂₈H₅₈N₂O₅OsP₂: C, 44.55; H, 7.74; N, 3.71. Found: C, 44.87; H, 8.03; N, 3.42. ESI-HRMS (m/z): calcd for $C_{28}H_{57}N_2O_5OsP_2\ [M\ -\ H]^+$ 755.3353; found 755.3313. IR (neat compound, cm⁻¹): ν (OH) 3336 (br); ν (Os–H) 2118 (w); ν (C=O) 1649 (s), 1619 (s); ν (C=C) 1523 (s). ¹H NMR (400 MHz, C₆D₆, 298 K): δ 7.30 (s, 1H, =CH), 6.24 (dd, J_{H-H} = 8.5, J_{H-H} = 6.8, 1H, NCH), 4.77 (br, 1H, CHOH), 4.18 (br, 3H, CHCH₂ and 2 OH), 3.90 (d, J_{H-H} = 11.2, 1H, CH₂OH), 3.82 (d, J_{H-H} = 11.2, 1H, CH₂OH), 2.59 (m, 1H, CH₂), 2.42 (m, 1H, CH₂), 1.99 (m, 6H, PCH(CH₃)₂), 1.79 (s, 3H, CH₃), 1.21 (dvt, $J_{H-H} = 9.4$, N = 14.5, 18H, PCH(CH₃)₂), 1.19 (dvt, $J_{H-H} = 9.1$, N = 13.8, 18H, PCH(CH₃)₂), -13.22 (br, 3H, OsH). ${}^{1}H{}^{31}P{}$ NMR (400 MHz, toluene- d_{8} , 203 K, high-field region): δ -10.50 (s, 1H, OsH), -13.58 (br, 1H, OsH), -15.36 (br, 1H, OsH). ³¹P{¹H} NMR (121.4 MHz, C₆D₆, 298 K): δ 33.7 (s). ¹³C{¹H} NMR plus HMBC (101 MHz, C₆D₆, 298 K): δ 176.6 (s, CO-Os), 154.4 (s, CO), 139.4 (s, =CH), 106.8 (s, =CCH₃), 89.7 (s, N-CH), 88.5 (s, CHCH₂), 72.4 (s, CHOH), 63.1 (s, CH₂OH), 40.8 (s, CH_2), 27.3 (vt, N = 23.6, $PCH(CH_3)_2$), 27.2 (vt, N = 23.8, PCH(CH₃)₂), 20.4, 20.3, 20.2, 20.1 (all s, PCH(CH₃)₂), 10.8 (s, CH₃).

Reaction of OsH_6(P^iPr_3)_2 with 5-Methyluridine: Preparation of 5. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of 5-methyluridine (49.9 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a white

solid. Yield: 87 mg (58%). Anal. Calcd for C₂₈H₅₈N₂O₆OsP₂: C, 43.62; H, 7.58; N, 3.63. Found: C, 43.27; H, 7.21; N, 3.35. ESI-HRMS (m/ z): calcd for C₂₈H₅₇N₂O₆OsP₂ [M - H]⁺ 771.3302; found 771.3265. IR (neat compound, cm⁻¹): ν (OH) 3371 (br); ν (OsH) 2142 (m); ν (C=O) 1648 (s), 1617 (s); ν (C=C) 1526 (s). ¹H NMR (400 MHz, C₆D₆, 298 K): δ 7.34 (s, 1H, =CH), 5.84 (d, J_{H-H} = 3.4, 1H, NCH), 5.35 (br, 1H, OH), 4.95 (br, 1H, OH), 4.85 (dd, $J_{H-H} = 4.8$, $J_{\rm H-H}$ = 4.8, 1H, CHOH), 4.71 (dd, $J_{\rm H-H}$ = 3.4, $J_{\rm H-H}$ = 4.8, 1H, CHOH), 4.33 (br s, 1H, CH₂CH), 4.08 (br, 1H, OH), 3.97 (d, J_{H-H} = 11.2, 1H, CH₂OH), 3.88 (d, J_{H-H} = 11.2, 1H, CH₂OH), 1.98 (m, 6H, $PCH(CH_3)_2$, 1.76 (s, 3H, CH₃), 1.20 (dvt, J_{H-H} = 6.8, N = 13.2, 18H, $PCH(CH_3)_2$, 1.19 (dvt, $J_{H-H} = 6.8$, N = 13.2, 18H, $PCH(CH_3)_2$), -13.22 (br, 3H, OsH). ${}^{1}H{}^{31}P{}$ NMR (400 MHz, toluene- d_8 , 203 K, high-field region): δ -10.49 (br, 1H, OsH), -13.57 (br, 1H, OsH), -15.37 (br, 1H, OsH). ³¹P{¹H} NMR (121.4 MHz, C₆D₆, 298 K): δ 33.6 (s). ¹³C{¹H} NMR plus HMBC (101 MHz, C₆D₆, 298 K): δ 176.8 (s, CO-Os), 155.2 (s, CO), 140.2 (s, =CH), 106.7 (s, =CCH₃), 95.5 (s, N-CH), 85.9 (s, CHCH₂), 75.6 (s, CHOH), 70.6 (s, CHOH), 62.2 (s, CH₂OH), 27.2 (vt, N = 23.9, PCH(CH₃)₂), 27.1 (vt, N = 23.9, PCH(CH₃)₂), 20.4, 20.3, 20.2, 20.1 (all s, PCH(CH₃)₂), 10.7 (s, CH₃)

