

of KEH (20 ml., 0.05 mole) in 1-butanol was added, and the MIBK was evaporated at reduced pressure (water pump) at 40° on the rotary evaporator to give 5.5 g. (38%) of white crystals after drying *in vacuo* (P_2O_5); $[\alpha]^{25}_D +229^\circ$ (c 1.02, water). A direct comparison of the n.m.r. spectrum of this specimen with the n.m.r. spectrum of pure potassium 6-[(+)- α -phenoxypropionamido]penicillanate¹⁶ indicated essential optical purity.

Anal. Calcd. for $C_{18}H_{21}KN_2O_5S \cdot H_2O$: C, 49.75; H, 5.33. Found: C, 49.40; H, 5.18.

Bis(6-phenylmercaptoacetamidopenicillanyl) Disulfide (14).—To 105 mg. (0.25 mmole) of potassium 6-(phenylmercaptoacetamido)thiopenicillanate dissolved in 10 ml. of water was added concentrated HCl to pH 2.5. The solution was layered with 10 ml. of ether and treated with 2 ml. of 0.1 N iodine in ether. The ether was washed with 2% aqueous $NaHCO_3$ solution and finally with water and dried ($MgSO_4$). The ether was evaporated and the residue was dried *in vacuo* (0.1 mm.) for 17 hr. to yield 70 mg. (75%) of amorphous solid. See Table I for analysis and properties. The major infrared absorptions (in cm^{-1}) in KBr were a broad absorption near 3370 (includes the amide NH), 1798 (β -lactam carbonyl), 1731 and 1715 (penicillanic acid disulfide

carbonyl), 1685 (amide carbonyl), and 740 and 690 (monosubstituted phenyl). The n.m.r. spectrum of a solution of the disulfide in $CDCl_3$ had absorption peaks which were assigned as follows: a doublet of spacing 9 c.p.s. at δ 7.59 due to the amide proton which is coupled to the C-6 proton, a singlet at 7.36 due to the 5 aromatic protons, a quartet centered at 5.84 ascribed to the C-6 proton which is coupled to the amide proton ($J = 9$ c.p.s.) and to the C-5 proton ($J = 4.5$ c.p.s.), the C-5 proton gave rise to a doublet of spacing 4.5 c.p.s. at 5.55, a singlet at 4.37 due to the C-3 proton, a singlet at 3.68 from the protons of the methylene group, and singlets at 1.60 and 1.51 due to the *gem*-dimethyl protons.

Acknowledgments.—We wish to express our thanks to David F. Whitehead and Albert Vulcano for the interpretation of the infrared and n.m.r. spectra and to R. M. Downing and C. Kalinowski for the elemental analyses. For the microbiological data we are indebted to Dr. Joseph Lein, Dr. Alexander Gourevitch, Dr. John A. Bach, and their associates.

Studies on 2-(α -Hydroxybenzyl)benzimidazole (HBB) Analogs. I. Synthesis of 8-(α -Hydroxybenzyl)purines, the Diaza Analogs of HBB^{1a,b}

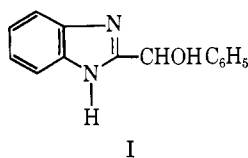
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The antiviral activity displayed by a number of 2-(α -hydroxybenzyl)benzimidazoles (HBB) and several purines has initiated the synthesis of some 8-(α -hydroxybenzyl)purines. These compounds were prepared by the cyclization of 4-amino-5-(acetylmandelamino)pyrimidines which were in turn prepared by the reaction of 4,5-diaminopyrimidines with acetylmandelyl chloride. Preliminary testing results of these diaza analogs of HBB against Sarcoma 180 and KB cell culture are reported. 8-(α -Hydroxybenzyl)purine was found to be inactive against type 1 and 2 polio virus *in vitro*.

