

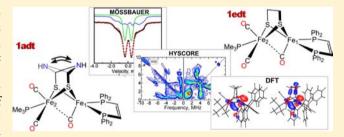
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EPR/ENDOR, Mössbauer, and Quantum-Chemical Investigations of Diiron Complexes Mimicking the Active Oxidized State of [FeFe]Hydrogenase

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Supporting Information

ABSTRACT: Understanding the catalytic process of the heterolytic splitting and formation of molecular hydrogen is one of the key topics for the development of a future hydrogen economy. With an interest in elucidating the enzymatic mechanism of the $[Fe_2(S_2C_2H_4NH)(CN)_2(CO)_2(\mu-CO)]$ active center uniquely found in [FeFe]hydrogenases, we present a detailed spectroscopic and theoretical analysis of its inorganic model $[Fe_2(S_2X)(CO)_3(dppv)(PMe_3)]^+$ [dppv =cis-1,2-bis(diphenylphosphino)ethylene] in two forms with S_2X = ethanedithiolate (1edt) and azadithiolate (1adt). These



complexes represent models for the oxidized mixed-valent Fe^IFe^{II} state analogous to the active oxidized "H_{ox}" state of the native H-cluster. For both complexes, the ³¹P hyperfine interactions were determined by pulse electron paramagnetic resonance and electron nuclear double resonance (ENDOR) methods. For 1edt, the ⁵⁷Fe parameters were measured by electron spin-echo envelope modulation and Mössbauer spectroscopy, while for 1adt, ¹⁴N and selected ¹H couplings could be obtained by ENDOR and hyperfine sublevel correlation spectroscopy. The spin density was found to be predominantly localized on the Fe(dppv) site. This spin distribution is different from that of the H-cluster, where both the spin and charge densities are delocalized over the two Fe centers. This difference is attributed to the influence of the "native" cubane subcluster that is lacking in the inorganic models. The degree and character of the unpaired spin delocalization was found to vary from 1edt, with an abiological dithiolate, to ladt, which features the authentic cofactor. For ladt, we find two 14N signals, which are indicative for two possible isomers of the azadithiolate, demonstrating its high flexibility. All interaction parameters were also evaluated through density functional theory calculations at various levels.

■ INTRODUCTION

The most efficient method of storing energy is within a chemical bond. Molecular dihydrogen is touted as a promising vessel for the storage of energy accumulated from renewable resources because of its low molecular weight and strong H–H single bond. ¹⁻³ Nature has engineered hydrogenase enzymes that promote the (reversible) heterolytic formation of dihydrogen from two protons and two electrons, i.e., 1,4,5

$$H_2 \leftrightarrows H^- + H^+ \leftrightarrows 2H^+ + 2e^- \tag{1}$$

Three classes of hydrogenases have been distinguished based on their active-site metal constituents: [FeFe], [NiFe], and [Fe].^{1,6}

The active center of [FeFe]hydrogenases, the "H-cluster", is a complex 6Fe cluster consisting of a "classical" four-cysteineligated [4Fe-4S] cluster appended to a catalytically active [2Fe] subcluster via a protein Cys-S ligand (see Figure 1).^{7,8} Both Fe centers are coordinated by CO and CN⁻ ligands that

stabilize low oxidation and spin states. 9,10 The two Fe ions are connected by a dithiolate bridge, whose central atom has sparked considerable debate because the available crystallographic data have insufficient resolution to discriminate O, C, or N. 11,12 Convincing evidence for N as a central atom has been obtained from an extensive pulse electron paramagnetic resonance (EPR) study of the H-cluster. 12 It has been proposed that this moiety might serve as a well-positioned base for relaying protons to and from the Fe center. 11-14

Two catalytically relevant states of the H-cluster have been experimentally identified thus far. ^{1,10,15} The active oxidized state, "H_{ox}", is characterized by a formal 2+ charge on the cubane subunit and a Fe^IFe^{II} configuration of the [2Fe] subcluster (Figure 1). This state is believed to be the entry point of the catalytic cycle. Whether this state directly

Received: June 27, 2012 Published: July 16, 2012

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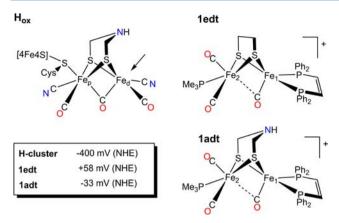


Figure 1. Schematic representation of the H-cluster in the H_{ox} state and model compounds used in this study. An arrow points to the vacant apical site on the distal Fe, which in the case of the enzyme is the site for binding H_2 . Inset: midpoint potentials for the H_{red}/H_{ox} and $[Fe_2(SX)_2]^{0/+}$ redox transition for the H-cluster and the models **1edt** and **1adt**, correspondingly. For the H-cluster, it is specified for pH 8^{15} in a Tris buffer with a KCl electrolyte; for the model compounds, the E_0 values are given for a $[(C_4H_9)_4N]PF_6$ electrolyte in CH_2Cl_2 and values are converted from versus $Fc^{0/+}$ to versus NHE scale by using $E_0(Fc^{0/+}) = +0.528$ versus NHE estimation.

Recently, we have provided evidence for another possible stable intermediate.²⁰ In this new state, [4Fe-4S] presumably exists in the reduced Fe^{II}₃Fe^{III} state. Because it allows one to store a second electron upon heterolytic splitting, it is likely that during the catalytic cycle the H-cluster passes through this state.

The catalytically pertinent [2Fe] subcluster can be viewed as a semiclassical diiron complex embedded in the protein pocket because only the Cys-S ligand of the cubane unit tethers the site covalently to the protein superstructure. Because the Fe₂ center is so lightly attached to the protein, it has proven to be a tempting target for synthetic modeling studies directed toward developing artificial hydrogen conversion systems.^{2,3} However, to date, only few of the published model compounds show significant conversion of protons to H₂, with the majority of cases requiring a substantial overpotential (i.e., the deviation of the midpoint potential from the ones observed for native systems).^{3,23}

These deficiencies of current synthetic models are likely to arise from differences in the electronic structure between these models and the active site. Specifically, we have shown that the electronic structure of the native [2Fe] cluster is strongly affected by the presence of the [4Fe–4S] subcluster;¹⁹ it is highly likely that this coupling is crucial for fine-tuning of the

electronic structure and thus the catalytic activity. Moreover, the pendant base, i.e., the bridging azadithiolate cofactor, might play an important role, acting as a relay for proton transfer. $^{1,12-14,22,24,25}$ Understanding these aspects will provide important insight into the mechanisms of $\rm H_2$ -activating catalysts, ultimately underpinning the development of efficient synthetic catalysts. To date, however, our understanding of the electronic structure of the catalytically active part of the H-cluster is rather limited. Model compounds mimicking the [2Fe] site provide a convenient means to study this part of the active center separately from the cubane moiety.

Recently, we conducted a study of $\{2\text{Fe3S}\}$ model compounds closely resembling the CO-inhibited oxidized state of the H-cluster, in which a CO ligand occupies the substrate binding site. The electronic structure of these models features a strong delocalization of the spin density over both Fe centers as well as some ligands. In contrast, our previous studies of the [FeFe]hydrogenase from *Desulfovibrio desulfuricans* indicated that the spin density is localized on the proximal Fe in the H_{ox} –CO state. This discrepancy was the first hint that the [4Fe–4S] subcluster significantly affects the electronic structure of the [2Fe] subcluster, despite the identical coordination spheres in the model and enzyme.

In this work, we present the first detailed EPR and Mössbauer studies to elucidate the ground-state electronic structures of biomimetic models resembling the dinuclear part of the active H_{ox} state of the H-cluster (Figure 1). Here we focus on the $[Fe_2(S_2X)(CO)_3(dppv)(PMe_3)]$ series of compounds, where dppv = cis-1,2-bis(diphenylphosphino)-ethylene. In this class, the bridging dithiolate ligand can be varied considerably $[X = (CH_2)_2, (CH_2)_2CH_2, (CH_2)_2NH, (CH_2)_2O, (CH_2)_2NCH_2C_6H_5]$. Although their redox processes are still far from the H_2/H^+ equilibrium potential (about -410 mV vs NHE at pH 7), they, nonetheless, provide an accessible Fe^IFe^{II} mixed-valent analogue of the "native" H_{ox} state.