Reaction of OsH₆(PⁱPr₃)₂ with Deoxyuridine: Preparation of 6. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of deoxyuridine (44.1 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a yellow solid. Yield: 85 mg (59%). Anal. Calcd for C₂₇H₅₆N₂O₅OsP₂: C, 43.77; H, 7.62; N, 3.78. Found: C, 44.06; H, 8.01; N, 4.03. ESI-HRMS (m/z): calcd for $C_{27}H_{55}N_2O_5OsP_2$ [M - H]⁺ 741.3197; found 741.3208. IR (neat compound, cm⁻¹): ν (OH) 3387 (br); ν (OsH) 2146 (w); ν (C=O) 1647 (s), 1632 (s); ν (C=C) 1536 (s). ¹H NMR (300 MHz, C₆D₆, 298 K): δ 7.26 (d, J_{H-H} = 7.5, 1H, =CH), 6.12 (dd, J_{H-H} = 8.5, J_{H-H} = 6.3, 1H, NCH), 5.24 (d, J_{H-H} = 7.5, 1H, =CH), 4.56 (br, 1H, CHOH), 4.01 (br s, 1H, CHCH₂), 3.80 (dd, J_{H-H} = 11.1, J_{H-H} = 3, 1H, CH_2OH), 3.68 (d, J_{H-H} = 11.1, 1H, CH_2OH), 3.17 (br, 2H, 2 OH), 2.50 (m, 1H, CH₂), 2.28 (m, 1H, CH₂), 2.00 (m, 6H, PCH(CH₃)₂), 1.24 (dvt, $J_{H-H} = 6.9$, N = 13.2, 18H, PCH(CH₃)₂), 1.19 (dvt, $J_{H-H} =$ 6.9, N = 11.7, 18H, PCH(CH₃)₂), -13.21 (br, 3H, OsH). ¹H(³¹P) NMR (300 MHz, toluene- d_8 , 223 K, high-field region): δ –10.44 (br, 1H, OsH), -13.61 (br, 1H, OsH), -15.41 (br, 1H, OsH). $^{31}P\{^{1}H\}$ NMR (121.4 MHz, C₆D₆, 298 K): δ 34.1 (s). $^{13}C\{^{1}H\}$ NMR plus HMBC (75 MHz, toluene-d₈, 353 K): δ 177.0 (br, CO-Os), 154.5 (s, CO), 142.3 (s, =CH), 98.9 (s, =CH), 89.6 (s, NCH), 88.5 (s, CHCH₂), 72.5 (s, CHOH), 63.3 (s, CH₂OH), 41.6 (s, CH₂), 27.8 (vt, N = 23.6, PCH(CH₃)₂), 20.8, 20.7 (both s, PCH(CH₃)₂).

Reaction of $OsH_6(P^iPr_3)_2$ with Uridine: Preparation of 7. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of uridine (47.2 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. The subsequent addition of cold pentane to the residue afforded a yellow solid. Yield: 70 mg (48%). Anal. Calcd for C₂₇H₅₆N₂O₆OsP₂: C, 42.84; H, 7.46; N, 3.70. Found: C, 43.17; H, 7.79; N, 3.82. ESI-HRMS (m/z): calcd for $C_{27}H_{55}N_2O_6OsP_2$ [M - H]⁺ 757.3146; found 757.3193. IR (neat compound, cm⁻¹): ν (OH) 3368 (br); ν (OsH) 2126 (w); ν (C=O) 1641 (s), 1608 (s); ν (C=C) 1534 (s). ¹H NMR (400 MHz, C₆D₆, 298 K): δ 7.75 (d, J_{H-H} = 7.6, 1H, =CH), 6.70 (br, 1H, OH), 5.98 (d, $J_{H-H} = 5.1, 1H, NCH), 5.65$ (br, 1H, OH), 5.30 (d, $J_{H-H} = 7.6, 1H,$ =CH), 4.66 (dd, J_{H-H} = 6.1, J_{H-H} = 4.7, 1H, CHOH), 4.62 (dd, J_{H-H} = 5.1, J_{H-H} = 4.7, 1H, CHOH), 4.31 (m, 1H, CH₂CH), 3.88 (d, J_{H-H} = 11.2, 1H, CH₂OH), 3.77 (d, J_{H-H} = 11.2, 1H, CH₂OH), 3.46 (br, OH), 1.99 (m, 6H, PCH(CH₃)₂), 1.17 (dvt, $J_{H-H} = 7.2$, N = 13.2, 18H, PCH(CH₃)₂), 1.16 (dvt, $J_{H-H} = 6.0$, N = 12.0, 18H, PCH(CH₃)₂), -13.21 (br, 3H, OsH). ¹H{³¹P} NMR (300 MHz, toluene- d_8 , 213 K, high-field region): δ –10.56 (br, 1H, OsH), –13.59 (br, 1H, OsH), -15.28 (br, 1H, OsH). ${}^{31}P{}^{1}H{}$ NMR (162 MHz, C₆D₆, 298 K): δ 34.1 (s). ${}^{13}C{}^{1}H{}$ NMR plus HMBC (101 MHz, C_6D_{62} 298 K): δ 176.9 (br, CO–Os), 155.1 (s, CO), 142.7 (s, =CH),

98.3 (s, =CH), 94.6 (s, NCH), 88.1 (s, CHCH₂), 76.0 (s, CHOH), 70.6 (s, CHOH), 61.9 (s, CH₂OH), 27.1 (vt, N = 23.6, PCH(CH₃)₂), 20.3, 20.2 (both s, PCH(CH₃)₂).

