A NMR Study of Lanthanide-NOTA Chelates as Aqueous Paramagnetic Shift and Relaxation Probes*

C. F. G. C. GERALDES[†], M. P. M. MARQUES

Chemistry Department, University of Coimbra, 3000 Coimbra, Portugal

and A. D. SHERRY

Department of Chemistry, The University of Texas at Dallas P.O. Box 830688, Richardson, Tex. 75083-0688, U.S.A.

One of the major limitations in the use of paramagnetic lanthanide(III) chelates as NMR aqueous shift reagents to probe dynamic molecular structures [1, 2] has been their lack of structural or effective axial symmetry [3, 4]. In an effort to optimize the characteristics of water soluble NMR shift and relaxation reagents, we have recently [5] studied the paramagnetic shifts and solution structure of the Ln(NOTA) chelate series in aqueous solution near pH 7 (Ln = lanthanide; NOTA is the axially symmetric macrocyclic triaza ligand 1,4,7-triazacyclononane-N, N', N''-triacetic acid) and its complexes with the monocarboxylate ligand cyclopropane carboxylic acid [6]. In this work we report a similar study of the binding of the Ln(NOTA) chelates to the flexible mononucleotide adenosine 5'-monophosphate (5'-AMP) which contains one phosphate binding group, and to the rigid endo-cis-bicyclo-[2.2.1] hept-5-ene-2,3-dicarboxylic acid (DCA), which has two carboxylate groups oriented in a favorable way to bind lanthanide chelates (Fig. 1). The symmetry of the dipolar shifts and the geometry of the adducts formed are discussed.



Fig. 1. Chemical structure and numbering scheme of adenosine 5'-monophosphate (5'-AMP) and *endo-cis*-bicyclo-[2.2.1] hept-5-ene-2,3-dicarboxylic acid (DCA).

Experimental

The preparation of NOTA and the 1:1 Ln(NOTA) complexes have been described previously [5]. 5'-AMP and DCA were obtained from Sigma and Aldrich, respectively. The NMR spectra were obtained on either a Varian XL-200 or JEOL FX-200 spectrometer. T_1 measurements were made using the standard inversion-recovery sequence.

Results and Discussion

LIS studies on 5'-AMP, previously undertaken using aqueous Ln³⁺ cations and Ln(EDTA) chelates [7] (EDTA is ethylenediaminetetraacetic acid), were extended to the entire Ln(NOTA) chelate series. Exchange broadening of the proton 5'-AMP resonances was observed at 25 °C for all Ln(NOTA) chelates; the early Ln(NOTA)-AMP complexes appear to be in rapid exchange at 85 °C but the heavy Ln-(NOTA)-AMP complexes are in slow exchange at all temperatures. The Ln(EDTA) chelates cause extensive exchange broadening at 25 °C only for the latter half of the series and not at all at 85 °C [8]. The ³¹P resonances show similar exchange broadening. The exchange rate differences between the two mixed complexes, Ln(NOTA)-AMP versus Ln(EDTA)-AMP, must reflect the larger association constants in the former complexes and hence longer bound lifetimes, $\tau_{\rm M}$. Thus, the Ln(EDTA)-AMP complexes come closer to meeting fast exchange conditions $(w_M \tau_M \ll 1)$ at a given temperature than do the Ln(NOTA)-AMP complexes.

Table I compares the proton shift ratios determined for 5'-AMP protons for the light Ln(NOTA) and Ln(EDTA) chelates at 85 °C. Since conditions of fast exchange apply at this temperature, the shift ratios were obtained in the usual way [7] from NMR titration curves of 5'-AMP with the chelates. The two sets of data agree with each other quite well and both agree with calculated ratios based on axially symmetric dipolar shifts determined from an average 5'-AMP conformation previously described [7] involving chelation at the phosphate groups. Table I also shows the measured proton spin-lattice relaxation rate ratios induced by binding of Gd(NOTA) and Gd(EDTA), which depend on the inverse sixth power of the distances of the protons to the paramagnetic center. The overall data generally support the 5'-AMP adduct structure described previously [7].

The mixed complexes formed by the dicarboxylate ligand DCA with aqueous Ln^{3+} cations and various chelates, including Ln(EDTA) and Ln-(HEDTA) (HEDTA is *N*-hydroxyethylenediamine-

^{*}Paper presented at the Second International Conference on the Basic and Applied Chemistry of f-Transition (Lanthanide and Actinide) and Related Elements (2nd ICLA), Lisbon, Portugal, April 6-10, 1987.

