# **Transport Properties of the Binary System L-(+)-Tartaric** Acid–Water at 25 °C. A Velocity Correlation Study

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Accurate mutual diffusion and intradiffusion coefficients have been measured for the binary system L-(+)-tartaric acid (2)-water (1) at 25 °C. The collected diffusion coefficients have been combined with osmotic coefficients present in the literature to calculate the velocity correlation coefficients (VCC's). The results have been interpreted in terms of molecular interactions.

#### Introduction

This research is part of a program devoted to the study of equilibrium and transport properties of the quaternary system ethanol-glucose-L-(+)-tartaric acid-water, with the aim to test the theory that some of us developed in the past (Vitagliano et al., 1992), giving criteria for both static and dynamic gravitational stability of diffusion boundaries with four (or more) components. In the preliminary phase of this study we have been collecting a set of data on the corresponding binary and ternary systems. We have already analyzed the system glucose-water (Castaldi et al., 1998).

In this work, we remeasured densities for the binary system L-(+)-tartaric acid (2)-water (1) in the composition range approaching saturation. Furthermore, for the same system, we measured mutual diffusion and intradiffusion coefficients. These data are not present in the literature, to the authors' knowledge.

The data have been discussed in terms of velocity correlation coefficients (VCC's), that show how the macroscopic thermodynamic properties are affected by the solution structure at the molecular scale.

L-(+)-Tartaric acid is a weak diprotic acid [ $K_1 = 9.268 \times 10^{-4}$  mol dm<sup>-3</sup> and  $K_2 = 4.305 \times 10^{-5}$  mol dm<sup>-3</sup> (Robinson and Stokes, 1955)]. Figure 1 shows the fraction of the various species present in solution, computed using these  $K_i$  values, drawn as a function of the square root of the stoichiometric tartaric acid molality.

The experimental techniques allow an exhaustive examination in the region where the dissociated species concentration is not appreciable (m > 0.09 mol kg<sup>-1</sup>), although measurements were also made in the more dilute composition range.

#### **Experimental Section**

**Materials.** L-(+)-Tartaric acid purchased from Sigma Chemical Co. (99.5% purity) was used without further purification. All solutions were prepared by mass using double-distilled water. In all calculations the molecular weight of L-(+)-tartaric acid was assumed to be 150.1 g mol<sup>-1</sup>.

**Density Measurements.** A set of density measurements were taken in the molality range  $0-9 \mod \text{kg}^{-1}$  with an Anton Paar 602 densimeter.

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**Figure 1.** Fraction of the various species in aqueous solutions of L-(+)-tartaric acid as computed from the  $pK_a$  (Robinson and Stokes, 1955): **1**, AcH<sub>2</sub>; **2**, AcH<sup>-</sup>; **3**, Ac<sup>--</sup>; **4**, H<sup>+</sup>.

Table 1. Densities of Aqueous L-(+)-Tartaric Acid Solutions at 25  $^{\circ}\text{C}$ 

$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	<i>m</i> / mol kg <sup>-1</sup>	$^{ ho/}_{ m kg~dm^{-3}}$	<i>m</i> / mol kg <sup>-1</sup>	$^{ ho/}_{ m kg~dm^{-3}}$	<i>m</i> ∕ mol kg <sup>−1</sup>	$^{ ho/}$ kg dm <sup>-3</sup>
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0.0000 0.0069 0.0932 1.4587 1.4614 1.5343 2.4070 2.5067	0.997 044 0.997 53 1.003 32 1.082 76 1.082 16 1.085 77 1.126 34 1.130 67	3.5241 4.4529 4.4577 4.5056 4.5215 4.5261 5.4207 5.4207 5.4207	1.170 85 1.202 68 1.202 76 1.204 26 1.204 76 1.204 74 1.204 81 1.231 70 1.232 49	6.0388 6.4237 6.5736 6.9892 7.3771 7.9845 8.7506 9.0057 9.0057	1.248 29 1.258 14 1.261 77 1.271 32 1.280 63 1.293 31 1.307 95 1.312 86