Reaction of Complex 5 with $OsH_2Cl_2(P^iPr_3)_2$ (8) in the Presence of NEt3: Preparation of Complex 9. A solution of complex 5 (100.0 mg, 1.3×10^{-4} mmol) in toluene (6 mL) was treated with the stoichiometric amount of 8 (75.7 mg, 1.3×10^{-4} mmol) and 4.0 equiv of NEt₃ (75.0 μ L, 5.6 × 10⁻⁴ mmol) during 20 min at room temperature. During this time the color changed from brown to orange. The resulting suspension was filtered through Celite to remove [HNEt₃]Cl and the solution thus obtained concentrated in vacuo to afford a yellow solid. Yield: 98.0 mg (59%). Anal. Calcd for C46H100N2O6Os2P4: C, 43.11; H, 7.86; N, 2.18. Found: C, 42.70; H, 7.82; N, 2.07. ESI-HRMS (m/z): calcd for $C_{46}H_{101}N_2O_6Os_2P_4$ [M + H]⁺ 1283.5814; found 1283.5936. IR (neat compound, cm⁻¹): ν (OH) 3358 (br), ν (OsH); 2155 (m), ν (C=O) 1663 (s), ν (C=C) 1523 (s). ¹H NMR (500 MHz, $C_6 D_{62}$ 298 K): δ 7.15 (s, 1H, =CH), 6.00 (d, $J_{\rm H-H} = 3.0, 1$ H, NCH), 5.01 (m, 1H, CHO–Os), 4.92 (dd, $J_{\rm H-H} = 5.4,$ $J_{\rm H-H}$ = 5.4, 1H, CHCH₂OH), 4.25 (m, 1H, CHO–Os), 4.10 (d, $J_{\rm H-H}$ = 11.5, 1H, CH₂OH), 3.94 (m, 1H, CH₂OH), 3.38 (br, 1H, OH), 2.12 (m, 6H, PCH(CH₃)₂), 2.04 (m, 3H, PCH(CH₃)₂), 1.98 (m, 3H, $PCH(CH_3)_2$, 1.75 (s, 3H, CH₃), 1.26 (dvt, $J_{H-H} = 6.5$, N = 13, 9H, $PCH(CH_3)_2$), 1.24 (dvt, J_{H-H} = 6.5, N = 12.5, 9H, $PCH(CH_3)_2$), 1.19 $(dvt, J_{H-H} = 7, N = 12.5, 9H, PCH(CH_3)_2), 1.18 (dvt, J_{H-H} = 7, N = 7)$ 12.5, 9H, PCH(CH₃)₂), 1.14 (dd, $J_{H-H} = 7$, $J_{H-P} = 12.5$, 36H, PCH(CH₃)₂), -13.17 (br, 3H, OsH), -17.63 (t, $J_{H-P} = 41$, 2H, OsH). ³¹P{¹H} NMR (162 MHz, C₆D₆, 298 K): δ 34.0 (s), 33.7 (s). ¹³C{¹H} NMR plus HMBC (75.4 MHz, C₆D₆, 298 K): δ 176.5 (s, CO-Os), 153.9 (s, CO), 140.5 (s, =CH), 105.8 (s, =CCH₃), 100.1 (s, NCH), 92.6 (s, CHO-Os), 90.9 (s, CHO-Os), 87.7 (s, CHCH2), 63.8 (s, CH₂OH), 27.4 (d, J_{C-P} = 31.4, PCH(CH₃)₂), 27.2 (vt, N = 23.4, PCH(CH₃)₂), 27.1 (vt, N = 23.4, PCH(CH₃)₂), 20.5, 20.4, 20.3, 20.2, 19.6, 19.5 (all s, PCH(CH₃)₂), 10.8 (s, CH₃).

Reaction of Complex 7 with OsH₂Cl₂(PⁱPr₃)₂ (8) in the Presence of NEt₃: Preparation of Complex 10. A solution of complex 7 (100 mg, 1.3×10^{-4} mmol) in toluene (6 mL) was treated at room temperature with a stoichiometric amount of 8 (77.1 mg, 1.3 \times 10⁻⁴ mmol) and 4.0 equiv of NEt₃ (73.0 µL, 5.6 \times 10⁻⁴ mmol) during 20 min. During this time the solution changed from brown to orange. The resulting suspension was filtered through Celite to remove [HNEt₃]Cl and the solution thus obtained concentrated in vacuo to afford a yellow solid. Yield: 106.6 mg (64%). Anal. Calcd for C45H98N2O6OS2P4: C, 42.64; H, 7.79; N, 2.21. Found: C, 42.40; H, 7.85; N, 2.09. ESI-HRMS (m/z): calcd for C₄₅H₉₉N₂O₆Os₂P₄ [M + H]⁺: 1269.5657; found: 1269.5683. IR (neat compound, cm⁻¹): ν (OH) 3348 (br), ν (OsH); 2156 (m), 2134 (m), ν (C=O) 1649 (s), 1610 (s), ν (C=C) 1540 (s). ¹H NMR (500 MHz, C₆D₆, 298 K): δ 7.46 (d, $J_{\rm H-H}$ = 7.0, 1H, =CH), 6.00 (d, $J_{\rm H-H}$ = 3, 1H, NCH), 5.17 (d, $J_{\rm H-H}$ = 7.0, 1H, =CH), 4.93 (m, 1H, CHO–Os), 4.80 (dd, $J_{\rm H-H}$ = 5.5, J_{H-H} = 2.5, 1H, CHCH₂OH), 4.21 (m, 1H, CHO–Os), 4.04 (dd, J_{H-H} = 12, J_{H-H} = 2.5, 1H, CH₂OH), 3.87 (d, J_{H-H} = 12, 1H, CH₂OH), 3.07 (br, 1H, OH), 2.12 (m, 6H, PCH(CH₃)₂, 2.05 (m, 3H, PCH(CH₃)₂), 1.99 (m, 3H, PCH(CH₃)₂), 1.27 (dvt, $J_{H-H} = 6.5$, N = 12.5, 9H, $PCH(CH_3)_2)$, 1.24 (dvt, $J_{H-H} = 7$, N = 13, 9H, $PCH(CH_3)_2)$, 1.20 13, 9H, PCH(CH₃)₂), 1.13 (dd, $J_{H-H} = 7$, $J_{H-P} = 12.5$, 36H, PCH(CH₃)₂), -13.16 (br, 3H, OsH), -17.68 (t, $J_{H-P} = 41$, 2H, OsH). ³¹P{¹H} NMR (162 MHz, C₆D₆, 298 K): δ 34.4 (s), 33.6 (s). ¹³C{¹H} NMR plus HMBC (101 MHz, C₆D₆, 298 K): δ 176.8 (s, CO-Os), 153.7 (s, CO), 143.3 (s, =CH), 98.9 (s, NCH), 97.7 (s, =CH), 93.2 (s, CHO-Os), 90.9 (s, CHO-Os), 87.2 (s, CHCH₂), 63.5 (s, CH₂OH), 27.4 (d, J_{C-P} = 33, PCH(CH₃)₂), 27.1 (vt, N = 20.2, PCH(CH₃)₂), 20.5, 20.4, 19.6, 19.5 (all s, PCH(CH₃)₂).