[†]Author to whom correspondence should be addressed.

Perturbation	Lanthanide chelate	H ₈	H ₂	H _I ′	H ₂ ′	H _{3'}	H ₄ ′
Shift	Pr(NOTA)	36	-4	15	34	44	36
	Nd(NOTA)	30	5	5	30	32	27
	Eu(NOTA)	42	2	13	22	b	b
	Pr(EDTA)	32	-1	7	24	35	31
	Nd(EDTA)	30	2	10	25	35	28
	Eu(EDTA)	30	-1	8	28	40	32
	Calculated ^c (axial)	32	1	13	28	38	33
Relaxation	Gd(NOTA)	97	5	7	b	b	b
	Gd(EDTA)	47	5	7	8	15	19
	Calculated ^c	41	2	3	10	9	13

TABLE I. Lanthanide-induced Proton Shift and Relaxation Ratios^a for 0.05 M 5'-AMP at pH 7.5 and 85 °C, using Ln(NOTA) and Ln(EDTA Chelates as Probes

^aRelative to $H_{5',5''} = 100$. ^bNot measured, ^cFrom ref. 7.

tetraacetic acid) have been studied previously using LIS and relaxation techniques [9]. We extended these studies to obtain ¹H and ¹³C LIS and relaxation data using the Ln(NOTA) chelates.

The ¹H spectra of the Ln(NOTA)–DCA complexes at 25 °C indicate that the Ce \rightarrow Eu complexes are in fast exchange whereas the Tb \rightarrow Yb complexes show exchange broadening even at 85 °C. For ¹³C nuclei, fast exchange conditions applied at 25 °C throughout the entire lanthanide series. Titration curves were consistent with formation of 1:1 Ln(NOTA)–DCA adducts which show increased stability over the corresponding Ln(EDTA)–DCA adducts [9]. Shift ratios and relaxation enhancements for ¹H and ¹³C nuclei are given in Tables II and III, respectively.

The ratios of induced relaxation rates obtained with Gd(NOTA) and avariety of Ln(HEDTA) chelates (including Gd) are quite similar and agree well with relaxation ratios estimated from a structure for the DCA-lanthanide chelate adduct as described in the literature [9]. ¹³C relaxation rate ratios induced by Gd(NOTA) also agree quite well with calculated values obtained, in a similar way, from molecular models (Table III). This indicates that DCA forms adducts of very similar structure with all lanthanide complexes studied, including the present NOTA chelates. Therefore, the large variations in shift ratios obtained for ¹³C LIS values (Table III) do not result from variations in molecular geometry [9]. They could instead result from (a) contact contribution to the measured LIS; (b) non-axial symmetry of the dipolar shifts; or (c) variations of the direction of the axial symmetry axis along the lanthanide cation series. Previous workers [9] have shown that (c) does not apply and proposed (b) as the explanation for the observed changes in proton shift ratios along the lanthanide series. Unfortunately, we were not able to measure proton shift ratios for all Ln(NOTA)-

Perturbation	Lanthanide chelate	H ₂ , H ₃	H ₁ , H ₄	H ₅ , H ₆	H ₇		H ₈
Shift	Ce(NOTA)	100	29	28		35b	
	Pr(NOTA)	100	46	36		36 ^b	
	Nd(NOTA)	100	42	19	31		27
	Sm(NOTA)	100	16	22		21 ^b	
	Pr(EDTA) ^c	100	42	22	39		28
	Calculated ^c (axial)	100	42	25	43		32
Relaxation	Gd(NOTA)	100	41	70		18 ^b	
	Gd(HEDTA) ^c	100	37	63	18		14
	Calculated ^c	100	30	65	16		12

TABLE II. Lanthanide-induced Proton Shift and Relaxation Ratios^a for 0.05 M DCA at pH 7.6 and 25 °C, using Ln(NOTA) Chelates as Probes

^aRelative to $H_{2,3} = 100$. ^bAverage value for H_7 and H_8 . ^cFrom ref. 9.