The temperature of the densimeter was regulated at  $(25.00 \pm 0.01)$  °C. For the densimeter calibration, air (at measured pressure and humidity) and distilled water, assumed density 0.997 044 kg dm<sup>-3</sup> (Lo Surdo et al., 1982), were chosen. The data, collected in Table 1, are in very good agreement with previous literature data (Thomsen, 1885; Dunstan and Thole, 1908). The experimental error is within  $(1-2) \times 10^{-5}$  kg dm<sup>-3</sup>. The coefficients of the polynomial equation fitted to the densities are given in Table 2. Figure 2 shows the deviation of experimental data from this equation. The limiting partial molar volume of undissociated L-(+)-tartaric acid was computed by interpolating the apparent molar volume of solute with a three-term polynomial, neglecting the first two experimental

Table 2. Coefficients of the Polynomial Equations Fitted to the Experimental Density and Diffusion Data (= $A_0 + A_1m + A_2m^2 + A_3m^3 + ... \pm \sigma$ )

	$ ho/{ m kg}~{ m dm}^{-3}$	$10^5 D_{12}/\mathrm{cm}^2 \mathrm{~s}^{-1}$	$10^5 D_2^*/{ m cm}^2~{ m s}^{-1}$	$10^5 D_1^*/{ m cm}^2~{ m s}^{-1}$
$A_0$ $A_1$	0.997044 (668.3 + 2.4) × 10 <sup>-4</sup>	$0.701 \pm 0.013 \ (-5.80 \pm 0.60)  imes 10^{-2}$	$\begin{array}{c} 0.798 \pm 0.009 \\ -0.208 \pm 0.010 \end{array}$	$2.299 \\ -0.72 \pm 0.02$
$A_2$	$(-67.1 \pm 2.0) \times 10^{-4}$ $(-67.1 \pm 5.0) \times 10^{-5}$	$(1.13 \pm 0.57) \times 10^{-3}$	$(2.2 \pm 0.3) \times 10^{-2}$	$0.102 \pm 0.007$ ( 5.2 + 0.7) + 10-3
$A_3 \\ A_4$	$(62.1 \pm 5.6) \times 10^{-6}$ $(-40.5 \pm 7.3) \times 10^{-6}$		$(-8 \pm 2) \times 10^{-4}$	$(-5.3 \pm 0.7) \times 10^{-5}$
$A_5 \sigma$	$egin{array}{c} (12.0\pm3.1) imes10^{-7}\ 1.4 imes10^{-4} \end{array}$	0.0105	0.0086	0.037



**Figure 2.** Comparison of L-(+)-tartaric acid aqueous solutions densities from different sources with eq 1:  $\bullet$ , our data;  $\diamond$ , (Thomsen, 1885);  $\Box$ , (Dunstan and Thole, 1908).

data, referring to solutions in which dissociation was appreciable. The computed value is:  $V_2^{\circ}(AcH_2) = 83.44 \pm 0.08 \text{ cm}^3 \text{ mol}^{-1}$ .

*Mutual Diffusion Coefficients.* Mutual diffusion coefficients were measured with a Gouy diffusiometer (Gosting, 1950; Tyrrell and Harris, 1984) using a two-lens apparatus in which parallel light passing through a single-channel cell generates the fringe pattern on its focus. The initial boundary was formed with the siphoning technique. The light source was a Unifas PHASE 0.8-nW neon-helium laser operating at  $\lambda = 632.8$  nm. A Model "II fx" MacIntosh computer was used to control the scanning apparatus and to determine fringe positions from fringe intensity profiles.

The mutual diffusion coefficients  $D_{12}$  were calculated using a series of programs well described in the literature (Albright and Miller, 1989; Miller et al., 1992).