The interesting biological activity of 2-(α -hydroxybenzyl)benzimidazole (HBB, I), which suppresses poliomyelitis virus infection in mice, was first described in 1958.² Later, Tamm, *et al.*,³ reported that HBB and its 6-chloro derivative showed selective in-



hibition against type 2 polio virus. Recently, O'Sullivan and Wallis⁴ showed that 1-alkyl-substituted HBB compounds had powerful activity in tissue culture against types 1, 2, and 3 polio virus and possessed protection to ERK cells against the cytopathogenicity of enteroviruses. Compounds of this type have been

shown to inhibit the synthesis of virus-directed RNA polymerase,⁵ of viral RNA,⁶ and viral coat protein.^{6a}

Much light has been cast on structure-activity relationships in this series of compounds.^{3,4,7,8} Hydrogen bonding and metal chelation have also been investigated. From the information available at present, it appears that the α -hydroxybenzyl moiety in I is not only of fundamental importance but rather specific for the virus inhibitory action of HBB. 2-Hydroxymethyl and 2-(α -hydroxyethyl) derivatives of benzimidazole, for instance, were inactive.³ The corresponding 2-benzoyl and 2-benzyl derivatives were less active with little selectivity of action.³ The fact that 2-(*o*-hydroxybenzyl)benzimidazole (II) possesses similar antiviral activities but the isomeric *p*-hydroxy derivative failed to do so⁹ indicated that the existence of intramolecular hydrogen bonding and/or steric requirements¹⁰ plays an important role in this type of

(1) (a) This investigation was supported by the Cancer Chemotherapy National Service Center, National Cancer Institute of the National Institutes of Health, Public Health Service, Contract SA-43-ph-3025. (b) Presented in part before the Division of Medicinal Chemistry, 147th National Meeting of the American Chemical Society, Philadelphia, Pa., April 1964. (c) To whom all inquiries should be directed.

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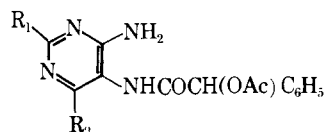
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(10) Examination of LaPine atomic models of I and II confirmed that the hydroxy group in II can take relatively the same position that is occupied by the hydroxyl group in I.

TABLE II

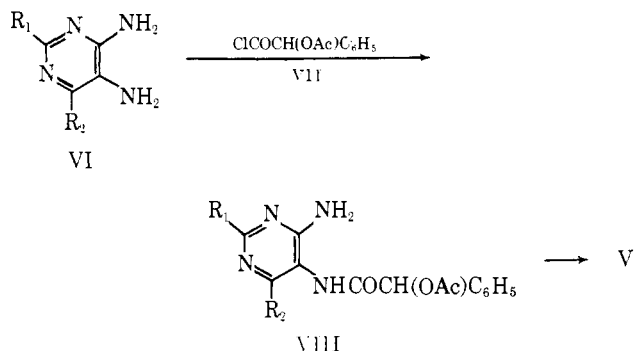
4-AMINO-5-(ACETYLMANDELAMIDO)PYRIMIDINES*



R ₁	R ₂	Recrystn. solvents	Yield, %	M.p., °C. dec.	Formula	Caled., %			Found, %			pH 1		pH 11	
						C	H	N	C	H	N	λ_{\max} , m μ	$\epsilon \times 10^{-3}$	λ_{\max} , m μ	$\epsilon \times 10^{-3}$
H	H	Ethyl acetate	53	198-200	C ₁₄ H ₁₄ N ₄ O ₂	58.7	4.93	19.6	58.6	5.08	19.1	251	12.0	236	8.5
H	CH ₃	Ethanol-ethyl acetate	83	240-242	C ₁₅ H ₁₆ N ₄ O ₂ ·HCl	53.5	5.06	16.6	53.6	5.21	16.5	249	11.1	234	7.7
H	NH ₂	Water-methanol	56	268	C ₁₄ H ₁₅ N ₅ O ₃	55.8	5.01	23.2	55.3	5.33	23.2	263	12.0	257	6.0
H	CH ₃ S	Acetone	71	230-232	C ₁₅ H ₁₆ N ₄ O ₃ S	54.2	4.85	16.9	54.3	5.14	16.6	242	17.3	230	18.6
NH ₂	H	Water-ethanol	81	252	C ₁₄ H ₁₅ N ₅ O ₃ ·HCl	49.9	4.77	20.8	49.8	4.97	20.9	294	12.6	286	8.0
CH ₃ S	H	Water-acetone	80	180-181	C ₁₅ H ₁₆ N ₄ O ₃ S	54.2	4.85	16.9	54.5	5.23	16.8	265 (s)	5.7	227	13.4
														287	7.4
														226	15.3
														253	11.6
														291	8.0
NH ₂	CH ₃ S				See Ref. 27										
CH ₃	NH ₂	Ethanol-ether	64	250-252	C ₅ H ₁₇ N ₅ O ₃ ·HCl	51.3	5.15	19.9	51.6	5.48	20.2	264	10.6	258	9.9
H	C ₆ H ₅ CH ₂ S	Hexane-acetone	66	176-178	C ₂₁ H ₂₀ N ₄ O ₃ S	61.8	4.95	13.7	61.8	5.21	13.7	240	16.3	233	20.4
H	OH	Water-dimethyl formamide	59	295-300	C ₁₄ H ₁₄ N ₄ O ₄ ·0.5H ₂ O	54.0	4.83	18.0	54.3	4.86	18.2	297	11.4	280	11.0
														258	6.3

