Fe(NTA)-catalyzed Dioxygenation of 4-t-Butylcatechol and the Mechanism of Non-heme Ferric Dioxygenases

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Abstract

Fe(NTA), activating the 'substrate' 4-t-butyl- Γ c(Γ), activating the substrate \pm -outyrcatechole, represents a functional active centre analogue of non-heme ferric dioxygenases. A Fe(NTA) catecholate complex with monodentate catecholate
is the reactive species to undergo dioxygenation.

Introduction

 $\frac{1}{2}$ is catalyzed oxidation of category $\frac{1}{2}$ $\frac{1}{2}$ reflic foll catalyzed oxidation of catechois $\frac{1}{2}$ with $O₂$ yields significantly different products in enzymic $(e.g.$ catechol 1,2-dioxygenase) or nonenzymic reactions, respectively. In both mechanisms, catecholate is initially coordinated to iron(III). However, catecholate chelate ligands in ferric complexes, succh as 3,5-di-t-but as 3,5-di-t-butyle-category-category-category-category-category-category-category-categoryt-Bu), undergrounder that the step with \mathbb{R} is \mathbb{R} in the step via \mathbb{R} t-Bu), undergo stepwise oxidation with oxygen via semiquinone to quinone (II) $[1-3]$, whereas at the non-heme ferric-active centre of catechol 1,2-dioxygenase, catechol (I), $R = H$) is oxidatively cleaved to muconic acid (III). Thus in the enzyme a 'simple' redox reaction is replaced by a dioxygenation, lead- $\frac{1}{100}$ in the dicarbonic action is replaced by a dioxygenation, readmetabolic actus which will lit lifts further metabolic pathways. This requires an as yet unknown activation of catecholate and/or O_2 at the non-heme ferric centre of the enzyme.

 $E =$ Catechol 1,2-Dioxygenase

We have shown [2, 51 that [Fe(NTA)(DBcat)12 we have shown [2, 3] that $[\text{Fe}(\text{N1A})(\text{D}\text{Ca})]$ $(NTA = nitrilotriacetate; DBcat = 3,5-di-t-butyl$ catecholate), as a first iron(III)-dioxygenase model system, reacts with O_2 effecting ring cleavage of the catecholate ligand. We now wish to demonstrate a similar reactivity of the analogous complex with the

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less oxidizable 4-t-butylcatechol $(BcatH₂)$ (VI); as condizable τ -courtrated for μ both η (vi), meenaman is presen

Experimental

 \mathbf{A} l reactions with out identifications with out identifications with out identifications with \mathbf{A} An reactions with oxygen were called out fuentically. A typical example is given below: to 10 mmol $BcatH₂$ in 100 ml ethanol, 115 ml 0.6 M borate buffer pH 8 and a solution of 10 mmol each Fe- $(CIO₄)₃$ and NTANa₃ in 30 ml ethanol/water 1:1 were added. The deeply blue-green solution (showing a glass electrode reading of 'pH' 8.2) was stirred under O_2 . After ten days, having consumed 10 mmol $O₂$, the solution turned reddish-brown. Evaporation of ethanol was followed by $CH₂Cl₂$ -extraction of first the basic and then the acidified (H_2SO_4) aqueous phase. EPR spectra were run on a Varian E109 spectrometer; temperature, 77 K; microwave frequency, 9.24 GHz; modulation amplitude, 10 G; microwave power, 50 mW. The signals were observed both in the frozen reaction medium $(H₂O/EtOH)$, and in frozen glasses (e.g. H_2O /propylene glycol).
For interpretation of the EPR-data see [6].

Results and Discussion

Oxidation of $[Fe(NTA)(DBcat)]^{2-}$ by O₂ in a σ_{A} buffer at pH σ_{B} buffer at pH σ_{B} at pH σ α are builet at β fricative total interviewing products (relative to BcatH₂ as starting material, total yield 99%): 62% lactone ester $(37\%$ (IVa) and 25% (Va)), 28% lactonic acid (9% (IVb) and 19% (Vb)), and 10% unreacted BcatH₂. All lactones (IV) and (V) are

IVb:R=H Vb:R=H

TABLE I. Oxidation of 4-t-Butylcatechol by O_2 .⁸

	No. Molar ratio			Time	Product, $\boldsymbol{\%}^{\mathbf{b}}$		
	Bcat	NTA Fe		[d]	Bcat (VI)	(VII)	Quinone Lactones $(V) + (V)$
				10	10		89
2	10	1.3	1	-20	18	3	71
3	100	1.3	1	20	35	15	45
4	3			5	30	60	
5				20	80	10	

^aIn EtOH/borate buffer 1:1, 'pH' 8.2 (see text). $\mathbf{^{b}$ Based on original BcatHz.

secondary products of muconic acid derivatives (XIII) [7], obtained from catecholate by dioxygenation. The total yield of ring cleavage products was 89% (Table I, No. 1). No alternative oxidation pathways were discernable, e.g. no quinone **(VII)** could be detected.

Thus, 4-t-butylcatechol in the ternary complex $[Fe(NTA)(DBcat)]^{2-}$ undergoes the same reaction as catecholate at the ferric active centre of catechol 1,2-dioxygenase. A $10-$ to 100-fold excess of catechol vs. Fe(NTA) (Table I, No. 2 and 3) gives rise to 71%, and 45% ring cleavage, respectively; i.e., Fe- (NTA) catalyzes, albeit slower than the enzyme, the catechol dioxygenation.