The experimental data, in the molality range 0–9 mol kg<sup>-1</sup>, are collected in Table 3. The experimental error is within  $(1-2) \times 10^{-3}$  cm<sup>2</sup> s<sup>-1</sup>. The coefficients of the polynomial equation fitted to the mutual diffusion coefficients in the molality range where the tartaric acid dissociation is negligible are given in Table 2. This equation was computed by least squares excluding the four diffusion data measured in the most diluted solutions where dissociation is appreciable. The experimental data and the polynomial trend are shown in Figure 3. The constant term,  $0.701 \times 10^{-5}$ , can be assumed as the limiting diffusion coefficient of undissociated tartaric acid,  $D_{12}^{\circ}$ (AcH<sub>2</sub>).

**Intradiffusion Measurement.** The intradiffusion measurements were made using the pulsed gradient spin–echo (PGSE) FT-NMR method (Stilbs, 1987; Callaghan, 1991; Weingärtner, 1994). Experiments were carried out on a Varian FT 80 NMR spectrometer operating in the <sup>1</sup>H mode, equipped with a pulsed magnetic field gradient unit, specially made by Stelar (Mede, Italy). The temperature was controlled to within  $\pm 0.1$  °C with a Stelar variable-temperature controller model VTC87. By using a pulse sequence where the echo delays are fixed and only the gradient pulse lengths are varied, the effects of relaxation

Table 3. Diffusion Data for the System L-(+)-Tartaric Acid–Water at 25  $^\circ C^a$ 

<i>m</i> ∕ mol kg <sup>−1</sup>	$\Delta m/$ mol kg $^{-1}$	J <sub>m</sub>	$10^5 D_{12}/ \ { m cm}^2 \ { m s}^{-1}$	$B(x_2)$	$10^5 D_{ m T}/\ { m cm}^2 { m s}^{-1}$
0.0000			1.171	1.000	1.171
0.0318	0.0623	45.46	0.783	1.004	0.779
0.0500	0.0862	62.16	0.776	1.007	0.771
0.2506	0.0602	40.01	0.731	1.035	0.706
0.6000	0.0719	51.40	0.695	1.086	0.640
0.7488	0.0605	37.31	0.669	1.108	0.603
1.4966	0.0755	41.49	0.612	1.222	0.501
2.4568	0.0997	45.65	0.560	1.371	0.409
3.4729	0.1023	39.94	0.511	1.524	0.335
4.4872	0.0687	21.64	0.451	1.663	0.271
5.5093	0.1773	55.56	0.429	1.782	0.241
6.4987	0.1499	36.66	0.363		
7.4841	0.2141	48.26	0.347		
8.4875	0.1604	32.25	0.298		
9.4992	0.1744	29.14	0.239		

<sup>*a*</sup> *m*, average molality of each diffusion run.  $\Delta m$ , molality difference between bottom and top solutions.  $J_m$ , total number of Gouy fringes; in terms of the refractive index difference,  $\Delta n$ , between the bottom and top solutions at the He–Ne laser red light ( $\lambda = 632.8 \text{ nm}$ ),  $J_m = (3.951 \times 10^6) \Delta n$ .  $D_{12}$ , diffusion coefficients.  $B(x_2)$ , thermodynamic factor.  $D_{\Gamma}$ , thermodynamic diffusion coefficient.



**Figure 3.** Comparison of mutual diffusion and intradiffusion coefficients of aqueous L-(+)-tartaric acid solutions at 25 °C:  $\diamond$ ,  $D_1^*$  in light water;  $\blacklozenge$ ,  $D_1^*$  in heavy water;  $\bigcirc$ ,  $D_2^*$  in light water;  $\blacklozenge$ ,  $D_1^*$ ; in heavy water;  $\bigcirc$ ,  $D_2^*$  in heavy water;  $\blacklozenge$ ,  $D_1^*$ ;  $\square$ , computed limiting mutual diffusion coefficients of monodissociated and bidissociated species.

are constant and need not be taken into consideration. The spin-echo peak amplitudes for a given line follow the equation

$$I = I_0 \exp\left[-\frac{2\tau}{T_2} - \gamma^2 g^2 D_i^* \delta^2 \left(\Delta - \frac{\delta}{3}\right)\right]$$
(1)

where  $\gamma$  is the gyromagnetic ratio of the nucleus,  $D_i^*$  is the intradiffusion coefficient of molecules, g is the gradient strength,  $\delta$  and  $\Delta$  are the length and spacing of the gradient pulses,  $\tau$  is the time lag between pulses at 90° and 180°, and  $T_2$  is the spin–spin relaxation time, respectively.