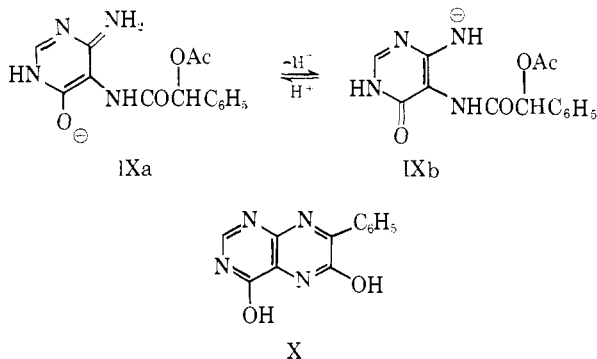
* Pyrimidine precursors (see text): VIa, D. J. Brown, *J. Appl. Chem.*, **2**, 239 (1952); VIb and VIc, A. Albert, D. J. Brown, and H. C. S. Wood, *J. Chem. Soc.*, 3832 (1954); VIe, prepared by the reduction of 4,6-diamino-5-nitropyrimidine, R. K. Robins, K. J. Dille, C. H. Willits, and B. E. Christensen, *J. Am. Chem. Soc.*, **75**, 263 (1953); VIe and VIj, A. Albert, D. J. Brown, and G. Cheeseman, *J. Chem. Soc.*, 474 (1951); VIi, A. Albert and D. J. Brown, *ibid.*, 2060 (1954); VIg, G. D. Daves, Jr., C. W. Noell, R. K. Robins, H. C. Koppel, and A. G. Beaman, *J. Am. Chem. Soc.*, **82**, 2633 (1960); VIh, prepared by the reduction of 2-methyl-4,6-diamino-5-nitrosopyrimidine, E. C. Taylor, O. Vogl, and C. C. Cheng, *ibid.*, **81**, 2442 (1959); VIi, G. B. Elion, W. H. Lange, and G. H. Hitchings, *ibid.*, **78**, 2858 (1956).

agent since the former caused some hydrolysis of the amide. In a similar manner, several substituted purine analogs (Vb-i) were also prepared. These purine analogs were either patterned after the known HBB derivatives, naturally occurring purines, pseudo-vitamin B₁₂ components,²¹ or purines possessing antitumor activity. The ultraviolet absorption maxima were in accord with those of the corresponding 8-unsubstituted purines.



- | | |
|---|---|
| a, R ₁ , R ₂ = H | g, R ₁ = NH ₂ ; R ₂ = CH ₃ S |
| b, R ₁ = H; R ₂ = CH ₃ | h, R ₁ = CH ₃ ; R ₂ = NH ₂ |
| c, R ₁ = H; R ₂ = NH ₂ | i, R ₁ = H; R ₂ = C ₆ H ₅ CH ₂ S |
| d, R ₁ = H; R ₂ = CH ₃ S | j, R ₁ = H; R ₂ = OH |
| e, R ₁ = NH ₂ ; R ₂ = H | k, R ₁ = H; R ₂ = SH |
| f, R ₁ = CH ₃ S; R ₂ = H | |