Two variants are studied, namely, **1edt** $[X = (SCH_2)_2]$ and **1adt** $[X = (SCH_2)_2NH]$ (Figure 1). The structural properties of **1edt** have been well characterized in previous studies using various methods. ^{29–31} X-band continuous-wave (CW) EPR spectra, showing an almost axial signal with a distinct 1:2:1 pattern originating from two equivalent ³¹P nuclei of dppv, suggested that the unpaired spin was confined to Fe₁ (coordinated by the dppv ligand) and, hence, formulated as Fe₂^{II}Fe₁^I. However, this result differed markedly from the fully delocalized description of the "native" H_{ox} state of the H-cluster. ¹⁹ This difference is analogous to the one found for the H_{ox} -CO state and the corresponding {2Fe3S} model compounds. ^{26,27}

The EPR spectrum of **1adt** is quite similar to that of **1edt**, suggesting that their electronic structures are similar as well. The deviation in the principal g values must be a consequence of introducing an amine group into the dithiolate bridge. A similar picture emerges from the IR spectroscopic analysis of these compounds. The CO-stretching IR bands differ for the two compounds (**1edt**, $\nu_{\rm CO}$ = 2015, 1962, and 1887 cm⁻¹; **1adt**, $\nu_{\rm CO}$ = 2013, 1956, and 1901 cm⁻¹). 32

In this work, we examine the impact of the amine-functionalized bridge on the distribution of the spin density in the $Fe^{II}Fe^{I}$ state and therein elucidate the ground-state electronic structures of **1edt** and **1adt**. The aforementioned catalytic properties are then contrasted with the native enzyme whose electronic structure of the H_{ox} state has been previously examined by an extensive multifrequency EPR approach. ¹²

Additionally, the present study investigates the applicability of density functional theory (DFT) to these model compounds. Although DFT methods are widely used for the investigation of [2Fe] low-valent constructs, 33 their performance was not yet fully tested against EPR data. Recently, we have demonstrated that a hybrid functional in conjunction with moderately large basis sets afforded reasonable estimates of the experimental spin-Hamiltonian parameters of the H_{ox}-CO mimicks.^{26,27} However, the present case is somewhat different because of the presumably localized nature of the spin density. It remains unclear whether DFT is suitable to track the impact of nuanced modifications to the second coordination sphere. In this work, we use the obtained experimental data to address these points for the first time. The results of the theoretical studies were then also used to obtain further insight into the electronic properties of the studied models.

MATERIALS AND METHODS

EPR Spectroscopy. EPR measurements were performed on a setup based on Bruker ELEXSYS E 580 X- and Q-band spectrometers as described earlier. ^{19,28} For Q-band measurements, we used a slightly overcoupled cylindrical TE011 home-built resonator. ³⁴

W-band electron spin-echo envelope modulation (ESEEM) and hyperfine sublevel correlation spectroscopy (HYSCORE) measurements were performed on a Bruker ELEXSYS E 680 FT EPR W-band spectrometer with a helium-flow cryostat and a variable-temperature TeraFlex Bruker resonator. The 84 GHz internal local oscillator source used for the W-band up/down converting unit was modified to achieve improved phase stability. The W-band bridge provided 300 mW output power. This setup allowed 24 ns as the minimum length of the $\pi/2$ pulse using a critically coupled resonator.

The pulse sequences used in this work can be found in the Supporting Information (SI; Figure S1). Conventional Q-band pulse electron nuclear double resonance (ENDOR) spectra were obtained using stochastic data acquisition³⁵ with one shot per point with $t_{\rm mix}=5$ μ s. Variable-mixing-time (VMT) ENDOR experiments were performed using the standard Davies and Mims ENDOR sequences in linear scan mode, recording 20–50 shots per point. In all cases, the shot repetition time was set to achieve an unsaturated ESE signal.

Pulse EPR data were simulated using home-written routines utilizing a conventional spin-Hamiltonian approach in the high-field approximation as previously detailed. In all simulations, the orientation selection was calculated, accounting for the two strongly coupled $^{31}\mathrm{P}$ nuclei. The $^{14}\mathrm{N}$ quadrupole coupling constant was accounted for by the quadrupole coupling parameter (K) and asymmetry parameter (η):

$$K = 2P_z = \frac{e^2 qQ}{4I(2I - 1)h} = \frac{e^2 qQ}{4h}, \qquad \eta = \frac{P_x - P_y}{P_z}$$
 (2)

where P_{x} , P_{y} , and P_{z} are the principal components of the traceless quadrupole tensor $(|P_{x}| \leq |P_{y}| \leq |P_{z}|)$ and I is the nuclear spin.

Mössbauer Spectroscopy. Mössbauer spectra were recorded on an alternating constant-acceleration spectrometer. The minimum experimental line width was 0.24 mm/s (full width at half-maximum). The sample temperature was maintained constant in either an Oxford Variox or an Oxford Mössbauer-Spectromag cryostat. The latter is a split-pair conducting magnet system for applying fields of up to 8 T to the samples that can be kept at temperatures in the range 1.5–250 K. The field at the sample is perpendicular to the γ beam. Isomer shifts are quoted relative to metallic iron at 300 K. Mössbauer spectra were simulated with a home-written spin-Hamiltonian program based on the usual nuclear Hamiltonian formalism.

DFT Calculations. All calculations were performed using the ORCA~2.8 software package³⁷ within the spin-unrestricted formalism. Geometry optimizations utilized the BP86 functional^{38,39} in concert with Ahlrichs triple- ζ valence basis set with three sets of polarization functions (TZVPP) on all atoms including the H atoms.⁴⁰ The

Ahlrichs (2df,2pd) polarization functions were obtained from the TurboMole basis set library under ftp.chemie.unikarlsruhe.de/pub/basen. The resolution of identity was used with the standard TZV/J expansion basis, as implemented in *ORCA*.

Calculations of spin-Hamiltonian parameters were performed using the three-parameter Becke–Lee–Yang–Parr (B3LYP) functional 41,42 with the Wachters basis set 43 on the Fe atoms and the TZVPP basis set on all other atoms. The calculated 57 Fe hyperfine (HF) coupling constants were obtained including a second-order spin–orbit coupling contribution. The calculated 57 Fe isomer shifts (δ) have been calibrated to the applied settings following previously published procedures. 44,45 The calibration $\rho(0)$ values were obtained using the geometries presented by Römelt et al. 46 The correlation between the experimental isomer shifts δ (mm/s) 46 and calculated $\rho(0)$ (au $^{-3}$) follows the linear response $\delta=0.1181-0.3821[\rho(0)-11614]$, where 11614 au $^{-3}$ has been added for convenience (see the SI).

Single-point energy calculations were obtained from both geometry optimization and spin-Hamiltonian calculations. However, because the differences in energies between different conformers are consistent between different types of calculations, here we only present those from the latter ones.

Synthesis of 57 Fe-Labeled Compounds. $[{}^{57}Fe_2(S_2C_2H_4)(CO)_6]$. This compound was prepared according to literature methods using isotopically enriched anhydrous FeCl₂. ^{31,47} A solution of FeCl₂ (0.26 g, 2.0 mmol) in 45 mL of acetone was treated with 1 atm of CO for 10 min. To the resulting yellow solution was added NEt₃ (0.30 mL, 22.0 mmol), changing the color to blue/gray. To this mixture was added H₂S₂C₂H₄ (0.1 mL, 10.6 mmol), generating a dark-brown solution. The mixture was stirred for 15 min, at which point zinc powder (0.120 g, 1.9 mmol) was added. After 18 h, the red solution was filtered through a Celite plug and then dried in vacuo. The solid was reconstituted in CH₂Cl₂ and filtered through a silica plug. The solvent was stripped under vacuum and the residue extracted into hexane and chromatographed on a plug of silica with hexane as the eluent. Two bands were observed: the desired product as a large red band that eluted first and a small pink band that was discarded. A small amount of brown insoluble material remained at the top of the column. The product fraction was dried in vacuo, affording red crystals. Yield: 0.480 g (30%). This product was spectroscopically identical (¹H NMR and IR) with the nonenriched material.

 $[^{57}Fe_2(S_2C_2H_4)(CO)_4(dppv)]$. Following published methods,³⁰ a slurry of 38% enriched $[^{57}Fe_2(S_2C_2H_4)(CO)_6]$ (0.260 g, 7.2 mmol) and dppv (0.270 g, 6.8 mmol) in 10 mL of MeCN was treated with Me₃NO (0.065 g, 8.7 mmol), resulting in vigorous bubbling and a darkening of the solution. After 3 h, the supernatant was filtered off, and the remaining precipitate was repeatedly recrystallized from CH₂Cl₂ by the addition of five volumes of hexane until the supernatant was clear. Yield: 0.344 g (67%).

 $[^{57}Fe_2(S_2C_2H_4)(CO)_3(dppv)(PMe_3)]$. This compound was prepared according to the literature recipe. ⁴⁷ IR and ¹H NMR spectra of the product were identical with the nonenriched form. ³¹P NMR (CD₂Cl₂): 95 (d, J_{Fe-P} = 53 Hz), 18 (d, J_{Fe-P} = 47 Hz).

Sample Preparation. The synthesis of reduced versions of **1edt** and **1adt** followed published procedures with minor modifications. These compounds were dissolved in toluene to a final concentration of 2–4 mM, treated with 1 equiv of Fc(BArF₄), and stirred vigorously for 30 min under an inert atmosphere. Then the resulting material was transferred to quartz EPR tubes or Mössbauer Teflon caps and frozen in liquid nitrogen.

