

was dried to constant weight *in vacuo*. The resulting powder was very hygroscopic, sintered at 50°, and decomposed at 62°. Fehling's solution was reduced and one major spot was noted on thin layer chromatograms on silica gel developed with BuOH-EtOH-H₂O (visualized by Tollens reagent). *Anal.* (C₁₁H₈N₂O₄·HCl) C, H; N: calcd, 15.18; found, 15.74.

Hydrogenolysis of 4.—A solution of 184 mg of N-hydroxy compound (4) as its hydrochloride in 25 ml of EtOH was hydrogenated at atmospheric pressure with PtO₂ (50 mg). After 6 hr the theoretical uptake was complete, the catalyst was removed, and the filtrate was taken to dryness. The product, dissolved in a few milliliters of H₂O, gave a negative Fehling's test. Electrophoresis performed with authentic samples of isoasparagine, asparagine, and aspartic acid at pH 3.5 (acetate buffer, 500 v, 4 hr) with ninhydrin as an indicator, or paper chromatography with 88% phenol, showed the product to contain but traces of isoasparagine and aspartic acid. The major component, DL-asparagine, precipitated on adjusting the solution to pH 3, had essentially zero rotation, and contained no isoasparagine.

Benzenitrone of Succinamide.—A solution of 2 g of 2 in 30 ml of 28% aqueous NH₃ was allowed to stand at room temperature for 24 hr. The diamide precipitated (600 mg, 28%) and was collected and recrystallized from 75% EtOH. It melted at 204–205° dec. *Anal.* (C₁₁H₁₃N₃O₃) C, H, N.

2-Hydroxylaminosuccinamide Hydrochloride.—A solution of 350 mg of the above nitron in 10 ml of 1 N HCl was concentrated *in vacuo* keeping the bath temperature below 30°. The resulting precipitate was taken up twice in EtOH and precipitated by the addition of ether. The white powder, dried *in vacuo* to constant weight, decomposed at about 80°. Chromatography with BuOH-EtOH-H₂O indicated one component with Tollens reagent at R_f 0.3. *Anal.* (C₄H₉N₃O₃·HCl) C, H, N.

Some 3-Methyl-2-butenylaminopurines and -pyrimidines¹

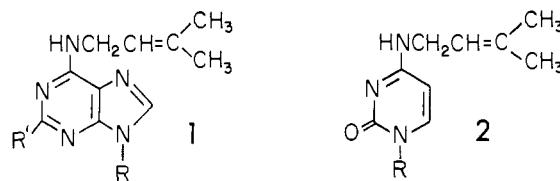
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6-(3-Methyl-2-butenylamino)-9-β-D-ribofuranosyl-purine has been recently reported as a minor component of yeast and liver soluble ribonucleic acid² and of yeast serine transfer ribonucleic acid.^{3,4} 6-(3-Methyl-2-butenylamino)purine has been found in extracts of *Corynebacterium fascians*.⁵ Both compounds have been synthesized and exhibit a high level of cytokinin activity.^{2,6} It was therefore of interest to synthesize other 3-methyl-2-butenylamino pyrimidines and purines and to test them for biological activity.

In all cases the syntheses were carried out by refluxing an excess of γ,γ-dimethylallylamine with the appropriate mercapto or alkylmercaptapurine or -pyrimidine according to the method of Elion, *et al.*⁷ The reactions were followed spectrophotometrically in ethanol and were terminated when the absorption maximum of the



1		2	
R	R'	R	
a. H	NH ₂	a. H	
b. ribofuranosyl	NH ₂	b. ribofuranosyl	
c. H	OH		

starting material (297–315 mμ) was no longer present. Proof of structure of the products **1a–c** and **2a, b** was shown by the similarity of their ultraviolet absorption spectra to the purine N⁶- or pyrimidine N⁴-mono-alkylated parent compounds (Table I). That the double bond in the side chain is in the position shown is demonstrated by the nmr methyl signals at δ 1.75 (DMSO-*d*₆, TMS).^{2,8}

In the leaf senescence test⁹ all of the compounds (**1** and **2**) showed about one-tenth the activity reported for 6-furfurylaminopurine (kinetin).¹⁰ None of these compounds exhibited any activity against mouse leukemia L1210 *in vivo*.¹¹ Compounds **2a** and **2b** were also inactive against Burkitt's cell cultures.^{11a} The cytidine analog **2b** was not deaminated by human liver or mouse kidney enzyme systems.^{11a}

Experimental Section

Melting points were taken on a Mel-Temp apparatus and are corrected. Spectra were determined using a Cary Model 15 spectrophotometer. Analyses were done by Spang Microanalytical Laboratory, Ann Arbor, Mich., and Galbraith Laboratories, Inc., Knoxville, Tenn. Where analyses are indicated only by the symbols of the elements, analytical results obtained for those elements were within ±0.4% of the theoretical values.