The PGSE-NMR method requires the presence of a deuterated substance as reference, in the measurement region. For aqueous solutions two different methods have been alternatively used. In the first method the measurements were carried out on solutions prepared with undeuterated solvent, using the coaxial tubes (Wilmad model WGS-5BL) with  $C_6D_6$  as external reference and lock. In dilute solutions, the water signal can be so strong it hinders the other ones.

The second method used D<sub>2</sub>O as solvent. This procedure. enhancing the signal intensity of the solute, allows us to analyze very dilute solutions. On the other hand it introduces some errors owing to the isotope effects on the intradiffusion rate. In this case a correction is needed to obtain the corresponding values in light water. Hertz (Goldammer and Hertz, 1970) proposed to multiply the experimental results measured in  $D_2O$  by the factor 1.23, which is the ratio of intradiffusion coefficients of normal and deuterated water, as well as the reciprocal of the ratio of their viscosities. In principle, this correction neglects the possible differences in the interactions of these two solvents with the solute. In fact,  $D_2O$  is thought to be slightly more structured than H<sub>2</sub>O (Nemethy and Scheraga, 1964), so that the hydrophobic effect and the formation of hydrogen bonds could be favored. As a consequence, the ratio of solute intradiffusion coefficients in D<sub>2</sub>O and H<sub>2</sub>O can also depend on the solute nature and concentration.

The two different techniques were applied with the aim of testing the correctness of this correction for the L-(+)-tartaric acid-water system. A good agreement was found between the two sets of measurements. A single polynomial interpolated both sets of experimental data. The polynomial coefficients are reported in Table 2.

In the same measurement sets  $D_{OH}^*$  values were determined. Since the proton exchange between L-(+)-tartaric acid OH's and water is much faster than the single spinecho sequence, the  $D_{OH}^*$  value is a mean value that can be split between L-(+)-tartaric acid and water contributions according to the expression

$$D_{\rm OH}^* = \frac{4m}{4m + (2 \times 55.5)} D_2^* + \frac{2 \times 55.5}{4m + (2 \times 55.5)} D_1^* \quad (2)$$

where  $m/\text{mol kg}^{-1}$  is the L-(+)-tartaric acid molality. This equation allows us to compute  $D_1^*$ . The measured values of  $D_{\text{OH}}^*$ ,  $D_2^*$ , and the computed  $D_1^*$  are collected in Table 4 a and b. The experimental error is within  $(1-2) \times 10^{-2} \text{ cm}^2 \text{ s}^{-1}$ . The  $D_1^*$  data obtained from the two experimental sets fit a polynomial whose coefficients are collected in Table 2. Figure 3 shows the experimental  $D_1^*$  and  $D_2^*$  data and the interpolating equations.

#### **Experimental Results**

In binary systems, the diffusive transport is described by the flux equation

$$J_2 = -M_2 \operatorname{grad} \mu_2 \tag{3}$$

correlating the flow  $J_2$ , expressed in the mass-fixed reference frame, to the chemical potential  $\mu_2$ ;  $M_2$  is the mobility that describes the actual tendency of molecules to diffuse.