Reaction of 2 with OsH_6(P^iPr_3)_2: Preparation of 11. A yellow solution of **2** (63 mg, 0.096 mmol) in toluene (6 mL) was treated with 1.0 equiv of $OsH_6(P^iPr_3)_2$ (50 mg, 0.096 mmol) and heated under reflux. Periodically, aliquots were removed and checked by ${}^{31}P{}^{1}H{}$ NMR spectroscopy to follow the reaction. After 8.5 h the complete conversion of **2** in **11** is observed. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. Addition of

methanol to the residue afforded a white solid that was washed with methanol and dried in vacuo. Yield: 71.8 mg (65%). Anal. Calcd for C41H94N2O2Os2P4: C, 42.76; H, 8.23; N, 2.43; found: C, 42.43; H, 8.48; N, 2.71. ESI-HRMS (m/z): calcd for C₄₁H₉₃N₂O₂Os₂P₄ [M -H]⁺ 1151.5390; found 1151.5437. IR (neat compound, cm⁻¹): ν (Os-H) 2149 (m), 2118 (m); ν (C=O) 1599 (s), 1542 (s); ν (C=C) 1505 (s). ¹H NMR (300 MHz, C_6D_{62} 298 K): δ 7.57 (s, 1H, =CH), 2.01 (m, 12H, PCH(CH₃)₂), 1.96 (s, 3H, CH₃), 1.35 (dvt, J_{H-H} = 6.6, N = 13.2, 18H, PCH(CH₃)₂), 1.31 (dvt, $J_{H-H} = 6.9$, N = 13.5, 18H, PCH(CH₃)₂), 1.13 (dvt, $J_{H-H} = 6.0$, N = 11.7, 18H, PCH(CH₃)₂), 1.05 (dvt, $J_{H-H} = 6.6$, N = 12.3, 18H, PCH(CH₃)₂), -13.50 (very br, 6H, OsH). $^{1}H{}^{31}P$ NMR (400 MHz, toluene- d_{8} , 193 K, high-field region): δ –9.98 (d, 1H, J_{H-H} = 9.6, Os–H), –12.00 (br, 1H, Os–H), -13.38 (br, 1H, Os-H), -13.54 (t, $J_{H-H} = 13.6$, 1H, Os-H), -13.97(br, 1H, Os–H), -15.97 (d, J_{H-H} = 13.6, 1H, Os–H). ³¹P{¹H} NMR (161.9 MHz, C_6D_6 , 298 K): δ 36.9 (s), 30.9 (s). ¹³C{¹H} NMR plus HSQC and HMBC (75 MHz, C_6D_6 , 298 K): δ 176.9 (t, $J_{C-P} = 1.5$, CO), 165.9 (t, $J_{C-P} = 1.5$, CO), 152.5 (s, =CH), 105.7 (s, =CCH₃), 28.1 (vt, N = 23.2, PCH(CH₃)₂), 27.1 (vt, N = 23.2, PCH(CH₃)₂), 21.5, 21.1, 19.9, 19.6 (all s, PCH(CH₃)₂), 11.4 (s, CH₃).

Reaction of OsH_6(P^iPr_3)_2 with 0.5 Equiv of Thymine: Preparation of Complex 11. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 0.5 equiv of thymine (12.2 mg, 0.096 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. ¹H and ³¹P{¹H} NMR spectra show the presence of complexes 13 and 11 in a ratio 0.05:1. Addition of methanol to the residue afforded a white solid (complex 11) that was washed with methanol and dried in vacuo. Yield: 74 mg (66%).

Reaction of 3 with OsH₆(PⁱPr₃)₂: Preparation of 12. A yellow solution of 3 (61.3 mg, 0.096 mmol) in toluene (6 mL) was treated with 1.0 equiv of $OsH_6(P^iPr_3)_2$ (50 mg, 0.096 mmol) and heated under reflux. Periodically, aliquots were removed and checked by ³¹P{¹H} NMR spectroscopy to follow the reaction. After 2 h the complete conversion of 3 in 12 is observed. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. Addition of methanol to the residue afforded a white solid that was washed with methanol and dried in vacuo. Yield: 76.3 mg (70%). Anal. Calcd for C40H92N2O2Os2P4: C, 42.23; H, 8.15; N, 2.46. Found: C, 41.93; H, 8.04; N, 2.24. ESI-HRMS (m/z): calcd for $C_{40}H_{91}N_2O_2Os_2P_4 [M - H]^+$ 1137.5234; found 1137.5230. IR (neat compound, cm⁻¹): ν (Os-H) 2146 (w), 2123 (m); ν (C=O) 1564 (s), 1553 (s); ν (C=C) 1501 (s). ¹H NMR (300 MHz, C₆D₆, 298 K): δ 7.55 (d, $J_{\rm H-H}$ = 6.0, 1H, CH), 5.47 (d, $J_{\rm H-H}$ = 6.0, 1H, CH), 2.04 (m, 6H, PCH(CH₃)₂), 1.93 (m, 6H, PCH(CH₃)₂), 1.37 (dvt, $J_{H-H} = 6.9$, $N = 13.2, 18H, PCH(CH_3)_2$, 1.29 (dvt, $J_{H-H} = 6.9, N = 13.2, 18H$, PCH(*CH*₃)₂), 1.17 (dvt, $J_{H-H} = 6.6$, N = 12.0, 18H, PCH(*CH*₃)₂), 1.04 (dvt, $J_{H-H} = 6.6$, N = 11.7, 18H, PCH(*CH*₃)₂), -13.48 (br, 6H, OsH). ¹H{³¹P} NMR (400 MHz, toluene- d_8 , 193 K, high-field region): δ -9.96 (br, 1H, Os-H), -11.99 (br, 1H, Os-H), -13.36 (br, 1H, Os-H), -13.55 (br, 1H, Os-H), -14.00 (br, 1H, Os-H), -16.10 (br, 1H, Os-H). ³¹P{¹H} NMR (161.9 MHz, C₆D₆, 298 K): δ 36.7 (s), 30.9 (s). ${}^{13}C{}^{1}H$ NMR (101 MHz, tol-d₈, 298 K): δ 178.8 (s, CO), 167.4 (s, CO), 155.4 (s, =CH), 99.4 (s, =CH), 29.2 (vt, N = 23.9, PCH(CH₃)₂), 28.1 (vt, N = 23.4, PCH(CH₃)₂) 22.4, 22.1, 20.9, 20.7 (all s, $PCH(CH_3)_2$)

Reaction of $OsH_6(P^iPr_3)_2$ with 0.5 Equiv of Uracil: Preparation of Complex 12. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 0.5 equiv of uracil (10.8 mg, 0.096 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. ¹H and ³¹P{¹H} NMR spectra show the presence of complexes 14 and 12 in a ratio 0.21:1. Addition of methanol to the residue afforded a white solid (complex 12) that was washed with methanol and dried in vacuo. Yield: 134 mg (61%).