RESULTS

Figure 2 shows EPR spectra of **1edt** and **1adt**, obtained by two-pulse-ESE-detected EPR at Q band. Both samples show signals consistent with the presence of one dominant species. The [1:2:1] splitting patterns observed at X-band originate from two presumably equivalent ³¹P nuclei of the dppv ligand. This structure is washed out at Q band because of the increased line width; however, the principal g values are more clearly resolved. The **1edt** model shows a larger anisotropy and a smaller

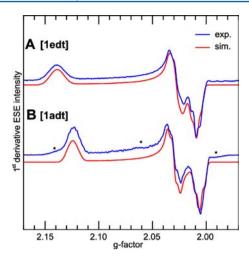


Figure 2. First derivative (pseudomodulated) Q-band two-pulse echodetected EPR spectra (blue) of frozen solutions of **1edt** (A) and **1adt** (B) and corresponding simulations (red) using principal g values: (A) $g^{\rm 1edt}=2.009,\ 2.028,\$ and $2.139;\$ (B) $g^{\rm 1adt}=2.006,\ 2.030,\$ and 2.124. Experimental conditions: T=20 K, $\nu_{\rm mw}=33.8560$ GHz (A), 33.918 GHz (B), and $\tau=340$ ns. The asterisks in B indicate minor contaminations of unknown origin.

rhombicity of the **g** matrix (g = 2.009, 2.028, and 2.139), whereas the **1adt** is more rhombic but less anisotropic (g = 2.006, 2.030, and 2.124). However, because the **g** matrix of both complexes is rather axial, it is apparent that both compounds have similar electronic structures that are only modestly influenced by the bridge alteration.

1. Spectroscopic Results on 1edt. Although the two strong ³¹P HF interactions (HFIs) can already be partially resolved in CW EPR, these interactions were also probed using orientation-selected pulse ENDOR measurements. Q-band Davies ENDOR spectroscopy revealed two different sets of ³¹P signals (Figure 3). One set corresponds to two strongly

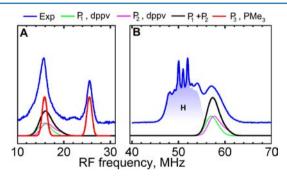


Figure 3. Q-band Davies ENDOR spectra (blue) of **1edt** measured at $T=20~\rm K$ and 1197.8 mT. Simulations were performed using $P_{1,2,3}$ parameters from Table 1. The $^1\rm H$ ENDOR region is shaded blue for clarity. Experimental conditions: (A) $t_{\rm RF}=35~\mu s$, $\nu_{\rm mw}=33.8708~\rm GHz$, and $t_{\rm inv}=200~\rm ns$; (B) $t_{\rm RF}=5.5~\mu s$, $\nu_{\rm mw}=33.8708~\rm GHz$, and $t_{\rm inv}=80~\rm ns$. See also Figure S2 in the SI for complete field dependence.

coupled ³¹P nuclei of the dppv ligand consistent with the splitting observed previously in the X-band CW EPR spectra. ³¹ The best fit to the field-dependent ENDOR signals was achieved assuming two slightly different ³¹P HF couplings: $A(P_1) = -72, -71$, and -80 MHz; $A(P_2) = -73, -72$, and -85 MHz. In the lower-frequency region, we have detected another doublet, centered at the Larmor frequency of ³¹P ($\nu_{31P} = 20.664$

MHz at 1197.8 mT), which has been assigned to a weakly coupled 31 P nucleus, A = 9.5, 11.0, and 8.5 MHz. The results of the simulations of these spectra are summarized in Table 1.

Table 1. Principal HFI Tensor Values (MHz) for 1edt and 1adt Obtained from Simulations of Experimental Spectra

					Euler	(deg)	
nucleus (assignment)	A_x	A_y	A_z	$A_{\rm iso}{}^a$	α	β	γ
			1edt				
$^{31}P_1$ (dppv)	-72	-71	-80	-74.3	0	30	0
$^{31}P_2$ (dppv)	-73	-72	-85	-76.6	0	30	0
$^{31}P_3$ (PMe ₃)	-9.5	-11.0	-8.5	-9.7	0	40	0
⁵⁷ Fe (Fe ₁)	-12.0	+16.6	-29.5	-8.3	10	10	0
⁵⁷ Fe (Fe ₂)	-3.0	-1.0	+0.5	-1.16	0	15	20
			1adt				
$^{31}P_1$ (dppv)	-70	-71	-78	-73.0	0	30	0
$^{31}P_2$ (dppv)	-69	-72	-85	-75.3	0	30	0
³¹ P (PMe ₃)	-4.3	-4.5	-3	-3.90	0	30	0
${}^{14}N_1^{\ b}$	+5.4	+3.7	+3.0	+4.0	10	25	0
${}^{14}N_2^{\ b}$	+0.0	+0.0	+3.5	+1.17	30	60	0
${}^{1}H_{1}$	+11.1	+15.8	+13.5	+13.4	40	30	0
$^{1}H_{2}$	+5.0	+8.5	+10.5	+8.0	30	30	0

 $^{a}A_{\rm iso} = (A_x + A_y + A_z)/3$. b The sign of the HF coupling is assumed based on DFT calculations (vide infra).

Because this ³¹P HF coupling is much smaller than those from the dppv ligand, we conclude that it originates from the PMe₃ ligand at the second iron (Fe₂), indicating some delocalization of the spin density to this site.

As will become apparent below, the sign of this small ^{31}P HF coupling plays an important role in understanding the orientation of the PMe₃ ligand. To resolve the sign of this HF coupling, we employed VMT ENDOR. 48,49 In this method, the sign of the HF coupling in question can be directly read out from polarization of the ENDOR signal upon an increase in the delay time between the radio-frequency (RF) pulse and the detection sequence, $t_{\rm mix}$ (Figure S1 in the SI), because of longitudinal electron-spin relaxation. As can be seen from Figure 4, even at relatively high temperatures (25 K), we have observed an almost completely polarized Q-band ENDOR spectrum when $t_{\rm mix}$ exceeds 100 μ s.

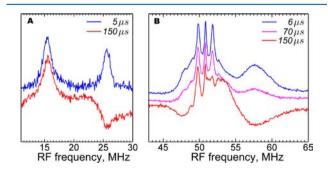


Figure 4. Q-band VMT (Davies) ENDOR of **1edt** measured for various $t_{\rm mix}$ delay times (see the SI) at maximum absorption of the EPR spectrum (g_y). Both low- (A) and high-frequency (B) ENDOR signals show a clear dependence on $t_{\rm mix}$ delay times (indicated in the figure). Experimental conditions: (A) T=25 K, $t_{\rm RF}=20$ μ s, $\nu_{\rm mw}=33.8269$ GHz, $B_0=1192.3$ mT, and $t_{\rm inv}=100$ ns; (B) T=25 K, $t_{\rm RF}=7$ μ s, $\nu_{\rm mw}=33.8865$ GHz, $B_0=1194.4$ mT, and $t_{\rm inv}=80$ ns.

For $S = \frac{1}{2}$ and $I = \frac{1}{2}$, the frequency of the ENDOR resonances is given by

$$\nu_{\alpha(\beta)} = 1 - g_{\rm p} \beta_{\rm p} B_0 + (-)A/2 \tag{3}$$

where g_n = nuclear g value, β_n = nuclear magneton, B_0 = strength of the magnetic field, A = orientation-dependent HF coupling constant. The assignment of the ENDOR signals to a specific M_s manifold (α or β) enables the direct determination of the absolute sign of A. As follows from the description of VMT ENDOR experiments, ⁴⁸⁻⁵⁰ at sufficiently long delay time between the RF pulse and detection (see the SI for the pulse sequence), $t_{\text{mix}} \ge T_{1\text{e}}$, the ENDOR signal from the $M_{\text{S}} = -\frac{1}{2}$ (β manifold) remains positive, while the $M_{\rm S}$ = $+^{1}/_{2}$ (α manifold) signal decreases relative to the other signal and even changes its sign. Considering that the g_n values for ³¹P and ¹H nuclei are positive, a positive A would result in $\nu_{\alpha} < \nu_{\rm L} = 1$ $g_n\beta_nB_0$ and $\nu_{\beta} > \nu_L$ (see eq 3). In this case, the lower-frequency component of the ENDOR doublet (i.e., ν_{α}) would be affected. However, in our experiment (Figure 4A), the opposite situation was observed, indicating that $\nu_{\alpha} > \nu_{\rm L}$. Thus, $A(^{31}{\rm P, PMe_3})$ is negative. Despite being unable to simultaneously observe α - and β -manifold signals, we can conclude that the ${}^{31}P(dppv)$ HF couplings similarly have negative signs because the highfrequency component of the ENDOR signal clearly decreases and becomes negative with increasing t_{mix} (Figure 4). In this experiment, polarization of the ¹H signals has also been detected; however, no definite assignment could be ascertained from the overlapping signals.

Although the observation of the weakly coupled ^{31}P nucleus indicates some degree of spin delocalization on Fe₂, more direct insight into the electronic structure for these model compounds is obtained by a study of the Fe centers themselves. Investigating the electronic environment of Fe atoms in coordination complexes and protein samples is routinely compromised by the absence of a nuclear spin of sufficient abundance. Naturally occurring 57 Fe ($I = ^{1}/_{2}$) has a meager 2.2% abundance, so isotropic enrichment is necessary. In this project, we benefit from the recently developed methods for the 57 Fe enrichment of **1edt**. ⁴⁷ By a combination of Mössbauer and pulse EPR spectroscopic methods to these highly enriched samples, we obtained the 57 Fe HF coupling constants with high accuracy.