Considering the difficulty of treating with the chemical potential gradient, we prefer Fick's phenomenological equation:

$$J_2 = -D_{12} \operatorname{grad} C_2 \tag{4}$$

Table 4. Intradiffusion Data on the System L-(+)-Tartaric Acid–Water at 25  $^\circ C$ 

<i>m</i> /	<i>X</i> <sub>2</sub>	$10^5 D_{\rm OH}^*$	$10^{5}D_{1}^{*}/$	$10^5 D_2^*/$	
mol kg <sup>-1</sup>		$cm^2 s^{-1}$	$cm^2 s^{-1}$	$cm^2 s^{-1}$	
(a) Measured in $D_2O$ and Multiplied by the Factor 1.23					
0.0000	0.0000	2.299	2.299		
0.4568	0.0091			0.69	
0.9716	0.0191	1.71	1.69	0.61	
1.1632	0.0285	1.47	1.44	0.53	
2.2841	0.0438	1.17	1.13	0.41	
3.4195	0.0641	0.90	0.86	0.30	
3.7815	0.0704	0.82	0.78	0.27	
4.7659	0.0872	0.60	0.56	0.21	
6.6794	0.1180	0.44	0.41	0.13	
	(b) M	leasured in H	I <sub>2</sub> O		
0.0000	0.0000	2.299	2.299		
1.5779	0.0303	1.35	1.32	0.53	
2.6237	0.0494	1.03	0.99	0.39	
3.5455	0.0656	0.90	0.86	0.31	
3.8944	0.0716	0.78	0.74	0.25	
4.2577	0.0777	0.74	0.70	0.24	
5.2125	0.0935	0.64	0.60	0.18	
6.4501	0.1132	0.52	0.48	0.14	
7.4924	0.1292	0.45	0.42	0.11	
8.3580	0.1420	0.39	0.41	0.10	

The diffusion coefficient  $D_{12}$  can be obtained multiplying M by the thermodynamic term B, accounting for the fact that the actual diffusion driving force is not the concentration gradient but the chemical potential gradient:

$$D_{12} = M_2 B_2 (5)$$

The thermodynamic factor *B* is correctly expressed in a molar concentration scale:

$$B(C_2) = RT\left(1 + \frac{\mathrm{d}\ln y_2}{\mathrm{d}\ln C_2}\right) \tag{6}$$

 $y_2$  being the activity coefficient expressed as a function of molarity.

In this paper we prefer to express the thermodynamic term B(x) as a function of mole fraction, according to the Laity notation (Laity, 1959):

$$B(x_2) = \left(1 + \frac{d \ln f_2}{d \ln x_2}\right) = \left(1 + \frac{d \ln f_1}{d \ln x_1}\right) = B(x_1) \quad (7)$$

where  $x_i$  and  $f_i$  are the mole fraction and rational activity coefficient of component *i*, respectively. In this way a unique mobility term is defined  $M_2 = M_1 = D_T$ , called the thermodynamic diffusion coefficient.

The thermodynamic factor was straighforwardly computed by differentiating the logarithm of solvent activity, ln  $a_1$ , obtained from literature osmotic data (Robinson et al., 1942), with respect to ln  $x_1$ , where  $x_1$  is the stoichiometric water mole fraction.

The term  $B(x_2)$  was well fitted by the equation

$$B(x_2) = 1 + 7.20x_2 + 53.4x_2^2 - 411x_2^3 \pm 0.01 \ (0.02 < x_2 < 0.09) \ (8)$$

The limiting diffusion coefficients at infinite dilution for the ionic species can be computed with the Nernst–Hartley equation from the literature equivalent conductivity data (Vanysek, 1993),  $\lambda_+^\circ$  = 349.65 and  $\lambda_-^\circ$  = 59.6  $m^2~S~mol^{-1}\times 10^4$ 

for the second dissociation:  $D_{12}^{\infty}(Ac^{2-}) =$ 

$$(2.662 \times 10^{-7}) \frac{3}{2} \cdot \frac{\lambda_+^{\circ} \lambda_-^{\circ}}{\lambda_+^{\circ} \lambda_-^{\circ}} = 2.033 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$$
(9)

for the 1st dissociation:  $D_{12}^{\infty}(AcH^{-}) =$ 

$$(2.662 \times 10^{-7}) \frac{\lambda_{+}^{\circ} \lambda_{-}^{\circ}}{\lambda_{+}^{\circ} + (\lambda_{-}^{\circ}/2)} = 1.462 \times 10^{-5} \,\mathrm{cm}^{2} \,\mathrm{s}^{-1}$$
(10)