2-Amino-6-(3-methyl-2-butenylamino)purine (1a).—2-Amino-6-methylthiopurine¹² (1.25 g, 6.9 mmoles) and γ,γ-dimethylallylamine¹³ (5.0 g, 59 mmoles) were refluxed for 30 hr. A precipitate which formed on cooling was washed with petroleum ether (30–60°) to remove the excess amine and was dissolved in 95% EtOH. After the addition of concentrated HCl, 1.18 g (67%) of **1a** hydrochloride was obtained. A sample for analysis was recrystallized from EtOH after treating with charcoal; mp 250–251° dec. *Anal.* (C₁₀H₁₄N₆·HCl) C, H, N, Cl.

2-Amino-6-(3-methyl-2-butenylamino)-9-β-D-ribofuranosyl-purine (1b).—6-Ethylthioguanosine¹⁴ (2.0 g, 6.1 mmoles) and γ,γ-dimethylallylamine (6.0 g, 71 mmoles) were refluxed for 60 hr. After the addition of petroleum ether (30–60°), a dark oil separated. The oil was triturated several times with petroleum ether to remove any excess amine and was then dissolved in 95% EtOH, treated with charcoal, and filtered. The crystals which formed were washed with cold EtOH and dried (yield 1.31 g, 48%). A sample for analysis was recrystallized from EtOH, mp 93–97°. *Anal.* (C₁₅H₂₂N₆O₄·EtOH) C, H, N.

2-Hydroxy-6-(3-methyl-2-butenylamino)purine (1c).—2-Hydroxy-6-methylthiopurine¹⁵ (0.42 g, 2.3 mmoles), γ,γ-dimethylallylamine (4.0 g, 47 mmoles), and EtOH (5.0 ml) were refluxed for 90 min. The crystals which formed on cooling were washed with EtOH, dried, dissolved in 1 N NaOH, and reprecipitated

(1) (a) This investigation was supported in part by National Cancer Institute Grant CA 08748. (b) The side chains of these compounds have also been referred to as γ,γ-dimethylallylamine or Δ²-isopentenyl groups.

(2) R. H. Hall, M. J. Robins, L. Stasiuk, and R. Thedford, *J. Am. Chem. Soc.*, **88**, 2614 (1966).

(3) H. G. Zachau, D. Dütting, and H. Feldmann, *Angew. Chem. Intern. Ed. Engl.*, **5**, 422 (1966).

(4) K. Bienmann, S. Tsunakawa, J. Sonnenbichler, H. Feldmann, D. Dütting, and H. G. Zachau, *ibid.*, **5**, 590 (1966).

(5) D. Klämbt, G. Thies, and F. Skoog, *Proc. Natl. Acad. Sci. U. S. A.*, **56**, 52 (1966).

(6) H. Q. Hamzi and F. Skoog, *ibid.*, **51**, 76 (1964).

(7) G. B. Elion, E. Burgi, and G. H. Hitchings, *J. Am. Chem. Soc.*, **74**, 411 (1952).

(8) Spectra were obtained with a Varian A-60 spectrometer. We would like to thank Mr. Marvin Olsen for obtaining these spectra.

(9) B. I. S. Srivastava and G. Ware, *Plant Physiol.*, **40**, 62 (1965).

(10) We would like to thank Drs. B. I. S. Srivastava and R. H. Hall for these preliminary results.

(11) (a) These preliminary data were obtained from M. R. Dollinger, *et al.*; manuscript in preparation; (b) Dr. Dorris Hutchison of this institute.

(12) E. O. Leonard, C. G. Skinner, E. M. Lansford, and W. Shive, *J. Am. Chem. Soc.*, **81**, 907 (1959).

(13) D. Semenov, C. H. Shih, and W. G. Young, *ibid.*, **80**, 5472 (1958).

(14) C. W. Noell and R. K. Robins, *J. Med. Pharm. Chem.*, **5**, 1074 (1962).