Reaction of $OsH_6(P^iPr_3)_2$ with 1.0 Equiv of Thymine: Preparation of 13. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of thymine (24.4 mg, 0.193 mmol) and heated under reflux during 3 h. During this time the solution changed from colorless to pale yellow. The resulting solution was filtered through Celite, and the solvent was removed in vacuo. $^1\!H$ and $^{31}P\{^1\!H\}$ NMR spectra show the presence of 13 and 11 in a ratio 1: 0.2. Addition of methanol to the residue afforded a white solid (complex 11) that was washed with methanol and dried in vacuo. The methanol solution was recollected and was taken to dryness. The subsequent addition of cold pentane to the residue afforded a white solid (complex 13). Yield: 81 mg (66%). Complex 13: Anal. Calcd for C₂₃H₅₀N₂O₂OsP₂: C, 43.24; H, 7.89; N, 4.38. Found: C, 42.89; H, 8.03; N, 4.55. ESI-HRMS (m/z): calcd for C₂₃H₄₉N₂O₂OsP₂ [M – H]⁺: 639.2879; found: 639.2912. IR (neat compound, cm⁻¹): ν (Os-H) 2133 (m); ν (C=O) 1661 (s), 1614 (s); ν (C=C) 1514 (s). ¹H NMR (400 MHz, C₆D₆, 298 K): δ 12.46 (s, 1H, NH), 6.51 (s, 1H, =CH), 1.98 (m, 6H, PCH(CH₃)₂), 1.64 (s, 3H, CH₃), 1.22 (dvt, J_{H-H} = 7.2, N = 14.0, 18H, PCH(CH_3)₂), 1.20 (dvt, J_{H-H} = 7.2, N = 13.6, 18H, PCH(CH_3)₂), -13.14 (br, 3H, OsH). ¹H(³¹P) NMR (400 MHz, toluene- d_8 , 203 K, high-field region): δ -10.53 (br, 1H, Os-H), -13.52 (br, 1H, Os-H), -15.05 (br, 1H, Os-H). $^{31}P{^{1}H}$ NMR (161.9 MHz, C_6D_{61} 298 K): δ 33.9 (s). ¹³C{¹H} NMR (75 MHz, C_6D_{67} 298 K): δ 178.0 (t, J_{C-P} = 1.6, CO), 158.0 (s, CO), 138.9 (s, =CH), 105.2 (s, =CCH₃), 27.3 (vt, N = 23.7, PCH(CH₃)₂), 20.5, 20.2 (both s, $PCH(CH_3)_2$), 10.4 (s, CH_3).

Reaction of OsH₆(PⁱPr₃)₂ with 1.0 Equiv of Uracil: Preparation of 14. A colorless solution of 1 (100 mg, 0.193 mmol) in toluene (10 mL) was treated with 1.0 equiv of uracil (21.7 mg, 0.193 mmol) and heated under reflux during 3 h, changing the color of the solution from colorless to pale yellow. After this time the mixture was cooled at room temperature and filtered through Celite, and the solvent was removed in vacuo. ¹H and ³¹P{¹H} NMR spectra showed quantitative conversion to a 1:0.2 mixture of complexes 14 and 12. Extraction of the resulting residue with methanol allows the isolation of both complexes in pure form: Complex 14 is extracted with methanol (10 mL), while complex 12 remains in the residue. Addition of methanol to the residue afforded a white solid (12) that was washed with methanol and dried in vacuo. The methanol solution containing complex 14 was taken to dryness. Subsequent addition of cold pentane to the resulting residue afforded a white solid (complex 14). Yield: complex 14: 73 mg (61%), complex 12: 16 mg (15%). Complex 14: Anal. Calcd for C₂₂H₄₈N₂O₂OsP₂: C, 42.29; H, 7.74; N, 4.48; found: C, 42.56; H, 7.82; N, 4.29. ESI-HRMS (m/z): calcd for $C_{22}H_{47}N_2O_2O_3P_2 \ [M - H]^+ \ 625.2723;$ found 625.2728. IR (neat compound, cm⁻¹): ν (OsH) 2122 (w); ν (C=O) 1659 (m), 1613 (m); ν (C=C) 1527 (s). ⁱH NMR (300 MHz, C₆D₆, 298 K): δ 12.14 (s, 1H, NH), 6.64 (d, J_{H-H} = 7.2, 1H, =CH), 5.02 (d, J_{H-H} = 7.2, 1H, =CH), 1.99 (m, 6H, PCH(CH₃)₂), 1.22 (dvt, J_{H-H} = 6.9, N = 12.9, 18H, PCH(CH₃)₂), 1.19 (dvt, $J_{H-H} = 6.9$, N = 12.6, 18 H, PCH(CH₃)₂, -13.13 (br, 3H, OsH). ¹H{³¹P} NMR (400 MHz, toluene- d_{8} , 203 K, high-field region): δ -10.51 (br, 1H, Os-H), -13.56 (br, 1H, Os-H), -15.12 (br, 1H, Os-H). ³¹P{¹H} NMR (161.9 MHz, C₆D₆, 298 K): δ 34.1 (s). ¹³C{¹H} NMR (101 MHz, C_6D_6 , 298 K): δ 178.1 (t, J_{C-P} = 1.6, CO), 157.8 (s, CO), 141.9 (s, =CH), 97.2 (s, =CH), 27.2 (vt, N = 24.0, PCH(CH₃)₂), 20.3 (s, $PCH(CH_3)_2$

Structural Analysis of Complexes 6, 11, and 14. Crystals suitable for the X-ray diffraction were obtained by slow diffusion of methanol into solutions of the complexes in toluene. X-ray data were collected on a Bruker Smart APEX (11, 14) and Bruker Apex II CCD (6) difractometers equipped with a normal focus, 2.4 kW sealed tube source (Mo radiation, $\lambda = 0.71073$ Å) operating at 50 kV and 30 (6, 11) or 40 (14) mA. Data were collected over the complete sphere by a combination of four sets. Each frame exposure time was 10 s (20 s for 6 and 14) covering 0.3° in ω . Data were corrected for absorption by The using a multiscan method applied with the SADABS program.¹⁵ structures were solved by the Patterson (Os atoms of 6, 11, and 14) method and conventional Fourier techniques and refined by full-matrix least-squares on F² with SHELXL97.²⁰ Anisotropic parameters were used in the last cycles of refinement for all non-hydrogen atoms. The hydrogen atoms were observed or calculated and refined freely or using a restricted riding model. Hydride ligands were observed in the difference Fourier maps but refined with restrained Os–H bond length (1.59(1) Å, CSD). The disordered phosphine groups observed in the structure of complex 14 were refined with two moieties, complementary occupancy factors, and isotropic thermal parameters. For all structures the highest electronic residuals were observed in the close proximity of the Os centers and make no chemical sense.