and for the intradiffusion:  $D_2^{*\infty} =$ 

$$(2.662 \times 10^{-7}) \frac{\lambda_{-}^{\circ}}{2} = 0.794 \times 10^{-5} \text{ cm}^2 \text{ s}^{-1}$$
 (11)

where the mobilities of  $Ac^{2-}$  and  $AcH^{-}$  were assumed to be equal. The extrapolated  $D_2^{*\infty}$  obtained from our NMR data (Table 2) is in perfect agreement with the value given in eq 11.

An attempt was made to correlate our mutual diffusion data with the values that could be computed in the very dilute solutions.

For the ionized species we assumed the following approximate expression:

$$D_i = D_i^{\infty} (1 + \Delta_1) \left( 1 + \frac{\partial \ln y_{\pm}}{\partial \ln C} \right)$$
(12)

where we used, for the thermodynamic term, the expression

$$\left(1 + \frac{\partial \ln y_{\pm}}{\partial \ln C}\right) = 1 - \frac{1.1722|z_{\pm}z_{-}|\sqrt{I}}{2(1 + 2\sqrt{I})^2}$$
(13)

and, for the Onsager correction term, the expression

$$\Delta_1 = -\frac{0.4351\sqrt{I}}{1+2\sqrt{I}} \tag{14}$$

where *I* is the ionic strength computed on the basis of the actual concentrations of the various ions.

The diffusion coefficient of the solution is given, at each molality, by the expression (Robinson and Stokes, 1955)

$$D_{12} = \alpha_{Ac^2-} D_{12} (Ac^{2-})_{+} \alpha_{AcH-} D_{12} (AcH^{-}) + \alpha_{AcH_2} D_{12} (AcH_2)$$
(15)

where the  $\alpha_i$  values are the fractions of each species present at the given molalities and  $D_{AcH_2}$  was taken from the polynomial whose coefficients are reported in Table 2.

Equation 15 is quite approximate; however, a good agreement was found in the range of overlapping between experimental and computed values. This can be seen in Figure 4.

## Discussion

Mutual diffusion coefficients  $D_{12}$  and thermodynamic diffusion coefficients  $D_{\Gamma}$  are shown in Figure 5 drawn as a function of the solute mole fraction.

Inspection of Figure 5 shows that both mutual and thermodynamic diffusion coefficients decrease with the mole fraction of solute, while the thermodynamic factors increase with  $x_2$  (Table 2). This gives evidence that the mobility contribution prevails on ruling the behavior of diffusion coefficients. An opposite effect can be found in



**Figure 4.** Mutual diffusion coefficients of tartaric acid:  $\bullet$  experimental data;  $\diamondsuit$ , computed data (see text).



**Figure 5.** Diffusion coefficients ( $\bullet$ ) and thermodynamic diffusion coefficients ( $\Box$ ) of L-(+)-tartaric acid aqueous solutions at 25 °C.

some systems, such as surfactant solutions, where both thermodynamic and diffusion coefficients decrease with solute concentration, while mobilities increase (Leaist, 1986; Paduano et al., 1997).

Computing the velocity cross-correlation coefficients, VCC's (McCall and Douglass, 1967; Mills and Hertz, 1980), a deeper insight into the system characteristics can be obtained. VCC's are more sensitive to specific interactions than the experimental diffusion coefficients and allow an analysis from the microscopic point of view (Weingärtner, 1990; Ambrosone et al., 1995).

The phenomenological coefficients can be expressed in terms of time integrals over velocity correlation functions, which give access to a more direct kinetic interpretation of observed physical quantities.

The intradiffusion coefficient  $D_i^*$  of component *i* can be defined as (Steele, 1969)

$$D_i^* = \frac{1}{3} \int_0^\infty \langle v_s^j(0) \cdot v_s^j(t) \rangle \,\mathrm{d}t \tag{16}$$

where the  $v_s^i$  is the velocity of a single particle numbered *s* of component *i* at time 0 and *t*, respectively. The pointed brackets indicate the ensemble average.

