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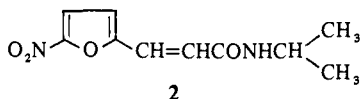
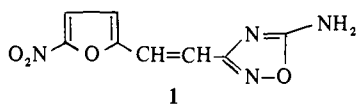
Structure and Antischistosomal Activity in the Nitrofurans Series. Requirement for a 5-Nitro Group in 3-(5-Nitro-2-furyl)acrylic Acid Derivatives†

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We have recently drawn attention^{1,2} to structural and conformational features associated with schistosomicidal action in nitrofurans derivatives. In particular, we observed that the absence of the 5-nitro grouping in *trans*-5-amino-3-[2-(5-nitro-2-furyl)vinyl]-1,2,3-oxadiazole (**1**) is associated with complete loss of antischistosomal activity. The question arises as to whether the nitro group in this and in related nitrofurans derivatives can be replaced by some other electronegative substituent without loss of antiparasitic activity. The precise role played by the nitro group is unknown, but possibilities include reduction by host or parasite to an unstable hydroxylamino or nitroso grouping.

We have attempted to answer the question by synthesizing and testing selected 5-carboxy-, 5-carboxymethyl-, and 5-cyano-2-furyl derivatives for comparison with a corresponding schistosomicidal 5-nitro-2-furyl compound. The similarity in both the type and sequence of biochemical effects and morphological changes induced by both the nitrofurans derivative **1** and by simple amides of 3-(5-nitro-2-furyl)acrylic acid (*e.g.*, **2**) has suggested^{1,2} a similar mode of action. For synthetic reasons we chose to work with the amide **2** rather than the more complex compound **1**, and carboxy, carboxymethyl, and cyano groups were selected for comparison with the nitro grouping for two reasons. Firstly, the Hammett normal substituent constants (*para*) for CO₂CH₃, CO₂H, and CN are³ respectively +0.38, +0.41, and +0.67 *vs.* 0.78 for NO₂. If the nitro group's contribution is based primarily on electron withdrawal, then one might expect a graded series of biological activities, with the activity of the cyanofuran derivative somewhat similar to the corresponding nitrofurans. Secondly, the conformational free



energies⁴ of these groupings suggest that their spatial requirements are similar to or less than that of the nitro group. In the course of the synthetic work we also prepared analogs of **2** in which the nitro group is replaced by CH₃, CH₂OH, and CH=NOH. These were also tested for antischistosomal activity.

Synthesis. The new *N*-isopropyl-3-(5-substituted 2-furyl)acrylamides synthesized in this study are collected in Table I. The known⁵ 3-(5-methyl-2-furyl)acrylic acid was converted to the isopropyl amide **3** *via* reaction with thionyl chloride to give acid chloride and then treatment with isopropylamine. Related acids were similarly converted to the corresponding isopropyl amide derivatives.

To prepare *N*-isopropyl-3-(5-cyano-2-furyl)acrylamide (**11**), we first oxidized *N*-isopropyl-3-(5-hydroxymethyl-2-furyl)acrylamide (**7**) to the 5-formyl derivative **8** with manganese dioxide. The mixture of *syn* and *anti* oximes, **9** and **10**, obtained from **8** was heated with acetic anhydride-sodium acetate to give the required 5-cyano compound **11**. Although the oximes **9** and **10** were separated and characterized, the crude oxime mixture was carried through successfully to the nitrile **11**. If oxime dehydration was effected by acetic anhydride alone, the nitrile **11** was accompanied by its *N*-acetyl derivative as well as by 3-(5-cyano-2-furyl)acrylic acid. The latter was presumably formed by hydrolytic cleavage of the *N*-acetyl compound. All compounds showed mass spectra and nmr spectra consistent with the postulated structures.

Pharmacological Results. Compounds **3-5**, **7**, **9**, **10**, and **11** were tested for antischistosomal activity in mice infected with *Schistosoma mansoni* and were found to be devoid of antiparasitic activity at dose levels (250 mg/kg *b.i.d.*, 10 doses) which caused significant reduction in the number of worms in the case of the parent 5-nitro compound **2**.

Lower doses of the latter and of other antischistosomal compounds of this group cause a reversible reduction of glycogen phosphorylase phosphatase activity and a glycogen depletion in the worms,^{1,2,6} thus providing a sensitive indicator for low antischistosomal activity. Since neither of these two biochemical changes was observed after administration of compounds **3-11**, they were devoid even of borderline activity.