The pure hypoxanthine analog (Vj) of HBB could not be synthesized by this general procedure. Cyclization of the intermediate amide VIIIj led to the isolation of a bright yellow product which showed an ultraviolet absorption maximum in the 380–390-m μ region in addition to the characteristic ultraviolet absorption spectrum for hypoxanthine. Paper chromatographic and other studies revealed the presence of a small amount of a side product, which is believed to be 4,6-dihydroxy-7-phenylpteridine (X). Compound X is presumably formed through a competitive cyclization *via* the intermediate IXb (formed in the presence of -OC₂H₅). Studies dealing with the formation of the by-product pteridine were not carried further in the present work.



Since much difficulty has been encountered in the separation of the purine (Vj) from the contaminated pteridine (X), pure Vj was obtained by the oxidation of 4-(methylthio)-8-(α -hydroxybenzyl)purine (Vd) with hydrogen peroxide followed by acid hydrolysis.²²

(21) 2-Methyladenine was found to be a component in pseudo-vitamin B₁₂; cf., E. L. Smith in "The Chemistry and Biology of Purines," Little, Brown and Co., Boston, Mass., 1957, pp. 160–168.

(22) Similar conversion of a methylthio group to a hydroxyl group by oxidation has been successfully applied in the pyrazolo[3,4-d]pyrimidine system. See R. K. Robins, *J. Am. Chem. Soc.*, **79**, 6407 (1957).

The corresponding 8-(α -hydroxybenzyl)-4-thiopyrimidine (Vk) was prepared by the debenzoylation of Vi by means of sodium and liquid ammonia.²³

Biological Results and Discussion

Preliminary anticancer testing results of some of the 8-(α -hydroxybenzyl)purines are given in Table III. 8-(α -Hydroxybenzyl)purine (Va), one of the diazo analogs of HBB, has been studied for possible antiviral properties by techniques similar to those used by Tamm and associates³ for HBB.²⁴ These tests were made in roller-tube tissue cultures of the LLC-MK₂ continuous cell line derived from rhesus monkey kidney.²⁵ The cultures had been grown in medium 199 supplemented with 1% normal horse serum. When confluent cell sheets had developed, the culture medium was removed, the cultures were rinsed with serum-free medium, fresh medium 199 (without serum) containing the compound in the desired concentration was added, and the tubes were inoculated with polio virus. Incubation was at 36–37° and cytopathogenic changes were observed daily. Control cultures were prepared and observed in the same manner except that the compound was not incorporated into the medium. Likewise, cultures were prepared and observed utilizing medium 199 containing HBB. Thus compound Va was directly compared to a known antiviral compound. (Compound Va and HBB were dissolved in absolute ethyl alcohol as concentrates and then diluted in medium 199 for addition to the cultures.)

When compound Va was tested against type 1 polio virus (Brunnhilde strain), no antiviral activity was observed with concentrations of 400 μ M of the compound in the medium. This was true for the two different levels of virus used: 10⁴ plaque-forming units (PFU)/tube and 10³ PFU/tube. However, HBB demonstrated significant antiviral activity in the same test. Both the 10⁴ and 10³ PFU concentrations of virus were suppressed for 7 days by 400 μ M HBB. With 100 μ M HBB in the medium the type 1 polio virus cytopathogenic effects were delayed approximately 2 days longer than observed with the untreated controls.

Data obtained with compound Va against polio virus type 2 (Brooks strain) were similar to those obtained for type 1 polio virus. Compound Va at 200 and 50 μ M levels failed to protect cultures against 1000 and 10 PFU of polio virus type 2. In the same test 200 μ M HBB delayed the cytopathologic changes due to the virus for approximately 3 days. No antiviral activity was observed against polio virus 2 when 50 μ M HBB was tested.

HBB and compound Va demonstrated very similar toxicity in the LLC-MK₂ cells. Both compounds were toxic to about 50% of the cells when used in 400 μ M concentrations. At a 100- μ M concentration HBB exhibited no toxicity and Va only caused a very few cells to become granular, somewhat elongated, and slough from the glass. These toxic effects were more pronounced from the third through the seventh days of

(23) The detailed debenzoylation conditions were described by G. B. Eison, W. H. Lange, and G. H. Hitchings, *ibid.*, **78**, 2858 (1956).