The velocity correlation coefficients reflect the correlation in the motion of two different particles *s* and *r*:

$$f_{ij} = \frac{N}{3} x_j \int_0^\infty \langle v_s^i(0) \cdot v_r^j(t) \rangle \,\mathrm{d}t \tag{17}$$

where N is the total number of particles in the system. Equation 17 characterizes the correlation motion between different particles of the same or of different components.



**Figure 6.** Velocity cross-correlation factors for the system L-(+)-tartaric acid–water at 25 °C: 1,  $f_{22}$ ; 2,  $f_{22}$ ; 3,  $f_{11}$ ; 4,  $f_{11}$ ; 5,  $f_{12}$ ; 6,  $f_{12}$ .

The VCC's can be expressed as a function of the experimental quantities  $D_1^*$ ,  $D_2^*$ , and  $D_{12}$  (McCall and Douglass, 1967)

$$f_{12} = -D_{12} \frac{M_1 M_2}{\left(x_1 M_1 + x_2 M_2\right)^2 B(x_1)} x_2 \tag{18}$$

$$f_{ii} = D_{12} \frac{M_j^* x_j}{\left(x_1 M_1 + x_2 M_2\right)^2 B(x_1)} - D_i^*$$
(19)

where  $M_i$  is the molecular weight of component *i* and  $B(x_1)$  is the thermodynamic factor in the mole fraction scale, as defined by eq 7.

According to Hertz's (1982) approach, formulas for standard velocity cross correlation coefficients  $f_{ij}^{o}$  can be derived, by using the law of effective linear momentum conservation and applying the ideal mixing rule:

$$f_{12}^{*} = -\frac{M_{1}x_{2}}{(x_{1}M_{1} + x_{2}M_{2})}D_{1}^{*}(1 + x_{1}P_{12})$$
(20)

$$f_{ii} = -\frac{M_i x_i}{(x_1 M_1 + x_2 M_2)} D_i^* (1 - x_j P_{ij})$$
(21)

where

$$P_{ij} = \frac{M_j D_j^*}{M_i D_i^*} - 1$$
 (22)

The quantities  $f_{ij}$  are representative of a fictitious ideal reference system formed by non-interacting components; in their computation the mutal diffusion coefficient, accounting for the global motion of the species in the system, is not involved. The relationship between the  $f_{ij}$  and  $f_{ij}$ coefficients play the role of indicator for the molecular association effects. As discussed in detail elsewhere (Mills and Hertz, 1980; Weingärtner, 1990), molecular association should lead to more correlated motions then those expected for an "ideal" system. The general association criterion is

$$f_{ij} > f_{ij}^{\circ} \tag{23}$$

The VCC's and the corresponding standard correlation coefficients are reported in Figure 6.

Inspection of Figure 6 shows that  $f_{22}$  coincides almost exactly with  $f_{22}$ , which implies, according to eq 23, that there is no self-association between L-(+)-tartaric acid molecules.

On the contrary the interactions L-(+)-tartaric acid– water and water–water appear to be sensible.

The existence of the interaction water–water is suggested by the observation that  $f_{11}$  is larger than the corresponding  $f_{11}^{*}$ .

Furthermore, according to the results shown in Figure 6, the interaction between L-(+)-tartaric acid and water seems to be the larger effect. This interaction is reflected in the behavior of  $f_{12}$ : in fact  $f_{12} \gg f_{12}^{\circ}$  in the whole range of explored concentration, stressing the presence of strong cross-associations between solute and water.

These evidences can be imputed to the strong hydrophilic behavior of L-(+)-tartaric acid, whose molecule can form hydrogen bonds with water. The solvent molecules participating in the L-(+)-tartaric acid hydration shell interact strongly among themselves, thus explaining the enhanced water—water interactions.

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