Control experiments demonstrate quinone formation instead of ring cleavage in the absence of NTA, i.e., upon oxidation of simple ferric catecholate complexes (Table I, No. 4). Further, it is shown (Table I, No. 5) that 'free' catecholate in alkaline medium is protected from oxidation by boric acid via ester formation [8].

 $[FeNTA)(DBcat)]^{2-}$ (VIII) was isolated as its tetrabutylammonium salt (dark blue-green crystals) and characterized as a catecholate chelate complex. The data are similar to those of the corresponding complex $[Fe(NTA)(DBcat)] (pipH)_2$ [2] (M. G. Weller and C. Ruh, unpublished). However, catechelates chelated to ferric ions are known to be either non-oxidizable $[4, 9-11]$ or to undergo oxidation to quinone (VII) via semiquinone ($[12, 12]$ 13]; $cf.$ Table I, No. 4); they are not subject to dioxygenation. Some catecholate complexes of other transition metal ions undergo non-catalytic ring cleavage with $O₂$ [2, 4].

A dioxygenase mechanism was proposed [4] involving the attack of $O₂$ on catecholate bound to the ferric active centre as a monodentate $\frac{1}{2}$ (see (IV). This high density of negative groups around Fe3+ in the chelate complex **(VIII)** coups around re the constant complex (vers) with monodentate catecholate **(IX).** In fact, the EPR-spectrum of **(VIII)** contains signals of *two high* spin ferric complexes; one with essentially rhombic symmetry of the electronic environ- μ memore symmetry of the electronic change. $\frac{64}{94}$ middle Kramer's doublet; $\alpha = 0.15$, lower $\begin{bmatrix} 0.04 \\ 0.04 \end{bmatrix}$, and a more axial species $(E/D = 0.000$ $(100; \text{g}) = 7.95, \text{g} = 5.64$. According to the b_6 by b_7 , b_8 b_7 , b_8 b_9 , b_9 , b_9 b_1 , b_1 , b_1 , b_1 , b_1 , b_1 , b_2 , b_3 , b_1 broad, rhombic signal of solid $[Fe(NTA) (DBcat)]$ -
(Bu₄N)₂ (g_{eff} = 4.2), the 'rhombic' component corresponds to the chelate compound (VIII), and the 'axial' one to (IX), *i.e.,* with monodentate catecholate. Also a perfectly rhombic species (E/D = $1/3$; $g_x = g_y = g_z = 4.29$, middle Kramer's doublet) $\frac{1}{2}$, $\frac{1}{2}$, eq. concentration of Fe(NTA)aq. **(IX)** is readily eq. concentration of $Fe(NTA)aq$. (IX) is readily ketonized to (X), thus allowing O_2 to attack the aromatic ring, facilitated by an iron-oxygen interaction (XI). Iron catecholate complexes with

Scheme 1

all coordination sites *cis* to catecholate occupied, thus preventing the formation of an additional Fe-O bond, e.g., Fe(salen)(catH) [4], do not carry out dioxygenation. Insertion of oxygen to yield the anhydride (XII), ring opening to muconic acid **(XIII)** and lactonization finally leads to (IV) or (V) as products.

The ternary complex (VIII), unlike the binary catecholate complex (Table I, No. 4), is not subject to oxidation via semiquinone to quinone (VII), although the relative rates of reaction (see Table I) would favour oxidation against dioxygenation. Supposedly, 'hardening' of Fe³⁺ by the hard ligand $NTA³⁻$ disallows the electron shuttling through iron, *i.e.*, intermediate formation of Fe²⁺, which is a prerequisite of semiquinone complex formation.

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References

- R. R. Grinstead, *Biochemistry, 3, 1308 (1964).*
- *M. G.* Weller and U. Weser,J. *Am. Chem. Sot., 104. 3152 (1982).*
- D. G. Brown and W. L. Johnson, III, Z. *Naturforsch., TeilB:, 34, 112 (1979).*
- L. Que, Jr., *Struct. Bonding (Berlin), 40, 39* (1980); *Coord. Chem Rev., 50, 73* (1983).
- M. G. Weller, in I. Bertini, R. S. Drago and C. Luchinat (eds.), 'The Coordination Chemistry of Metalloenzymes', Reidel, Dordrecht, 1983, p. 273.
- J. W. Whittaker, J. D. Libscomb, T. A. Kent and E. Miinck, J. *Biol. Chem., 259, 4466 (1984).*
- *Y.* Sawaki and C. S. Foote, J. *Am.* Chem. Sot., 105, 5035 (1983).
- U. Weser, *Hoppe-Seyler's Z. Physiol.* Chem., 349, 982 (1968);
- **9 Barrish Life C. J. Carrano, S. R. Cooper, S. R. Corporation** L. Babcock and R. Pizer, *Inorg. Chem.*, 22, 174 (1983).
- 10 B. Howlin, A. R. Mohd-Nor, J. Silver and P. W. C. A. Avdeef, J. V. McArdle and K. N. Raymond, *J. Am. Chem. Sot., 101, 6097 (1979).*
- 11 D. A. Buckingham, C. R. Clark, M. G. Weller and G. J. Barnard, *Inorg. Chim. Acta*, 91, 153 (1984).
- Gainsford, *J. Chem. Soc., Chem. Commun.*, 779 (1982).
- 12 M. W. Lynch, M. Valentine and D. N. Hendrickson, J. 1¹ International Leon and D. T. Sawyer, *Incre Chem.*, **Camera** *Am. Chem. Sot., 104, 6982* (1982).
- *21, 3692 (1982)* and refs. therein.