Crystal data for 6: $C_{27}H_{56}N_2O_5OsP_2$, M_W 740.88, colorless, irregular block (0.15 × 0.08 × 0.05), orthorhombic, space group $P2_12_12_1$, *a*: 11.0603(15) Å, *b*: 16.493(2) Å, *c*: 17.632(2) Å, *V* = 3216.3(8) Å³, *Z* = 4, D_{calc} : 1.530 g cm⁻³, F(000): 1512, T = 100(2) K, μ 4.100 mm⁻¹. 35436 measured reflections (2θ : 3–58°, ω scans 0.3°), 8389 unique (R_{int} = 0.0335); minimum/maximum transmission factors 0.645/ 0.862. Final agreement factors were R^1 = 0.0200 (8048 observed reflections, I > $2\sigma(I)$) and wR² = 0.0447; data/restraints/parameters 8389/3/359; GoF = 1.009. Largest peak and hole 1.080 and -0.391 e/ Å³.

Crystal data for **11**: $C_{41}H_{94}N_2O_2O_5{}_2P_4$, M_W 1151.46, colorless, prism (0.18 × 0.16 × 0.08), monoclinic, space group P2(1)/n, *a*: 14.6768(7) Å, *b*: 16.0063(7) Å, *c*: 21.5187(10) Å, *α*: 90.00°, *β*: 99.2180(10)°, γ : 90.00°, *V* = 4989.9(4) Å³, *Z* = 4, D_{calc} : 1.533 g cm⁻³, F(000): 2328, T = 100(2) K, μ 5.249 mm⁻¹. 59788 measured reflections (2 θ : 3–58°, ω scans 0.3°), 12026 unique (R_{int} = 0.0380); minimum/maximum transmission factors 0.445/0.589. Final agreement factors were R¹ = 0.0266 (9745 observed reflections, I > 2 σ (I)) and wR² = 0.0566; data/restraints/parameters 12026/6/509; GoF = 0.984. Largest peak and hole 1.584 and -1.619 e/Å³.

Crystal data for 14: $C_{22}H_{48}N_2O_2OsP_2$, M_W 624.76, colorless, irregular block (0.10 × 0.06 × 0.03), monoclinic, space group P2(1)/*c*, *a*: 17.7824(17) Å, *b*: 20.108(2) Å, *c*: 30.682(3) Å, *α*: 90.00°, *β*: 96.9740(10)°, γ : 90.00°, *V* = 10889.8(18) Å³, *Z* = 16, D_{calc} : 1.524 g cm⁻³, F(000): 5056, T = 100(2) K, μ 4.820 mm⁻¹. 131407 measured reflections (2 θ : 3–58°, ω scans 0.3°), 26270 unique (R_{int} = 0.1160); minimum/maximum transmission factors 0.514/0.799. Final agreement factors were R¹ = 0.0420 (12537 observed reflections, I > 2 σ (I)) and wR² = 0.0737; data/restraints/parameters 26270/64/1122; GoF = 0.774. Largest peak and hole 2.069 and -1.844 e/Å³.

ASSOCIATED CONTENT

S Supporting Information

CIF files giving positional and displacement parameters, crystallographic data, and bond lengths and angles of compounds 6, 11, and 14. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*E-mail: maester@unizar.es.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

Financial support from the MINECO of Spain (Projects CTQ2011-23459 and Consolider Ingenio 2010 CSD2007-00006), the Diputación General de Aragón (E35), and FEDER is acknowledged.

REFERENCES

(1) (a) Jamieson, E. R.; Lippard, S. J. Chem. Rev. 1999, 99, 2467.
(b) Lippert, B. Coord. Chem. Rev. 2000, 200-202, 487. (c) Jung, Y.; Lippard, S. Chem. Rev. 2007, 107, 1387.

(2) (a) Ronconi, L.; Sadler, P. J. Coord. Chem. Rev. 2007, 251, 1633.
(b) Lynam, J. M. Dalton Trans. 2008, 4067. (c) Pizarro, A. M.; Sadler, P. J. Biochimie 2009, 91, 1198. (d) Sava, G.; Bergamo, A.; Dyson, P. J. Dalton Trans. 2011, 40, 9069.