Conclusions

Our experimental results demonstrate that replacement of the nitro group in *N*-isopropyl-3-(5-nitro-2-furyl)acrylamide (**2**) by cyano, carboxy, or carboxymethyl groupings results in complete loss of schistosomicidal activity. We infer that electronegativity alone cannot explain the crucial role played by the nitro group and suggest that *in vivo* reduction to a hydroxylamino or nitroso group may be involved.

Experimental Section

General Procedure for Preparation of *N*-Isopropyl-3-(5-substituted 2-furyl)acrylamides of Table I from the Corresponding Acids, Illustrated by the Synthesis of *N*-Isopropyl-3-(5-methyl-2-furyl)acrylamide (3**).** A solution of 3-(5-methyl-2-furyl)acrylic acid⁵ (3.04 g, 0.02 mol) in Et₂O (75 ml) and SOCl₂ (2.86 g, 0.024 mol) was heated under reflux for 1.5 hr. The solvents were evaporated *in vacuo*, and the crude acid chloride (3.40 g) was dissolved in Et₂O (30 ml) and cooled in a Dry Ice-acetone mixture. Isopropylamine (2.60 g, 0.044 mol) was added; the mixture was kept at -70° for 15 min and was then diluted with H₂O and extracted with CHCl₃. The CHCl₃ extract was washed with H₂O, dried (Na₂SO₄), and evaporated *in vacuo*. The residue was crystallized from CHCl₃-Et₂O to give 1.22 g (32%) of **3**, mp 117-120°, and a second crop of 1.18 g

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Table I

Compd no.	R ^a	Mp, °C	Crystn solvents	Formula
3	CH ₃	121-122	CHCl ₃ -Et ₂ O	C ₁₁ H ₁₅ NO ₂
4	COOCH ₃	164-165	C ₆ H ₆ -Et ₂ O	C ₁₂ H ₁₅ NO ₄
5	COOH	201-203	MeOH-Et ₂ O	C ₁₁ H ₁₃ NO ₄
6	CH ₂ OCOCH ₃	116-118	EtOAc-ligroin	C ₁₃ H ₁₇ NO ₄
7	CH ₂ OH	185-186	MeOH	C ₁₁ H ₁₅ NO ₃
8	CHO	157-159	Me ₂ CO-Et ₂ O	C ₁₁ H ₁₃ NO ₃
9	CH=NOH	186-191	MeOH-Et ₂ O	C ₁₁ H ₁₃ N ₂ O ₃
10	CH=NOH	142-146	Et ₂ O-C ₆ H ₆	C ₁₁ H ₁₄ N ₂ O ₃
11	CN	139-140	Et ₂ O-ligroin	C ₁₁ H ₁₂ N ₂ O ₂

^aAll compounds were analyzed for C, H, and N, and all gave values within 0.3% of the calculated figures.

(31%), mp 115-118°. Recrystallization raised the melting point to 121-122°.

N-Isopropyl-3-(5-carboxy-2-furyl)acrylamide (5). The known⁷ 3-(5-carboxymethyl-2-furyl)acrylic acid was converted to the isopropyl amide 4 by the general method described above, and 4 (0.52 g, 2.2 mmol) was dissolved in MeOH (10 ml) and 1% aqueous KOH (16.5 ml) and kept at 25° for 55 min. Acidification and extraction with EtOAc gave crude product (0.48 g) which was crystallized from MeOH-Et₂O to give 5, mp 201-203° (0.406 g, 83%).

N-Isopropyl-3-(5-hydroxymethyl-2-furyl)acrylamide (7). The known⁸ 3-(5-acetoxymethyl-2-furyl)acrylic acid was converted to the isopropyl amide 6 by the general method described above, and a solution of 6 (7.80 g, 0.031 mol) in 40 ml of MeOH and 21 ml of 10% aqueous KOH was kept at 25° for 30 min. The solution was acidified, and the precipitate was filtered off, washed with water, and dried *in vacuo*. Crystallization from MeOH-Et₂O gave 7 (4.75 g, 73%), mp 182-185°; second crop (1.20 g, 18%), mp 181-183°. Recrystallization from MeOH-Et₂O raised the melting point to 185-186°.