(15) E. O. Leonard, W. H. Orme-Johnson, T. R. McMurtry, C. G. Skinner, and W. Shive, *Arch. Biochem. Biophys.*, **99**, 16 (1962).

TABLE I
 ULTRAVIOLET ABSORPTION

Compound	0.1 N HCl		λ , $m\mu$ ($\epsilon \times 10^{-3}$) pH 7		0.1 N NaOH	
	λ_{max}	ϵ_{max}	λ_{max}	ϵ_{max}	λ_{max}	ϵ_{max}
1a	280 (14.3)	261 (9.0)	281 (13.2)	261 (7.3)	286 (13.1)	273 (4.8)
	247.5 (12.6)	237.5 (11.3)	248 (7.9)	242 (7.6)		
1b	291 (12.9)	271 (8.8)	281 (15.5)	243 (6.8)	281 (15.5)	243 (6.8)
	254 (12.1)	238 (8.0)	263 sh (10.9)		263 sh (10.9)	
1c	287 (17.3)	250 (1.9)			285 (15.9)	256 (3.7)
2a	278 (12.4)	241 (2.5)	267.5 (9.2)	248 (6.9)	282 (8.1)	253 (3.7)
			232 (8.5)		230 sh (8.3)	
2b	282.5 (15.8)	243 (2.9)	271 (13.0)	247.5 (9.2)	271 (13.0)	247.5 (9.2)
			239 (9.6)	228 (9.1)	239 (9.6)	228 (9.1)

by neutralization with glacial AcOH. The precipitate was washed with H₂O and dried to give 300 mg (55%) of **1c** monohydrate. An analytical sample was prepared by recrystallization from 1 N NaOH by acidification with AcOH; mp >300° dec. *Anal.* (C₁₀H₁₃N₃O·H₂O) C, N; H: calcd, 6.37; found, 5.92.

2-Hydroxy-4-(3-methyl-2-butenylamino)pyrimidine (2a).—4-Methylthiouracil¹⁶ (1.5 g, 10.5 mmoles), γ,γ -dimethylallylamine (5.0 g, 59 mmoles), and EtOH (5.0 ml) were refluxed for 2.5 hr. The precipitate which formed on cooling was washed with petroleum ether (30–60°) and recrystallized from 95% EtOH and dilute H₂SO₄ to give 1.82 g (95%) of **2a**·0.5H₂SO₄. A sample for analysis was recrystallized twice from EtOH–H₂O and dried; mp 229–230°. *Anal.* (C₉H₁₃N₃O·0.5H₂SO₄) C, H, N.

2-Hydroxy-4-(3-methyl-2-butenylamino)-1- β -D-ribofuranosylpyrimidine (2b).—1-(2,3,5-Tri-O-benzoyl- β -D-ribofuranosyl)-4-thiouracil¹⁷ (2.0 g, 4.3 mmoles) was refluxed with γ,γ -dimethylallylamine (10 g, 118 mmoles) for 90 min. The excess amine was removed under vacuum to give a brown oil. The oil was dissolved in 10 ml of EtOH and 300 ml of Et₂O was added. The pale tan oil which separated was shown by thin layer chromatography (silica gel; GF₂₅₄, CHCl₃–MeOH 3:1) to be essentially free of N-(3-methyl-2-butenyl)benzamide. Final separation was carried out on a silica gel column (2.5 × 18 cm, Fisher silica gel 100–200 mesh, grade 923) by elution with CHCl₃–MeOH 3:1. The solvent was removed under vacuum to give an amorphous and somewhat unstable product. *Anal.* (C₁₇H₂₁N₃O₅) H, N; C: calcd, 54.01; found, 53.02.

A tetrabenzoyl derivative of **2b** was prepared by heating 100 mg of **2b** with an excess of benzoyl chloride in pyridine at 60° until the (silica gel GF₂₅₄, C₆H₆–Et₂O 2:1) showed a single spot at R_f ~0.50. The product was poured over ice and gave a light tan oil. The oil was triturated with H₂O and allowed to stand for a few days. The needles which formed were recrystallized twice from EtOH; mp 193–194°. *Anal.* (C₂₂H₂₇N₃O₅) C, H, N.

(16) Prepared according to the method of T. Ueda and J. J. Fox, *J. Med. Chem.*, **6**, 697 (1963).