Applied-field ⁵⁷Fe Mössbauer spectra of **1edt** measured at 4.2 K consist of two signals corresponding to the two Fe sites in the complex (Figure 5). One of the signals at 0.1 T represents a typical quadrupole doublet with HF coupling well hidden under the line width, thus indicating relatively weak ⁵⁷Fe HFI. The other contributing signal has a rather complex and broad shape even at low magnetic fields. The overall width of this signal remains invariant with increasing magnetic field. The spectral profile of this signal remains unchanged at 1.8 K, eliminating the possibility for a strong relaxation case that could also result in considerable broadening of the spectra. We therefore conclude that this second contributor is subject to a relatively large and very anisotropic ⁵⁷Fe HF tensor.

Because of the complexity of the signal at all measured magnetic fields, no unique set of Mössbauer parameters could be unambiguously extracted. To increase the number of constraints and thus obtain a unique set of spin-Hamiltonian parameters, additional pulse EPR experiments were conducted on the ⁵⁷Fe-labeled **1edt** sample.

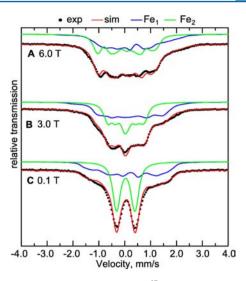


Figure 5. Mössbauer spectra of the fully 57 Fe-enriched **1edt** measured at three magnetic field strengths at a temperature of 4.2 K (black) and respective simulations (red) accounting for two contributing 57 Fe nuclei: Fe₁ (blue) and Fe₂ (green) with parameters presented in Tables 1 and 2.

The simulations exhibited in Figure 5 were obtained from a simultaneous fit of both Fe nuclei with the inclusion of fixed coupling parameters obtained from high-frequency ESEEM and HYSCORE data (vide infra). The ⁵⁷Fe-labeled **1edt** samples show distinct signals in the low-frequency region of the Q- and W-band ESEEM spectra (Figures 6 and 7) that are consistent with a weakly coupled ⁵⁷Fe nucleus. A better resolution was obtained from HYSCORE measurements at the Q-band frequency. The noticeable field dependence of these ⁵⁷Fe signals indicates a very anisotropic character of the HF coupling. The simulation could be performed using one set of parameters with a good fit to the experimental spectra (Figure 7). To corroborate these assignments, we have also performed these HYSCORE experiments at W-band frequency (see the SI). The resulting ⁵⁷Fe parameters match the Q-band data completely.

Concerning the second ⁵⁷Fe signal, as in the case of the Mössbauer data, the high-frequency W-band ESEEM signals cannot be fit with a unique set of parameters. However, because Mössbauer and pulse EPR data impose different constraints, a simultaneous simulation of all of the data (Q/W-band ESEEM and HYSCORE and Mössbauer) yielded a unique set of parameters of two ⁵⁷Fe HF couplings. Figures 5–7 include simulations obtained from this process. It is worth noting that this approach also allowed identification of the sign of each ⁵⁷Fe HF coupling despite their pronounced anisotropy.

The ⁵⁷Fe HF coupling constants are consistent with the weakly coupled ⁵⁷Fe signals resolved in Mössbauer experiments. The W-band ESEEM spectra at selected field positions showed additional signals that could be attributed to a second strongly coupled ⁵⁷Fe nucleus. Figure 7 shows two field-dependent series of three-pulse ESEEM spectra obtained for two different τ values. On the basis of these experiments, we can confirm that the spin density is predominantly located at Fe₁. However, as indicated by the presence of weakly coupled ⁵⁷Fe and ³¹P nuclei, about 5–10% of the spin is found on Fe₂. Table 1 summarizes the ⁵⁷Fe and ³¹P HF coupling parameters. Table 2 shows ⁵⁷Fe Mössbauer-specific parameters, extracted from the spectra.

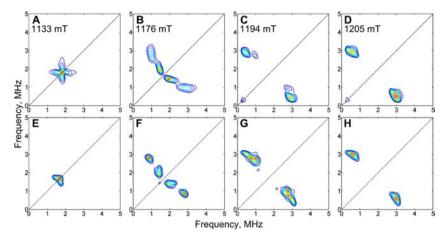


Figure 6. Experimental (A–D) Q-band HYSCORE spectra of **1edt** measured at the indicated field positions and corresponding simulations (E–H) that account for one ⁵⁷Fe HF coupling [$A(\text{Fe}_2)$; Table 1]. Experimental conditions: T = 20 K, $\tau = 420$ ns, $\nu_{\text{mw}} = 33.8883$ GHz, $t(\pi/2) = 40$ ns, and $\Delta T_{1,2} = 52$ ns.

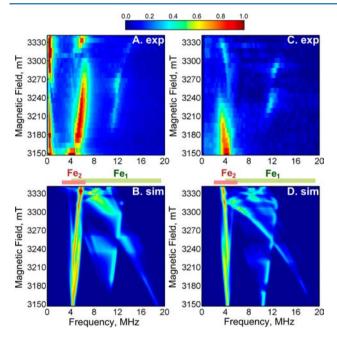


Figure 7. W-band three-pulse ESEEM spectra as a function of the applied magnetic field B_0 of ⁵⁷Fe-enriched **1edt**, measured at two different τ values: 164 ns (A) and 288 ns (C), together with the respective simulations (B and D) accounting for two contributing ⁵⁷Fe HF couplings (Table 1). Experimental conditions: T=10 K, $\tau=164$ ns, $\nu_{\rm mw}=94.0928$ GHz, $t(\pi/2)=24$ ns, and $\Delta T=12$ ns. The color coding of the relative intensities of the signal is represented by the color bar.

Table 2. ⁵⁷Fe Quadrupole and Isomer Shift Parameters Obtained from Mössbauer Data for 1edt

				Euler angles (deg.)		
nucleus	$\Delta E_{\rm Q} ({\rm mm/s})$	η	$\delta \; (\text{mm/s})$	α	β	γ
⁵⁷ Fe (Fe ₁)	-0.58(3)	+0.90(10)	+0.20(1)	20	60	0
⁵⁷ Fe (Fe ₂)	+0.70(3)	+0.92(10)	+0.04(1)	0	-10	0

2. Spectroscopic Results on 1adt. To assess the impact of a nitrogen atom in the dithiolate bridge (azadithiolate), we have pursued similar EPR investigations of **1adt** (Figure 1). Qband ENDOR spectra of **1adt** displayed in Figure 8 were more intricate than those of its **1edt** counterpart (Figure 3). Aside

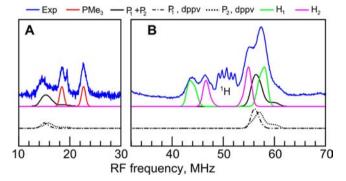


Figure 8. Q-band ENDOR spectra of **1adt** measured at 20 K and field position 1192.8 mT (g_y). Low-frequency spectra (A) were recorded using the refocused Mims ENDOR sequence with $\tau=108$ ns and $t(\pi/2)=16$ ns, whereas the high-frequency part (B) was recorded using Davies ENDOR, $t_{\rm inv}=44$ ns (suppressing matrix $^1{\rm H}$ signals), $t(\pi/2)=16$ ns, and $\tau=400$ ns. Simulations were performed using the HF coupling constants listed in Table 1. See also Figure S3 in the SI for complete field dependence.

from the strongly coupled ³¹P signals, a distinct set of ¹H signals was observed (Figure 8B). The pattern with a splitting in the range 10–15 MHz remained centered at the ¹H Larmor frequency and unperturbed by variation in the magnetic field, indicating a mostly isotropic HF coupling (Table 1). Tentatively, we attribute the observed ¹H signals to the methylene and/or amine protons of the azadithiolate moiety. This assignment is supported by DFT calculations, presented in the following section. The additional structure at the high-frequency side of the proton range reveals ³¹P signals, most clearly seen at the lowest measured magnetic field position (Figure 8B). These arise from two signals from either inequivalent nuclei of the dppv ligand or the presence of two isomeric forms of the complex.

In the low-frequency region, the doublet at the ³¹P Larmor frequency is assigned to the weakly coupled PMe₃ ligand (Figure 8A). Its field dependence (see Figure S3 in the SI) showed this coupling to be mostly isotropic. It is interesting to note that this ³¹P HF coupling is significantly smaller in magnitude than that for **1edt**.

From Q-band VMT (Davies) ENDOR experiments of **1adt** (Figure S4 in the SI), we conclude that all ³¹P signals (from PMe₃ and dppv) correspond to negative HF couplings, as for

1edt. Conversely, the polarization pattern of the ¹H signals in Figure S4 in the SI identifies the strongly coupled ¹H HF couplings as positive. Despite the lack of viable synthetic access to ⁵⁷Fe-enriched variants of **1adt** that precludes ⁵⁷Fe Mössbauer and EPR studies similar to those conducted on enriched **1edt**, it is, nevertheless, apparent from the ³¹P(PMe₃) HF couplings that the electronic structure of **1adt** is distinct from that of **1edt**.