(24) J. O. MacFarlane and R. D. Lamb, to be published.

(25) R. N. Holl, W. R. Cherry, and O. J. Tritsch, *J. Exptl. Med.*, **115**, 963 (1962).

amide (VIIb and h) was formed, solid product could be obtained by triturating the resulting syrup with acetone rather than with water.²⁷

B. VIIIc and e.—An equimolar mixture of finely ground triaminopyrimidine (free base, freshly prepared from the corresponding 5-nitropyrimidine by Raney nickel reduction) and acetylmandelyl chloride was cautiously warmed on the steam bath. At ca. 90° a vigorous reaction occurred. After the reaction subsided the heterogeneous mixture was warmed for another 15 min. with constant stirring. The reaction mixture was then diluted with a large amount of acetone. The crude solid product was then filtered and purified.

8-(α -Hydroxybenzyl)purines (Va-i, Table I).—To a potassium ethoxide solution (prepared by dissolving 2.1 g. of K in 20 ml. of absolute ethanol) was added 5 g. of the amide. The mixture was warmed on a steam bath for 4 hr., poured into 250 ml. of water, and acidified with glacial acetic acid. The resulting precipitate was filtered and recrystallized.

6-Hydroxy-8-(α -hydroxybenzyl)purine (Vj).—To a solution of 200 ml. of water, 10 ml. of concentrated HCl and 10 ml. of 30% of H₂O₂ was added 4.5 g. of 6-methylthio-8-(α -hydroxybenzyl)purine (Vd). The mixture was refluxed for 20 min. and, without cooling, excess acid was carefully neutralized with dilute NaOH. After refrigeration overnight, 2.0 g. of white,

(27) In the case of the preparation of VIIIg, a mixture of mono- and bis(acetylmandelamino)pyrimidine²⁸ was formed. The monoacetyl derivative was difficult to purify. The bisacetyl derivative, after recrystallization from ethanol, gave the following information: m.p. 208–209°; λ_{max}^{220} 231 m μ (ϵ 23,000), 303 (14,700); χ_{max}^{220} 290 m μ (ϵ 10,400). *Anal.* Calcd. for C₂₁H₂₃N₃O₅·0.511H₂O: C, 56.4; H, 4.88; N, 12.9. Found: C, 56.5; H, 5.11; N, 12.8. The crude intermediate, however, was readily cyclized to the desired compound Vj by normal procedure in a 60% over-all yield.

(28) Formation of bis(acetylamido)pyrimidines has also been reported by other investigators: cf. A. Albert and D. J. Brown, *J. Chem. Soc.*, 2060 (1954).

analytically pure product was isolated from the reaction mixture (see Table I for additional data).

Preparation of Vj by the cyclization of 4-amino-5-acetylmandelamino-6-hydroxypyrimidine (VIIj) in potassium ethoxide yielded a yellow solid which gave an additional ultraviolet absorption maximum at 380–390 m μ . Proper analysis for Vj was obtained (*Anal.* Calcd. for C₂₁H₂₃N₃O₅: C, 59.5; H, 4.17; N, 23.1. Found: C, 59.3; H, 3.78; N, 23.0.) after the crude product was repeatedly recrystallized from water and ethanol. However, the 380–390-m μ absorption was still present in the final product and paper chromatographic measurements of the recrystallized product still indicated the presence of a trace amount of a fluorescent substance. Work on the formation of this by-product, which is believed to be a pteridine, is not included in the present work.

6-Thio-8-(α -hydroxybenzyl)purine (Vk).—To a three-necked 500-ml. flask equipped with stirrer, Dry Ice cold finger, and drying tube was introduced 200–250 ml. of liquid ammonia followed by the addition of 5.0 g. of 6-benzylthio-8-(α -hydroxybenzyl)purine (Vi). To the light yellow solution was added portionwise, with stirring, 0.9 g. of sodium. After the addition was complete, the mixture was stirred for 30 min. Excess NH₃ was allowed to evaporate and the residue was dissolved in 60 ml. of water. This was then acidified with glacial acetic acid and the resulting yellow gum was recrystallized from a mixture of water and ethanol to yield 2.3 g. of a bright yellow solid (see Table I for additional data).