(3) (a) Dorcier, A.; Ang, W. H.; Bolaño, S.; Gonsalvi, L.; Juillerat-Jeannerat, L.; Laurenczy, G.; Peruzzini, M.; Phillips, A. D.; Zanobini, F.; Dyson, P. J. *Organometallics* **2006**, *25*, 4090. (b) Schmid, W. F.; John, R. O.; Arion, V. B.; Jakupec, M. A.; Keppler, B. K. Organometallics 2007, 26, 6643. (c) Peng, H.; Zhang, L.; Kjällman, T. H. M.; Soeller, C.; Travas-Sejdic, J. J. Am. Chem. Soc. 2007, 129, 3048. (d) Peacock, A. F. A.; Sadler, P. J. Chem Asian J. 2008, 3, 1890. (e) Kostrhunova, H.; Florian, J.; Novakova, O.; Peacock, A. F. A.; Sadler, P. J.; Brabec, V. J. Med. Chem. 2008, 51, 3635. (f) van Rijt, S. H.; Hebden, A. J.; Amaresekera, T.; Deeth, R. J.; Clarkson, G. J.; Parsons, S.; McGowan, P. C.; Sadler, P. J. J. Med. Chem. 2009, 52, 7753. (g) van Rijt, S. H.; Peacock, A. F. A.; Jhonstone, R. D. L.; Parsons, S.; Sadler, P. J. Inorg. Chem. 2009, 48, 1753. (h) Bergamo, A.; Masi, A.; Peacock, A. F. A.; Habtemariam, A.; Sadler, P. J.; Sava, G. J. Inorg. Biochem. 2010, 104, 79. (i) van Rijt, S. H.; Mukherjee, A.; Pizarro, A. M.; Sadler, P. J. J. Med. Chem. 2010, 53, 840. (j) Fu, Y.; Habtemariam, A.; Pizarro, A. M.; van Rijt, S. H.; Healey, D. J.; Cooper, P. A.; Shnyder, S. D.; Clarkson, G. J.; Sadler, P. J. J. Med. Chem. 2010, 53, 8192. (k) van Rijt, S. H.; Kostrhunova, H.; Brabec, V.; Sadler, P. J. Bioconjugate Chem. 2011, 22, 218. (l) Shnyder, S. D.; Fu, Y.; Habtemariam, A.; van Rijt, S. H.; Cooper, P. A.; Loadman, P. M.; Sadler, P. J. Med. Chem. Commun. 2011, 2, 666. (m) Fu, Y.; Habtemariam, A.; Basri, A. M. B. H.; Braddick, D.; Clarkson, G. J.; Sadler, P. J. Dalton Trans. 2011, 40, 10553.

(4) Brabec, V.; Nováková, O. *Drug Resist. Updates* **2006**, *9*, 111. (5) Atwood, J. D. *Inorganic and Organometallic Reaction Mechanisms*; Wiley-VCH Publishers: New York, 1997; Chapter 3.

(6) Ni, W.-X.; Man, W.-L.; Cheung, M. T.-W.; Sun, R. W.-Y.; Shu, Y.-L.; Lau, Y.-W.; Che, C.-M.; Lan, T.-C. Chem. Commun. 2011, 47, 2140. (7) (a) Barea, G.; Esteruelas, M. A.; Lledós, A.; López, A. M.; Oñate, E.; Tolosa, J. I. Organometallics 1998, 17, 4065. (b) Barrio, P.; Castarlenas, R.; Esteruelas, M. A.; Lledós, A.; Maseras, F.; Oñate, E.; Tomàs, J. Organometallics 2001, 20, 442. (c) Barrio, P.; Castarlenas, R.; Esteruelas, M. A.; Oñate, E. Organometallics 2001, 20, 2635. (d) Barrio, P.; Esteruelas, M. A.; Oñate, E. Organometallics 2004, 23, 1340. (e) Barrio, P.; Esteruelas, M. A.; Oñate, E. Organometallics 2004, 23, 3627. (f) Baya, M.; Eguillor, B.; Esteruelas, M. A.; Lledós, A.; Oliván, M.; Oñate, E. Organometallics 2007, 26, 5140. (g) Baya, M.; Eguillor, B.; Esteruelas, M. A.; Oliván, M.; Oñate, E. Organometallics 2007, 26, 6556. (h) Eguillor, B.; Esteruelas, M. A.; Oliván, M.; Puerta, M. Organometallics 2008, 27, 445. (i) Esteruelas, M. A.; Masamunt, A. B.; Oliván, M.; Oñate, E.; Valencia, M. J. Am. Chem. Soc. 2008, 130, 11612. (j) Esteruelas, M. A.; Forcén, E.; Oliván, M.; Oñate, E. Organometallics 2008, 27, 6188. (k) Eguillor, B.; Esteruelas, M. A.; García-Raboso, J.; Oliván, M.; Oñate, E. Organometallics 2009, 28, 3700. (1) Esteruelas, M. A.; Fernández, I.; Herrera, A.; Martín-Ortiz, M.; Martínez-Alvarez, R.; Oliván, M.; Oñate, E.; Sierra, M. A.; Valencia, M. Organometallics 2010, 29, 976. (m) Eguillor, B.; Esteruelas, M. A.; García-Raboso, J.; Oliván, M.; Oñate, E.; Pastor, I. M.; Peñafiel, I.; Yus, M. Organometallics 2011, 30, 1658.

(8) (a) Atencio, R.; Esteruelas, M. A.; Lahoz, F. J.; Oro, L. A.; Ruiz, N. Inorg. Chem. 1995, 34, 1004. (b) Esteruelas, M. A.; Lahoz, F. J.; López, A. M.; Oñate, E.; Oro, L. A.; Ruiz, N.; Sola, E.; Tolosa, J. I. Inorg. Chem. 1996, 35, 7811. (c) Castillo, A.; Barea, G.; Esteruelas, M. A.; Lahoz, F. J.; Lledós, A.; Maseras, F.; Modrego, J.; Oñate, E.; Oro, L. A.; Ruiz, N.; Sola, E. Inorg. Chem. 1999, 38, 1814. (d) Buil, M. L.; Esteruelas, M. A.; Garcés, K.; García-Raboso, J.; Oliván, M. Organometallics 2009, 28, 4606. (e) Esteruelas, M. A.; García-Raboso, J.; Oliván, M. Organometallics 2011, 30, 3844.

(9) (a) Engelking, H.; Krebs, B. J. Chem. Soc., Dalton Trans. 1996, 2409. (b) De Napoli, L.; Iacovino, R.; Messere, A.; Montesarchio, D.; Piccialli, G.; Romanelli, A.; Ruffo, F.; Saviano, M. J. Chem. Soc., Dalton Trans. 1999, 1945. (c) Fish, R. H.; Jaouen, G. Organometallics 2003, 22, 2166. (d) Romanelli, A.; Iacovino, R.; Piccialli, G.; Ruffo, F.; De Napoli, L.; Pedone, C.; Di Blasio, B.; Messere, A. Organometallics 2005, 24, 3401. (e) Ruiz, J.; Lorenzo, J.; Sanglas, L.; Cutillas, N.; Vicente, C.; Villa, M. D.; Aviles, F. X.; López, G.; Moreno, V.; Perez, J.; Bautista, D. Inorg. Chem. 2006, 45, 6347. (f) Futera, Z.; Klenko, J.; Šponer, J. E.; Šponer, J.; Burda, J. V. J. Comput. Chem. 2008, 30, 1758. (10) Ruiz, J.; Villa, M. D.; Rodríguez, V.; Cutillas, N.; Vicente, C.; López, G.; Bautista, D. Inorg. Chem. 2007, 46, 5448.