A combination of ESEEM and HYSCORE measurements enabled determination of the HF and quadrupole coupling constants of the ¹⁴N atom in the azadithiolate ligand because it is the only N atom in the system. However, the observed X-band HYSCORE spectra (Figures 9 and S6 in the SI) reveal

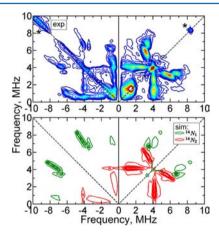


Figure 9. X-band HYSCORE spectra of **1adt** (top) and corresponding simulations (bottom) accounting for two independent ¹⁴N signals that correspond to two isomeric forms of the amine group (see also Figure S6 in the SI). Experimental parameters: $B_0 = 342.5$ mT, T = 20 K, $\nu_{\rm mw} = 9.720$ GHz, $t(\pi/2) = 8$ ns, and $\tau = 86$ ns. For clarity, the presented spectra are cropped to the low-frequency range, neglecting the ¹H signals. Asterisks denote instrumental artifacts.

rather complex 14 N signals. The majority of the signals in the (++) quadrant can be attributed to a single 14 N nucleus. Because the ridges in the HYSCORE spectra change considerably with the position of the magnetic field (Figure S6 in the SI), we assume a highly anisotropic 14 N HF coupling. Closer examination of the spectra reveals additional signals that are most prominent in HYSCORE measurements with relatively short τ values. These signals have a weaker magnetic field dependence and are attributed to a predominantly isotropic HF coupling.

Additional Q-band HYSCORE measurements produced two sets of signals as well (see the SI). Spectral simulations depicted in Figure 9 yielded two distinct sets of ¹⁴N HF parameters (Table 1). This observation can only be rationalized by attributing each HF coupling to one of two isomers of **1adt**, presumably arising from a bridge flip (eq 4). The almost identical magnitude of the quadrupole coupling for the observed ¹⁴N signals corroborates this assignment (Table 3).

Table 3. ¹⁴N Quadrupole Parameters Obtained from X-band HYSCORE Analysis of 1adt

			Eule	r angles (d	eg)
	K(MHz)	η	α	β	γ
¹⁴ N ₁	1.12(1)	+0.20(5)	10	20	0
$^{14}N_{2}$	1.15(1)	0.00(10)	0	70	0

Although the geometry of the amine ligand in the model is presumably identical with that of the H-cluster, the measured ^{14}N quadrupole coupling in 1adt is $\sim\!10\%$ smaller than the one observed in the native enzyme [K(1adt)=1.12/1.15 MHz] versus $K(H_{ox})=1.23\text{ MHz}].^{12}$ We believe that this difference stems from the absence of neighboring protein residues that affect the site symmetry of the N atom in the enzyme.

3. Quantum-Chemical Study of 1edt and 1adt. Previous studies of inorganic models of the H_{ox} -CO state, $[Fe_2(\mu\text{-CO})(CO)_3(CN)_2(\text{dithiolate})]^-$ [where dithiolate = $\{\text{MeSCH}_2\text{C}(\text{Me})(\text{CH}_2\text{S})_2\}$ (2)²⁶ or $\{\text{MeS}(\text{CH}_2)_2\text{N}(\text{CH}_2\text{S})_2\}$ (3)²⁷], produced calculated values in good agreement with the physical observables. However, while 2 and 3 show a highly delocalized spin density, the present compounds evidently produce a highly localized spin distribution.³¹

On the basis of geometry optimizations and earlier studies,^{29,31} two principal geometric configurations of the ligands about Fe₁ in 1edt are considered: (i) dithiolate and dppv ligands form a square-pyramidal arrangement with a semibridging position of the CO ligand (designated as "flat") and (ii) twisted arrangement of the dppv ligand with respect to the dithiolate bridge and CO in a terminal basal position and henceforth referred to as "twist" (Figure 10). For each of these two arrangements, two positions of the PMe₃ ligand at Fe₂, being apical ("ap") or basal ("bas") with respect to the dithiolate bridge, are critical to the degree of spin delocalization in the molecule. An earlier X-ray crystallographic study of the reduced variant of ledt revealed an apical position of this ligand.²⁹ However, it should be noted that the conditions for sample preparation as well as the measurement conditions are different in the current case. Because the formation of the crystal implies a tight packing of molecules, lattice forces might as well favor a different isomer. Hence, in this analysis, we consider the four geometric alternatives shown in Figure 10A.

Following the previous approach for 2 and 3, i.e., spin-unrestricted B3LYP-DFT calculations with all-electron valence triple- ζ basis sets (TZVP), the ³¹P HF coupling constants for 1edt and 1adt were nicely reproduced. However, the correspondence between the experimental and computed ⁵⁷Fe HF coupling constants was less favorable. This situation did not improve when alternative popular hybrid functionals such as PBE0 and TPSSh were used or by including a dielectric continuum.

The calculated 57 Fe $A_{\rm iso}$ values were found to be strongly affected by the basis set. Figure 11 presents variation in the 57 Fe $A_{\rm iso}$ values with the different basis sets used. TZVPP was maintained for all other atoms, having no substantial impact on the 57 Fe HF coupling constants. Surprisingly, we observed that, depending on the flexibility of the core region of the iron basis sets, the results fall into two distinct categories. The only exception is the rather large atomic natural orbital (ANO) basis set Roos-ANO-TZP. Illustrative of the overall sensitivity of the computed HF values to the basis set is the finding that decontraction of a relatively small basis set SVP (indicated as

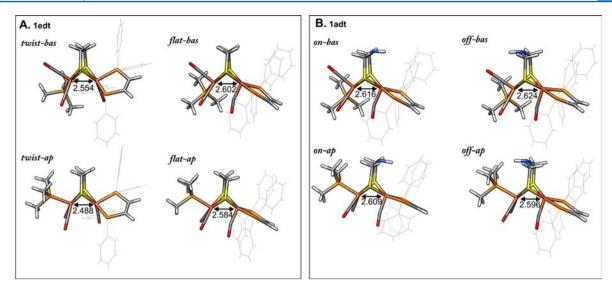


Figure 10. Geometry-optimized isomers of 1edt (A) and 1adt (B). Fe₁—Fe₂ distances are indicated in angstroms. "bas"/"ap" stands for the basal or apical position of the PMe₃ ligand, respectively; "flat"/"twist" stands for the flat (square-pyramidal) configuration of dithiolate and dppv ligands and the twisted configuration; "on"/"off" indicates the position of the amine with respect to the dppv ligand.

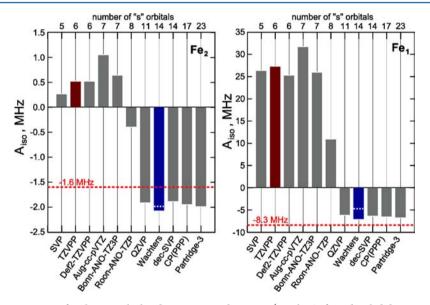


Figure 11. 57 Fe A_{iso} coupling constants of $1edt_{flat-bas}$ calculated using various basis sets (see the SI for a detailed description) in conjunction with the B3LYP functional and TZVPP basis set on all other atoms, not taking into account second-order spin—orbit coupling contributions. The red dashed lines indicate the experimentally obtained values from Table 1. Brown and blue bars identify TZVPP and Wachters basis sets, respectively. The white dashed lines in the blue bars indicate spin—orbit coupling corrections to the isotropic HF coupling.

"dec-SVP") leads to a completely different result, which is, in fact, much closer to the experiment.

Overall, the "Wachters" basis set⁴³ was preferred for computing the EPR and Mössbauer parameters; the data for **1edt** are given in Table 4. For calculations in vacuo, the energy differences between the various structures were found to be relatively small (<6 kcal/mol). Nevertheless, as expected based on the structural differences, the "flat" and "twist" conformations differ considerably in the spin-density distribution (see the SI, Figure S10). In the "flat" case, the spin density is predominantly localized on Fe₁, while it is delocalized over both Fe₁ and Fe₂ in the twisted isomers. The two classes of isomers also differ dramatically with respect to the electric field gradient constant on Fe₁. Given that a substantial quadrupole splitting is computed for the twisted isomers, we infer that only "flat"

isomers are present in the measured samples in accordance with the experimental quadrupole splitting.