TABLE III. Lanthanide-induced ¹³C Shift and Relaxation Ratios^a for 0.05 M DCA at pH 7.6 and 25 °C, using Ln(NOTA) Chelates as Probes

Perturbation	Lanthanide chelate	C ₁ , C ₄	C ₇	C ₂ , C ₃	C ₅ , C ₆	CO ₂
Shift	Ce(NOTA)	40	16	100	14	90
	Pr(NOTA)	26	24	100	23	77
	Nd(NOTA)	7	10	100	6	- 30
	Eu(NOTA)	13	13	100	7	-191
	Tb(NOTA)	155	41	100	123	498
	Dy(NOTA)	108	70	100	91	420
	Ho(NOTA)	220	122	100	175	835
	Er(NOTA)	-100	53	100	- 93	-66
	Tm(NOTA)	-70	37	100	-51	220
	Yb(NOTA)	0	85	100	10	195
Corrected	$(Ce \rightarrow Eu)$	40	24	100	20	312
dipolar shift	$(Tb \rightarrow Yb)$	42	38	100	33	220
	Calculated (axial)	50	36	100	32	220
Relaxation	Gd(NOTA)	39	33	100	46	833
	Calculated	30	14	100	40	745

^aRelative to $C_{2,3} = 100$

DCA complexes because of exchange broadening. Nevertheless, the sign and magnitude of the ¹³C LIS values, especially for those nuclei near the lanthanide binding site on DCA, indicate that the contact contribution to the LIS values cannot be zero. Using standard procedures [10], we separated the contact and dipolar contributions to the measured LIS values. The proton LIS values for Ce \rightarrow Sm were found to be overwhelmingly dipolar, and the LIS ratios agreed quite well with the ratios calculated from the published structure using the axial symmetry model (Table II). The analysis of the ¹³C data, after purification of the contact contribution to each shift, gave similarly good agreement with the axial symmetry model (Table III).

In conclusion, dipolar shifts induced by Ln(NOTA) chelates compare quite well with values calculated assuming axial symmetry and the previously defined solution structures for the 5'-AMP and DCA adducts of lanthanide chelates [7, 9]. This contrasts with non-axial contributions to the LIS values observed in some cases when non-axially symmetric chelates like Ln(EDTA) or Ln(HEDTA) have been used [9]. Therefore, the fact that the axial symmetry of the Ln(NOTA) chelates [5] is preserved when these form adducts with mono- [8] or even polyfunctional ligands like DCA indicates that the symmetry of NOTA determines the overall symmetry of the dipolar induced shifts on the second ligand. This observation makes Ln(NOTA) an attractive series of chelates to be used as aqueous shift probes of molecular structure. Their main limitations seem to result from extensive problems of exchange broadening for the second half of the lanthanide series, caused by stronger interactions with phosphate or carboxylate groups than what is found, e.g., with Ln(EDTA) chelates.

Acknowledgements

This work was supported in part by grants from the Robert A. Welch Foundation (AT-584) and Mallinckrodt, Inc. Support from I.N.I.C., Portugal, is also acknowledged.

References

- 1 C. M. Dobson and B. A. Levine, New Tech. Biophys. Cell Biol., 3, 19 (1976).
- 2 R. J. P. Williams, Struct. Bonding (Berlin), 50, 79 (1982).
- 3 W. D. Horrocks Jr., J. Am. Chem. Soc., 96, 3022 (1974).
- 4 J. D. Marinetti, G. H. Snyder and B. D. Sykes, *Biochemistry*, 15, 4600 (1976).
- 5 A. D. Sherry, M. Singh and C. F. G. C. Geraldes, J. Magn. Reson., 66, 511 (1986).
- 6 C. F. G. C. Geraldes, M. Singh and A. D. Sherry, J. Less-Common Met., 112, 255 (1985).
- 7 C. M. Dobson, C. F. G. C. Geraldes, R. G. Ratcliffe and R. J. P. Williams, *Eur. J. Biochem.*, 88, 259 (1978).
- 8 A. D. Sherry, C. A. Stark, J. R. Ascenso and C. F. G. C. Geraldes, J. Chem. Soc., Dalton Trans., 2078 (1981).
- 9 M. Delepierre, C. M. Dobson and S. L. Menear, J. Chem. Soc., Dalton Trans., 678 (1981).
- 10 C. N. Reilley, B. W. Good and R. D. Allendoerfer, Anal. Chem., 48, 1446 (1976).