(11) Phosphine complexes of Ag(I) and Au(I) and phosphine ligands by themselves have been shown to be anticancer or antimitochondrial agents. See: Berners-Price, S. J.; Sadler, P. J. *Coord. Chem. Rev.* **1996**, *151*, 1.

(12) Metal complexes with pyrimidinic nucleoside ligands characterized by X-ray diffraction analysis are extremely rare. From 324 structures located in the Cambridge Structural Database, only 4 of them contain metals. See: (a) Guay, F.; Beauchamp, A. L. Can. J. Chem. 1985, 63, 3456. (b) Galy, J.; Mosset, A.; Grenthe, I.; Puigdomènech, I.; Sjöberg, B.; Hultén, F. J. Am. Chem. Soc. 1987, 109, 380. (c) Begum, N. S.; Manohar, H. Polyhedron 1992, 11, 2823. (d) Toma, P. H.; Toma, J. M. D. R.; Bergstrom, D. E. Acta Crystallogr, Sect. C: Cryst. Struct. Commun. 1993, 49, 2047.

(13) Barrio, P.; Esteruelas, M. A.; Lledós, A.; Oñate, E.; Tomàs, J. Organometallics 2004, 23, 3008.

(14) (a) Aracama, M.; Esteruelas, M. A.; Lahoz, F. J.; López, J. A.; Meyer, U.; Oro, L. A.; Werner, H. Inorg. Chem. 1991, 30, 288.
(b) Kuhlman, R.; Streib, W. E.; Huffman, J. C.; Caulton, K. G. J. Am. Chem. Soc. 1996, 118, 6934. (c) Wolf, J.; Stuer, W.; Grünwald, C.; Gevert, O.; Laubender, M.; Werner, H. Eur. J. Inorg. Chem. 1998, 1827.
(d) Battacharya, S.; Gupta, P.; Basuli, F. C.; Pierpont, G. Inorg. Chem. 2002, 41, 5810. (e) Baya, M.; Esteruelas, M. A.; Oñate, E. Organometallics 2011, 30, 4404.

(15) Gusev, D. G.; Kuhlman, R.; Rambo, J. R.; Berke, H.; Eisenstein, O.; Caulton, K. G. J. Am. Chem. Soc. **1995**, 117, 281.

(16) (a) Tayebani, M.; Gambarotta, S.; Yap, G. Organometallics 1998, 17, 3639. (b) Gandelman, M.; Milstein, D. Chem. Commun. 2000, 1603. (c) Cameron, T. M.; Abboud, K. A.; Boncella, J. M. Chem. Commun. 2001, 1224. (d) Ozerov, O. V.; Guo, C.; Papkov, V. A.; Foxman, B. M. J. Am. Chem. Soc. 2004, 126, 4792. (e) Fan, L.; Yang, L.; Guo, C.; Foxman, B. M.; Ozerov, O. V. Organometallics 2004, 23, 4778. (f) Ozerov, O. V.; Guo, C.; Fan, L.; Foxman, B. M. Organometallics 2004, 23, 5573. (g) Weng, W.; Guo, C.; Moura, C.; Yang, L.; Foxman, B. M.; Ozerov, O. V. Organometallics 2005, 24, 3487. (h) Burling, S.; Mahon, M. F.; Powell, R. E.; Whittlesey, M. K.; Williams, J. M. J. J. Am. Chem. Soc. 2006, 128, 13702. (i) Wang, X.; Chen, H.; Li, X. Organometallics 2007, 26, 4684. (j) Ye, J.; Zhang, X.; Chen, W.; Shimada, S. Organometallics 2008, 27, 4166. (k) Cabeza, J. A.; del Rio, I.; Miguel, D.; Sanchez-Vega, M. G. Angew. Chem., Int. Ed. 2008, 47, 1920. (I) Liu, L.; Wang, F.; Shi, M. Eur. J. Inorg. Chem. 2009, 1723. (m) Häller, L. J. L.; Page, M. J.; Erhardt, S.; Macgregor, S. A.; Mahon, M. F.; Naser, M. A.; Vélez, A.; Whittlesey, M. K. J. Am. Chem. Soc. 2010, 132, 18408. (n) Kundu, S.; Brennessel, W. W.; Jones, W. D. Inorg. Chim. Acta 2011, 379, 109. (o) Surawatanawong, P.; Ozerov, O. V. Organometallics 2011, 30, 2972.

(17) See for example: (a) Esteruelas, M. A.; Lledós, A.; Oliván, M.; Oñate, E.; Tajada, M. A.; Ujaque, G. Organometallics 2003, 22, 3753.
(b) Eguillor, B.; Esteruelas, M. A.; Oliván, M.; Oñate, E. Organometallics 2004, 23, 6015. (c) Eguillor, B.; Esteruelas, M. A.; Oliván, M.; Oñate, E. Organometallics 2005, 24, 1428. (d) Esteruelas, M. A.; Hernández, Y. A.; López, A. M.; Oliván, M.; Oñate, E. Organometallics 2005, 24, 5989. (e) Esteruelas, M. A.; Hernández, Y. A.; López, A. M.; Oliván, M.; Oñate, Y. A.; López, A. M.; Oliván, M.; Rubio, L. Organometallics 2008, 27, 799. (f) Castarlenas, R.; Esteruelas, M. A.; Oñate, E. Organometallics 2008, 27, 3240.

(18) Esteruelas, M. A.; Lahoz, F. J.; López, J. A.; Oro, L. A.;
Schlünken, C.; Valero, C.; Werner, H. Organometallics 1992, 11, 2034.
(19) Blessing, R. H. Acta Crystallogr. 1995, A51, 33. SADABS: Areadetector absorption correction; Bruker-AXS: Madison, WI, 1996.

(20) SHELXTL Package v. 6.10; Bruker-AXS: Madison, WI, 2000. Sheldrick, G. M. Acta Crystallogr. 2008, A64, 112.