Although the calculated magnitude of the ^{31}P HF coupling of the PMe₃ ligand is similar in the apical and basal conformations, its sign switches. This difference can be understood by considering that the spin density resides in a d_z^2 -like orbital at Fe₂, and therefore the sign is positive for the position of the ligand along the axial direction of the d_z^2 orbital (apical isomer) and negative in the equatorial plane of the d_z^2 orbital (basal isomer). On the basis of the sign determination using VMT ENDOR measurements, we conclude that the PMe₃ ligand in **ledt** is in the basal position. Although the crystal structure of **ledt** (as its BF₄⁻ salt) shows an apical PMe₃ in the solid, the structure of a related Fe₂(pdt)(CO)₄(Imes)(PMe₃)]PF₆ features basal PMe₃ on Fe₂. Sure assignment is further corroborated by the fact that the "flat-bas" isomer has the

Table 4. Comparison of Experimental and Calculated Parameters for 1edt

	twist-ap	twist-bas	flat-ap	flat-bas	exp
ΔE (kcal/mol)	5.73	5.76	2.03	0.00	
⁵⁷ Fe, Fe ₁ ^a					
A _{iso} (MHz)	-0.55	+0.97	-4.59	-4.74	-8.30
$A_{\rm dip}~({ m MHz})$	+11.14	+9.64	+15.97	+16.58	+12.45
$A_{ m rh}$	+0.68	+0.99	+0.46	+0.35	+ 0.70
$\Delta E_{\rm Q} ({\rm mm/s})^b$	+1.47	-1.17	-0.41	-0.49	-0.58
η	0.87	0.87	0.91	0.97	0.90
$\delta (\text{mm/s})^c$	+0.15	+0.14	+0.17	+0.16	+0.20
Mulliken	0.7448	0.7023	0.9740	1.0189	
⁵⁷ Fe, Fe ₂ ^a					
$A_{\rm iso}~({ m MHz})$	-0.57	0.55	-2.25	-1.99	-1.16
$A_{ m dip}$ (MHz)	+5.77	+7.17	+2.02	+1.43	+0.90
A _{rh} (MHz)	+0.64	+0.79	+0.30	+0.23	+0.82
$\Delta E_{\rm Q} ({\rm mm/s})^b$	+0.70	-0.59	+0.67	+0.80	+0.70
η	0.99	0.69	0.52	0.48	0.92
$\delta \; (\text{mm/s})^c$	-0.03	-0.01	+0.01	+0.01	+0.04
Mulliken	0.3946	0.4178	0.1147	0.0797	
31 P (dppv), A_{iso} (MHz)					
P_1	-66.85	-60.13	-85.17	-82.29	-74.3
P_2	-60.78	-58.69	-84.39	-85.70	-76.6
³¹ P (PMe ₃), A _{iso} (MHz)					
P_3	+8.15	-40.22	+6.10	-11.91	-9.70

 $[^]aA_{\rm iso}$ = isotropic part, $A_{\rm dip}$ = magnitude of the anisotropic part, $A_{\rm rh}$ = rhombicity of anisotropic part, i.e., degree of deviation from axial character as such: $A = A_{\rm iso} + A_{\rm dip}[-1 + A_{\rm rh}, -1 - A_{\rm rh}, 2]$. bB ecause the asymmetry parameter is almost 100%, the sign of $\Delta E_{\rm Q}$ does not have a physical meaning. $^c\delta = 0.1181 - 0.3821[\rho(0) - 11614]$; see the Materials and Methods section.

Table 5. Comparison of Experimental and Calculated Parameters for 1adt

	on-ap	on-bas	off-ap	off-bas	\exp^c
ΔE (kcal/mol)	2.79	0.00	4.16	0.32	
¹⁴ N (adt) ^a					
A _{iso} (MHz)	+3.92	+2.91	+0.59	+0.83	+4.00/+1.17
$A_{\rm dip}~({ m MHz})$	+0.21	+0.22	+0.66	+0.72	+0.68/+1.17
$A_{ m rh}$	+0.34	+0.24	+0.03	+0.03	+0.51/+0.0
K (MHz)	-1.24	-1.25	-1.24	-1.22	1.12/1.15
η	0.09	0.07	0.07	0.11	0.20/0.0
³¹ P (dppv), A _{iso} (MHz)					
P_1	-88.27	-83.85	-86.23	-82.08	-73.0
P_2	-92.01	-88.30	-86.22	-87.38	-75.3
31 P (PMe ₃), A_{iso} (MHz)					
P_3	+8.39	-9.61	+9.59	-6.67	-3.90
1 H (adt), $A_{\rm iso}$ (MHz) b					
H_1	+2.68	+1.37	+7.05	+4.95	+8.30
H_2	+1.95	+1.78	+7.07	+7.67	+13.50
H_3	-1.30	-0.81	-0.87	-1.19	
H_4	-1.32	-0.77	-1.07	-1.00	
1 H (NH), $A_{\rm iso}$ (MHz)					
H_{s}	-0.60	-0.41	-1.07	-0.92	

[&]quot; $A_{\rm iso}$ = isotropic part of HF coupling, $A_{\rm dip}$ = magnitude of the anisotropic part, $A_{\rm rh}$ = rhombicity of anisotropic part, i.e., degree of deviation from axial character as such: $A = A_{\rm iso} + A_{\rm dip}[-1 + A_{\rm rh}, -1 - A_{\rm rh}, 2]$. For assignment of the 1 H HF couplings, see Figure S11 in the SI. If no sign is given, it is not determined experimentally.

lowest calculated single-point energy, be it by only 2 kcal/mol. Moreover, the spin-density distribution mapped by DFT for the "flat-bas" isomer, with the unpaired electron essentially localized in a d_z^2 -like orbital of Fe₁ with minimal delocalization to the other Fe site, best matches the experiment. Knowing that we can reproduce the general electronic structure using DFT methods, we can now compare data for the **1edt** and **1adt** variants.

For **1adt**, we include two possible arrangements of the azadithiolate ligand as an additional degree of freedom. We will call the orientation of the amine facing the dppv ligand "on" and the orientation to the other side "off". We found that the overall electronic structure was insensitive to the orientation of the amine proton. In all cases examined, the amine proton projects toward the metal core because this afforded the most stable geometry. Additionally, we disregarded the "twist" isomer

because the overall similarity in the EPR spectra of **1edt** and **1adt** implies that the "flat" isomer is dominant. When only the "flat" arrangement is selected for the dppv ligand, four isomers remain to be considered for **1adt**, as depicted in Figure 10B.

We noticed that the position of the amine perturbs the arrangement of the dppv phenyl substituents; projection of the NH group into this space results in a minor tilt of the dppv ligand, which could explain the slight inequivalence of the two ³¹P(dppv) HF couplings. If the amine is flipped in the opposing direction, it affects the position of the PMe₃ ligand when this is in the apical position. Hence, the "off-ap" isomer has the highest calculated single-point energy. As was the case for **1edt**, the negative sign of the PMe₃ ³¹P HF coupling corresponds to a basal orientation of this ligand for **1adt**.

Concerning the orientation of the amine, the two experimentally extracted sets of ¹⁴N HF coupling constants match the ones calculated for the "on" and "off" positions. If the nitrogen points toward the dppv ligand (and is closer to the spin density "cloud" near Fe), a large HF coupling constant is encountered. A much smaller ¹⁴N HF coupling constant is obtained when the amine is pointing to the other side. Interestingly, only in the "off" isomers are the methylene protons of the dithiolate bridge (but not the amine proton) calculated to have relatively large HF couplings (Table 5). The sign of these ¹H HF couplings matches the experimental assignment. We therefore conclude that the two isomers of 1adt detected in our experiments are "on-bas" and "off-bas".

Comparing the calculated values for these isomers with the experimental data, we notice that the ¹H and ¹⁴N HF coupling constants are considerably underestimated while the ³¹P HF coupling for the PMe₃ ligand is highly overestimated. While for **1edt** the calculated ³¹P HF couplings matched very well with the experimental results, the changes induced by the amino group in the bridging ligand, i.e., a reduction of the spin density at Fe₁ with a concomitant increase on the dithiolate bridge, were not as clearly reproduced in the calculations. However, the HOMO is indeed more distributed toward the dithiolate bridge in **1adt** than in **1edt** (Figure S10 in the SI).

DISCUSSION

DFT Accuracy. DFT calculations have reproduced the salient electronic structural features of the two models 1edt and 1adt quite well. The calculated single-point energy differences between the various isomers are consistent with the experimental observations. The spin density was found to be localized on the five-coordinated Fe₁ site, with some delocalization toward the six-coordinated Fe2 site. A satisfactory agreement was also found for the ⁵⁷Fe Mössbauer parameters. Conversely, the calculated values of the ⁵⁷Fe HF coupling constants are strongly influenced by the flexibility of the basis set. This is not unexpected, but, surprisingly, the dependence is not monotonic. Instead, two quite distinct cases were obtained with respect to the 57Fe isotropic HF coupling constant for 1edt. This apparently relates to two potential minimum solutions that could be reached depending on the basis set core functions. This is a rather important finding because ⁵⁷Fe data provide critical insight into the electronic structure of diiron compounds and their reproducibility by DFT is crucial. Nevertheless, analysis yields a logical trend: a larger basis set provides more accurate results. To verify, whether the same holds for the previously published models of the H_{ox}-CO state (2), we performed test calculations at the B3LYP level. Similarly, two different cases for the ⁵⁷Fe HF coupling could be

achieved depending on the basis set (see the SI). Quite interestingly, the completely opposite scenario arises; i.e., a *smaller* basis set provides a better fit to the experimental data than a large one. For 2, the original standard contracted SVP basis set gives a very satisfactory agreement (see the SI, Figure S9). Therefore, we conclude that, despite the fact that the used methods can indeed reproduce the EPR parameters reasonably well, the DFT methods lack predictive power, and for each state of the system, the basis sets should be "tailored" to fit the experimental data. It remains to be seen if this problem is inherent to the DFT method or if further theoretical developments in DFT might come up with strategies to alleviate or circumvent this "bistability" effect.