N-Isopropyl-3-(5-formyl-2-furyl)acrylamide (8). The hydroxymethyl compound 7 (1.5 g, 7.2 mmol) in Me₂CO (300 ml) was stirred with activated MnO₂ (5.0 g) at 25° for 2 hr. More MnO₂ (1.0 g) was added, and stirring was continued for 1 hr. Another portion (1.0 g) of MnO₂ was added and stirring was continued for 1.75 hr. The mixture was filtered and the filtrate was evaporated *in vacuo* to give crude aldehyde (1.38 g). Crystallization from Me₂CO-Et₂O gave 1.28 g (85%) of 8, mp 155-157°. Recrystallization raised the melting point to 157-159°.

Formation of Oximes 9 and 10 from *N*-Isopropyl-3-(5-formyl-2-furyl)acrylamide (8). A solution of the aldehyde 8 (3.41 g, 0.016 mol) in MeOH (15 ml) and H₂O (30 ml) containing hydroxylamine sulfate (2.08 g, 0.016 mol) and NaOAc (2.62 g, 0.032 mol) was heated under reflux for 1 hr. Tlc (EtOAc-ligroin, 4:1) showed formation of two products. Removal of MeOH *in vacuo* resulted in the crystallization of the less polar oxime 9 (2.25 g, 61%) which was recrystallized from MeOH-Et₂O to give 0.814 g (22%), mp 187-191°. The aqueous mother liquor was extracted with CHCl₃ to give 1.50 g (41%) comprising mainly the more polar oxime 10. Crystallization from Et₂O-C₆H₆ gave pure 10, mp 142-146°.

N-Isopropyl-3-(5-cyano-2-furyl)acrylamide (11). A solution of the more polar oxime 10 (1.22 g, 5.5 mmol) in Ac₂O (1.5 ml) containing NaOAc (0.675 g) was heated under reflux for 70 min. By dilution with water and extraction with Et₂O-CHCl₃ (4:1), crude nitrile (1.04 g) was obtained. Crystallization from C₆H₆-ligroin gave 0.706 g (63%) of 11, still slightly impure. Dry-column chromatography on silica gel (10 g) with CHCl₃ as eluent gave 0.629 g (56%) of 11, which was crystallized from Et₂O-ligroin to give pure material, mp 139-140°. The nitrile 11 could also be obtained in comparable amount if the crude oxime mixture, 9 and 10, was used in the reaction.

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Nucleoside Peptides. 5.¹ Synthesis of Certain *N*-(5-Uridineacetyl)amino Acids from 5-Uridineacetic Acid

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Modified nucleosides isolated from transfer ribonucleates have been proposed (for a recent review, see ref 2) to exhibit a regulatory influence on protein biosynthesis as well as influence the tertiary structure of these macromolecules. Inhibition of the growth of certain cancer cell lines by 4-thiouridine³ and the cytokinin activity of isopentenyl adenosine⁴ have been noted and suggest that other minor nucleosides and their derivatives may possess interesting biological activities.

It was, therefore, of interest to study the biological properties of the modified nucleoside 5-uridineacetic acid (1) (Scheme I) isolated from yeast transfer ribonucleates^{5,6} and several of its analogs. Recently 1 was synthesized^{7,8} and the synthetic product found to be identical with the product isolated from natural sources.⁸ Since we have investigated the biological properties of a number of nucleoside peptide derivatives and found inhibition of protein biosynthesis^{1,9} and antiviral activity¹⁰ in cell culture, we now report the synthesis and biological properties of a number of *N*-(5-uridineacetyl)amino acids prepared from 1.

The active ester procedure of Bodanszky¹¹ provided the method of choice for peptide bond formation. Since this technique allows selective bond formation to an amino group in the presence of free hydroxyl groups,⁹ it was unnecessary to block the sugar moiety of the nucleoside. A facile preparation of the active ester 5-uridineacetic acid *p*-nitrophenyl ester (2a) (Scheme I) was achieved by the action of *p*-nitrophenol and DCC on 5-uridineacetic acid (1). This useful intermediate was found to be readily susceptible to nucleophilic attack. Treatment of 2a with alcoholic ammonia or alcoholic methylamine gave 5-uridineacetamide (2b) and 5-uridine-*N*-methylacetamide (2c) in yields of 58 and 80%, respectively. Facile peptide bond formation was also achieved by treatment of 2a with appro-