Acknowledgment.—The authors wish to express their appreciation to Mr. Hal P. Van Fossen, Mrs. Margaret L. Rounds, and Mr. John R. Gravatt for the analytical and instrumental measurements. They are also indebted to Dr. John O. MacFarlane and Mr. R. D. Lamb for the antiviral evaluation.

Substituted 5,6-Dihydro-2-(2-, 3-, and 4-pyridyl)-4H-1,3,4-oxadiazines¹

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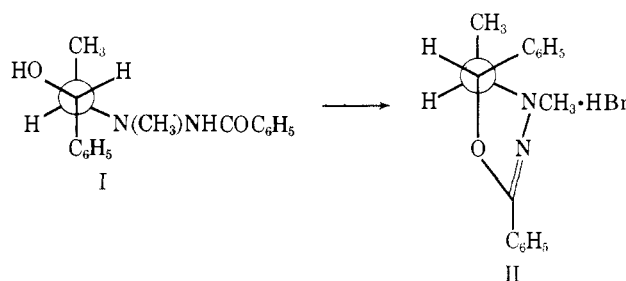
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Received May 21, 1965

Various 2-(β -hydroxyalkyl)nicotinic, -isonicotinic, and -picolinic acid hydrazides were prepared by treating appropriate hydrazino alcohols with either the chloride, mixed anhydride, or anhydride of nicotinic, isonicotinic, and picolinic acids. Cyclodehydration of the 2-(β -hydroxyalkyl)nicotinic, isonicotinic, and picolinic acid hydrazides, containing either a primary, secondary, or tertiary hydroxyl group, to substituted 5,6-dihydro-2-(2-, 3-, and 4-pyridyl)-4H-1,3,4-oxadiazines was accomplished utilizing the four following methods: (1) concentrated H₂SO₄, (2) hydrogen bromide in acetic acid, (3) O-tosylation followed by solvolysis, and (4) replacement of OH by Cl followed by NaOH dehydrochlorination. All 2-(4-pyridyl)oxadiazines in this series antagonized the effects of tremorine in mice. The 2-(2- and 3-pyridyl) isomers were inactive in this test. It is suggested that the 2-(4-pyridyl)oxadiazines interfere with the metabolic conversion of tremorine since they did not antagonize its active metabolite oxotremorine.

As part of a continuing exploratory research program in heterocyclic syntheses, we turned our attention to the substituted 5,6-dihydro-2-(2-, 3-, and 4-pyridyl)-4H-1,3,4-oxadiazines. Previously,² we had discovered that the acid-catalyzed dehydration of certain 2-(β -hydroxyalkyl)carboxylic acid hydrazides proceeds *via* neighboring-group participation with concomitant formation of a substituted 5,6-dihydro-4H-1,3,4-oxadiazine. For example, treatment of *erythro*-(–)-2-methyl-2-(α -methyl- β -hydroxy- β -phenethyl)benzoic

acid hydrazide (I) with gaseous hydrogen bromide in glacial acetic acid (HBr-AcOH) at ambient temperature gave *cis*-(–)-4,5-dimethyl-2,6-diphenyl-5,6-dihydro-4H-1,3,4-oxadiazine hydrobromide (II) in 87% yield.^{2d}



(1) Presented in part before the Division of Medicinal Chemistry at the 150th National Meeting of the American Chemical Society, Atlantic City, N. J., Sept. 1965.

(2) (a) D. L. Trepanier, V. Sprancmanis, and K. G. Wiggs, *J. Org. Chem.*, **29**, 668 (1964); (b) D. L. Trepanier and V. Sprancmanis, *ibid.*, **29**, 673 (1964); (c) *ibid.*, **29**, 2151 (1964); (d) D. L. Trepanier, V. Sprancmanis, D. S. Tharpe, and P. E. Krieger, *J. Heterocyclic Chem.*, in press.