A more subtle problem lies in the fact that the DFT methods did not identify electronic structural dissimilarities between **1edt** and **1adt**. According to the calculated spin-Hamiltonian parameters, the electronic structures of these two compounds are virtually identical while experimentally there are obvious distinctions.

1edt vs 1adt. Although a spin-localized structure is adopted by both variants, subtleties in this distribution are evident in the spectroscopic analysis. From ⁵⁷Fe HF couplings in 1edt, about 10% of the spin density is dispersed on Fe2, which commutes with a substantial ³¹P HF coupling constant of the PMe₃ ligand. Upon insertion of an amine, the electronic structure changes. The ^{31}P (PMe₃) A_{iso} coupling decreases by a factor of 3, attended by a loss of spin density from that ligand. We can exclude a conformational change from basal to apical because the sign of the HF coupling is retained. Furthermore, the EPR spectrum of ladt is less anisotropic and more rhombic, corresponding to a modification of the frontier molecular orbital composition. The similar ³¹P HF coupling of the dppv ligand underscores a nearly identical spin population on Fe1 in 1edt and 1adt. Interestingly, the ¹H ENDOR spectra resolve strongly coupled methylene protons on the azadithiolate bridge in contrast to the case of ledt, where the methylene protons are much more weakly coupled. The PMe₃ ³¹P HFI in **1adt** is only one-third that in 1edt. This would suggest that the azadithiolate bridge perturbs the charge distribution in the diiron core, resulting in a decrease of the spin population on Fe₂ and a concomitant increase of the spin population on the dithiolate bridge. This accounts for variation in the electronic properties of the two dithiolates without amending the geometry. The steric and electronic impact of the amine functionality in the dithiolate bridge is expressed in the composition of the molecular orbitals in the Fe₂S₂ unit and is consistent with the ease with which ladt can be reduced.³¹ Additionally, preliminary calculations suggested that the S-C bonding is affected by interactions with the nitrogen lone pair via the so-called anomeric effect, leading to enhanced coupling of the heteroatom with the Fe_2S_2 core.⁵³

analogous trend was identified in the native H-cluster, wherein the spin-density distribution was also found to shift toward the proximal Fe upon CO inhibition of the H_{ox} state. However, there is a very significant difference between the actual spin distributions in the models and native system. In contrast to the model systems, a delocalized picture has been observed in the H_{ox} state and a localized one in the H_{ox} -CO state.

As previously detailed, the electronic structure of the oxidized states of the H-cluster is heavily influenced by intercluster Heisenberg exchange interactions. ^{19,26} However, accounting for a Heisenberg spin—spin exchange mechanism alone seems insufficient to account for the discrepancy between the $H_{\rm ox}$ state and its inorganic mimic, **1adt**. Moreover, the exchange interaction is approximately 3–4 times weaker than that in the $H_{\rm ox}$ –CO state, and therefore the [4Fe–4S] cluster presumably has a minimal impact on the [2Fe] center. Contrarily, the DFT-computed HOMO for the diiron unit is strikingly similar to those obtained for our model complexes, ^{12,18} and therefore we expect to see a localized spin density for the $H_{\rm ox}$ state. There could be several reasons for this discrepancy:

(i) The coordination geometry of the H_{ox} state may deviate from the depiction given in Figure 1 owing to the inclusion of water in the vacant (sixth) coordination site of Fe_d, as inferred from crystallographic data of [FeFe]hydrogenase from Clostridium pasteurianum⁷ and its recent refinement.¹⁷ In recent DFT studies, the presence of such a molecule has been shown to significantly affect the electronic structure. 12,18 However, because the authors specify the use of the reductant dithionite in the crystallization buffer, it is doubtful that under these conditions the Hox state is dominant in the crystal. Moreover, low-temperature illumination experiments questions presence of water at the same site (see the arrow in Figure 1). Below 60 K, the external CO ligand can be photodissociated, regenerating the H_{ox} state, as indicated by IR and EPR measurements. 10,15,54,55 Such a conversion argues against binding water at the same site. It is worth noting that CO photodissociation was observed even at 8 K. Taking into consideration the compact arrangement of amino acids about the open coordination site, dissociation of a CO ligand and instantaneous binding of a water ligand would require substantial motional freedom of the protein matrix, which is clearly unrealistic at these temperatures. Thus, it is very likely that no ligand is present at the sixth coordination site of the distal Fe in the Hox state.

(ii) The interaction with the surrounding protein environment may have a substantial effect on the electronic structure of the H-cluster. Our recent DFT calculations showed a surprisingly small influence of the protein matrix on the spin distribution in the H-cluster, 18 although Szilagyi et al. reported the opposite effect on the spin populations by including key amino acid residues.⁵⁶ Unfortunately, these results raise more questions about the reliability of such calculations on spinexchange-coupled Fe cofactors such as the H-cluster. However, because the binuclear cluster is a "dangling" unit with only one covalent bond to the protein (i.e., the Cys-S bridge between the 2Fe and 4Fe-4S subclusters), the protein matrix is expected to exert only a minor influence on the electronic structure of the H-cluster. Nonetheless, the influence of the protein on the electronic structure of the active site remains an important area of investigation.

(iii) The distinctive electronic interaction between the [2Fe] and [4Fe-4S] subclusters seems to be the most likely

explanation for the differences between the model systems and the H-cluster. It clearly transcends a simple Heisenberg exchange approximation because this would not result in a redistribution of the spin density in the binuclear subcluster. Fig. 8 Increasing the exchange interaction merely attenuates the observable spin population of the [2Fe] subcluster, and the Heisenberg approach cannot accommodate alternative mechanisms such as double exchange. It would be interesting to study this phenomenon in more detail in model complexes featuring a second redox center, thus mimicking the intracluster interaction in the H-cluster.

The properties of the amine in the dithiolate bridge is a further variable to consider in this system. It is apparent from the experimental data that two conformations of the adt ligand are probable, highlighting the flexibility of this bridge. A similar conclusion resulted from DFT calculations of the H-cluster, ¹⁷ where this flexibility may be pivotal in modulating catalytic rates because it relays protons during turnover. A recently synthesized family of [Ni(diphosphine)₂] catalysts exploits the flexibility of the pendant base throughout the catalytic cycle. ²⁵

CONCLUSIONS

A detailed EPR and Mössbauer study of two structural diiron models for the Hox state of the H-cluster is presented. The spectroscopic data indicate that the ligand environment of the 1edt and 1adt models is characterized by "flat" (dppv) and "basal" (PMe3) configurations, which geometrically parallel the H-cluster. While the H_{ox} state of the H-cluster shows a delocalized spin density, 1edt and 1adt are appropriately formulated as mixed-valent Fe2 IFe1 pairs with the spin predominantly found on Fe₁ (in a d_z²-like orbital). Inserting an amine into the dithiolate bridging ligand, i.e., going from **1edt** to **1adt**, pushes the spin density away from Fe₂ toward the adt ligand accompanied by large HF couplings of the methylene protons and a measurable HF and quadrupole interaction of the ¹⁴N at the apex of the bridge. The adt ligand exhibits conformational flexibility, leading to two configurations ("on" and "off"). DFT calculations corroborate the general picture derived from the experimental data. The ligand HFIs (31P, 1H, and ¹⁴N) are well reproduced, but the properties calculated for the iron core, i.e., the ⁵⁷Fe HFIs, are highly dependent on the chosen basis set. Finally, the calculated spin-density distribution was found to be essentially the same for 1edt and 1adt, which is not evident in the experiment.

This study provides valuable insight into the electronic and structural properties of the presented model systems. The differences between the resolved spectroscopic parameters of **1edt** and **1adt** and those obtained for the H-cluster underscore the difficulty in reproducing the precise electronic environment programmed into the native system, with all parts playing an important role in tuning this efficient hydrogen catalyst for the best performance.

ASSOCIATED CONTENT

S Supporting Information

Extended experimental data on both **1edt** and **1adt** and details of the DFT calculations. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We are grateful to Dr. E. Bill for many useful discussions and to B. Mienert, G. Klihm, and F. Reikowski for their technical expertise and invaluable assistance during the experiments. This work was financially supported by the EU/Energy Network Project SOLAR-H2 (FP7 Contract 212508) and National Institutes of Health (Grant GM061153).