(17) J. J. Fox, D. Van Praag, I. Wempen, I. L. Doerr, L. Cheong, J. E. Knoll, M. L. Eidinoff, A. Bendich, and G. B. Brown, *J. Am. Chem. Soc.*, **81**, 178 (1959).

Nucleosides. XLVII. Syntheses of Some N⁴-Substituted Derivatives of 1- β -D-Arabinofuranosylcytosine and -5-fluorocytosine¹

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The clinical usefulness of 1- β -D-arabinofuranosylcytosine (**4**) in the treatment of acute leukemias and

lymphomas has been demonstrated,^{2a} but various pharmacological studies^{2b,c} have shown that this drug is rapidly deaminated to the inactive nucleoside, 1- β -D-arabinofuranosyluracil (**1**). It has been shown previously that alkylation of the exocyclic amino function of cytosine nucleosides, such as 2'-deoxy-5-fluorocytidine produced a marked decrease of the susceptibility of such compounds to deamination by bacterial deaminases.³ It was further shown⁴ that **4** also serves as a substrate for bacterial deaminases. The 5-fluoro analog (**4b**, R'' = H)⁵ was also effective against several mouse leukemias, but this analog, too, is degraded by human liver or mouse kidney deoxyctidine deaminases to **1b**, R = H.⁶ These results^{6,7} suggested the synthesis of N⁴-substituted derivatives of **4a** and **4b** as potential chemotherapeutic agents and/or as deoxyctidine deaminase inhibitors. The present report describes the synthesis of several of these N⁴-substituted derivatives selected for biological evaluation.

The syntheses of N⁴-substituted arabinosylexycytosine nucleosides (**4**) were accomplished by the thiation process^{5,8} which involved thiation of a suitably protected nucleoside (**1**) with phosphorus pentasulfide in pyridine to give the 4-thiones (**2**) followed by alkylation to the 4-methylthio analogs (**3**). Treatment of **3** with various nucleophiles (*e.g.*, NH₂OH, NH₂NH₂, CH₃NH₂) gave the desired products (**4**) (Scheme I).

Though the synthesis of **2b** (R = COCH₃) had been reported,⁹ its preparation by thiation of **1b** was rather difficult and several retreatments with P₂S₅ were necessary to drive the reaction to completion. Moreover, during the extended thiation-reaction time, some of the acetyl-protecting groups were apparently lost and **2b** could not be obtained in pure form. Although crude **2b**, thus obtained, was satisfactory for subsequent conversions, it was hoped that benzoyl blocking groups

(2) (a) R. Talley and V. K. Viatkevichs, *Blood*, **21**, 352 (1963); E. S. Henderson and P. J. Burke, *Proc. Am. Assoc. Cancer Res.*, **6**, 26 (1965); R. W. Carey and R. R. Ellison, *Clin. Res.*, **13**, 337 (1965); K. P. Yu, J. P. Howari, and B. D. Clarkson, *Proc. Am. Assoc. Cancer Res.*, **7**, 78 (1966); R. R. Ellison, J. F. Holland, T. Silver, J. Bernard, and M. Boiron, *Proc. 9th Intern. Cancer Cong., Tokyo, 1967*, in press; (b) R. Papac, W. A. Creasey, P. Calabresi, and A. D. Welch, *Proc. Am. Assoc. Cancer Res.*, **6**, 50 (1965); (c) G. W. Camiener and C. G. Smith, *Biochem. Pharmacol.*, **14**, 1405 (1965).

(3) I. Wempen, R. Duschinsky, L. Kaplan, and J. J. Fox, *J. Am. Chem. Soc.*, **83**, 4755 (1961).

(4) L. I. Pizer and S. S. Cohen, *J. Biol. Chem.*, **235**, 2387 (1960).

(5) J. J. Fox, N. Miller, and I. Wempen, *J. Med. Chem.*, **9**, 101 (1966).

(6) M. R. Dollinger, J. H. Burchenal, W. Krise, and J. J. Fox, *Biochem. Pharmacol.*, **16**, 689 (1967).

(7) G. W. Camiener, *ibid.*, in press.

(8) J. J. Fox, D. Van Praag, I. Wempen, I. L. Doerr, L. Cheong, J. E. Knoll, M. L. Eidinoff, A. Bendich, and G. B. Brown, *J. Am. Chem. Soc.*, **81**, 178 (1959).

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