REFERENCES

- (1) Lubitz, W.; Reijerse, E.; van Gastel, M. Chem. Rev. 2007, 107 (10), 4331–4365.
- (2) Barton, B. E.; Olsen, M. T.; Rauchfuss, T. B. Curr. Opin. Biotechnol. 2010, 21 (3), 292-297.
- (3) Tard, C.; Pickett, C. J. Chem. Rev. 2009, 109 (6), 2245-2274.
- (4) Cammack, R.; Frey, M.; Robson, R. L. Hydrogen as a fuel: learning from nature; Taylor & Francis: London, 2001.
- (5) Lubitz, W.; Tumas, W. Chem. Rev. 2007, 107 (10), 3900-3903.
- (6) Salomone-Stagni, M.; Stellato, F.; Whaley, C. M.; Vogt, S.; Morante, S.; Shima, S.; Rauchfuss, T. B.; Meyer-Klaucke, W. *Dalton Trans.* **2010**, 39 (12), 3057–3064.
- (7) Peters, J. W.; Lanzilotta, W. N.; Lemon, B. J.; Seefeldt, L. C. Science 1998, 282 (5395), 1853-1858.
- (8) Nicolet, Y.; Piras, C.; Legrand, P.; Hatchikian, C. E.; Fontecilla-Camps, J. C. Structure 1999, 7 (1), 13–23.
- (9) Pierik, A. J.; Hulstein, M.; Hagen, W. R.; Albracht, S. P. Eur. J. Biochem. 1998, 258 (2), 572-578.
- (10) Albracht, S. P.; Roseboom, W.; Hatchikian, E. C. J. Biol. Inorg. Chem. 2006, 11 (1), 88–101.
- (11) Nicolet, Y.; de Lacey, A. L.; Vernede, X.; Fernandez, V. M.; Hatchikian, E. C.; Fontecilla-Camps, J. C. J. Am. Chem. Soc. 2001, 123 (8), 1596–1601.
- (12) Silakov, A.; Wenk, B.; Reijerse, E.; Lubitz, W. Phys. Chem. Chem. Phys. **2009**, 11 (31), 6592–6599.
- (13) Fan, H. J.; Hall, M. B. J. Am. Chem. Soc. 2001, 123 (16), 3828–3829.
- (14) Barton, B. E.; Olsen, M. T.; Rauchfuss, T. B. J. Am. Chem. Soc. **2008**, 130 (50), 16834–16835.
- (15) Roseboom, W.; de Lacey, A. L.; Fernandez, V. M.; Hatchikian, E. C.; Albracht, S. P. *J. Biol. Inorg. Chem.* **2006**, *11* (1), 102–118.
- (16) Nicolet, Y.; Lemon, B. J.; Fontecilla-Camps, J. C.; Peters, J. W. *Trends Biochem. Sci.* **2000**, 25 (3), 138–143.
- (17) Pandey, A. S.; Harris, T. V.; Giles, L. J.; Peters, J. W.; Szilagyi, R. K. J. Am. Chem. Soc. **2008**, 130 (13), 4533–4540.
- (18) Greco, C.; Silakov, A.; Bruschi, M.; Ryde, U.; De Gioia, L.; Lubitz, W. Eur. J. Inorg. Chem. **2011**, 1043–1049.
- (19) Silakov, A.; Reijerse, E. J.; Albracht, S. P.; Hatchikian, E. C.; Lubitz, W. J. Am. Chem. Soc. 2007, 129 (37), 11447-11458.
- (20) Silakov, A.; Kamp, C.; Reijerse, E.; Happe, T.; Lubitz, W. *Biochemistry* **2009**, 48 (33), 7780–7786.
- (21) Cheah, M. H.; Tard, C.; Borg, S. J.; Liu, X.; Ibrahim, S. K.; Pickett, C. J.; Best, S. P. J. Am. Chem. Soc. 2007, 129 (36), 11085–11092
- (22) Olsen, M. T.; Rauchfuss, T. B.; Wilson, S. R. J. Am. Chem. Soc. **2010**, 132 (50), 17733–17740.
- (23) Tschierlei, S.; Ott, S.; Lomoth, R. Energy Environ. Sci. 2011, 4 (7), 2340–2352.

(24) Nicolet, Y.; Cavazza, C.; Fontecilla-Camps, J. C. J. Inorg. Biochem. 2002, 91 (1), 1–8.

- (25) Yang, J. Y.; Chen, S. T.; Dougherty, W. G.; Kassel, W. S.; Bullock, R. M.; Dubois, D. L.; Raugei, S.; Rousseau, R.; Dupuis, M.; Dubois, M. R. Chem. Commun. 2010, 46 (45), 8618–8620.
- (26) Silakov, A.; Shaw, J. L.; Reijerse, E. J.; Lubitz, W. J. Am. Chem. Soc. 2010, 132 (49), 17578–17587.
- (27) Erdem, Ö. F.; Schwartz, L.; Stein, M.; Silakov, A.; Kaur-Ghumaan, S.; Huang, P.; Ott, S.; Reijerse, E. J.; Lubitz, W. *Angew. Chem., Int. Ed.* **2011**, *50* (6), 1439–1443.
- (28) Silakov, A.; Wenk, B.; Reijerse, E.; Albracht, S. P.; Lubitz, W. J. Biol. Inorg. Chem. **2009**, 14 (2), 301–313.
- (29) Justice, A. K.; Rauchfuss, T. B.; Wilson, S. R. Angew. Chem., Int. Ed. 2007, 46 (32), 6152–6154.
- (30) Justice, A. K.; Zampella, G.; De Gioia, L.; Rauchfuss, T. B.; van der Vlugt, J. I.; Wilson, S. R. *Inorg. Chem.* **2007**, *46* (5), 1655–1664.
- (31) Justice, A. K.; De Gioia, L.; Nilges, M. J.; Rauchfuss, T. B.; Wilson, S. R.; Zampella, G. *Inorg. Chem.* **2008**, 47 (16), 7405–7414.
- (32) Olsen, M. T.; Barton, B. E.; Rauchfuss, T. B. *Inorg. Chem.* **2009**, 48 (16), 7507–7509.
- (33) Siegbahn, P. E. M.; Tye, J. W.; Hall, M. B. Chem. Rev. 2007, 107 (10), 4414–4435.
- (34) Reijerse, E.; Lendzian, F.; Isaacson, R.; Lubitz, W. J. Magn. Reson. 2012, 214, 237–243.
- (35) Epel, B.; Arieli, D.; Baute, D.; Goldfarb, D. J. Magn. Reson. 2003, 164 (1), 78-83.
- (36) Gütlich, P.; Bill, E.; Trautwein, A. X. Mossbauer Spectroscopy and Transition Metal Chemistry: Fundamentals and Applications; Springer-Verlag: Berlin, 2011.
- (37) Neese, F. Wiley Interdiscip. Rev.—Comput. Mol. Sci. 2012, 2, 73–78.
- (38) Perdew, J. P. Phys. Rev. B 1986, 33 (12), 8822.
- (39) Becke, A. D. Phys. Rev. A 1988, 38 (6), 3098.
- (40) Schäfer, A.; Horn, H.; Ahlrichs, R. J. Chem. Phys. 1992, 97 (4), 2571–2577.
- (41) Lee, C. T.; Yang, W. T.; Parr, R. G. Phys. Rev. B 1988, 37 (2), 785-789.
- (42) Becke, A. D. J. Chem. Phys. 1993, 98 (7), 5648-5652.
- (43) Wachters, A. J. J. Chem. Phys. 1970, 52 (3), 1033.
- (44) Filatov, M. J. Chem. Phys. 2007, 127, 8.
- (45) Sinnecker, S.; Slep, L. D.; Bill, E.; Neese, F. Inorg. Chem. 2005, 44 (7), 2245–2254.
- (46) Römelt, M.; Ye, S.; Neese, F. Inorg. Chem. **2009**, 48 (3), 784–785.
- (47) Volkers, P. I.; Boyke, C. A.; Chen, J.; Rauchfuss, T. B.; Whaley, C. M.; Wilson, S. R.; Yao, H. *Inorg. Chem.* **2008**, 47 (15), 7002–7008.
- (48) Epel, B.; Poppl, A.; Manikandan, P.; Vega, S.; Goldfarb, D. J. Magn. Reson. 2001, 148 (2), 388-397.
- (49) Epel, B.; Niklas, J.; Antonkine, M.; Lubitz, W. Appl. Magn. Reson. **2006**, 30 (3–4), 311–327.
- (50) Doan, P. E. J. Magn. Reson. 2011, 208 (1), 76-86.
- (51) Roos, B. O.; Lindh, R.; Malmqvist, P.-A.; Veryazov, V.; Widmark, P. O. J. Phys. Chem. A 2005, 109 (29), 6575–6579.
- (52) Liu, T.; Darensbourg, M. Y. J. Am. Chem. Soc. 2007, 129 (22), 7008-7009.
- (53) Lawrence, J. D.; Li, H.; Rauchfuss, T. B.; Benard, M.; Rohmer, M. M. Angew. Chem., Int. Ed. **2001**, 40 (9), 1768–1771.
- (54) Chen, Z. J.; Lemon, B. J.; Huang, S.; Swartz, D. J.; Peters, J. W.; Bagley, K. A. *Biochemistry* **2002**, *41* (6), 2036–2043.
- (SS) Silakov, A.; Reijerse, E. J.; Lubitz, W. Eur. J. Inorg. Chem. 2011, 1056–1066.
- (56) Giles, L. J.; Grigoropoulos, A.; Szilagyi, R. K. Eur. J. Inorg. Chem. 2011, 2677–2690.
- (57) Popescu, C. V.; Münck, E. J. Am. Chem. Soc. 1999, 121 (34), 7877–7884.
- (58) Pereira, A. S.; Tavares, P.; Moura, I.; Moura, J. J. G.; Huynh, B. H. J. Am. Chem. Soc. **2001**, 123 (12), 2771–2782.