

Palmitoyl chloride (49 g, 0.178 mol) was dissolved in 500 ml of ether and cooled to 0°, and 500 ml of 30% H₂O₂ was added dropwise. After the solution was stirred for 20 min, 50 ml of pyridine was added, and the resulting solution was stirred for an additional 30 min at room temperature. The ether layer was washed with 250 ml of 5% HCl, water, and 2% NaOH, dried over MgSO₄, and evaporated; yield 30 g (62%).

6-Hydroxy-4,7-dioxobenzothiazole (4.0 g, 0.022 mol) was dissolved in 500 ml of acetic acid. The temperature was raised to 90°, and palmitoyl peroxide (16 g, 0.031 mol) in 300 ml of ether was added over a 4-hr period. The solution was filtered to give 1.7 g of starting material. The filtrate was stirred overnight, and the solvent was evaporated. The remaining oil was placed on a silica gel column and eluted with ether. The yellow fractions were combined, and the solvent was removed. Two recrystallizations from hexane-ether gave 0.32 g of orange solid (6.5%); mp 129–130°; nmr (CDCl₃) δ 0.90, 1.26 (m, 29 H), 2.61 (t, 2 H), 9.16 (s, 1 H). *Anal.* C₂₂H₃₃N₂O₃S) C, H, N.

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Acute Oral Toxicity of 2-Alkyl- and 2,6-Dialkylanilines. Correlation with Lipophilicity

John A. Durden, Jr.

Research and Development Department, Chemicals and Plastics Division, Union Carbide Corporation, South Charleston, West Virginia 25303. Received May 24, 1973

Jacobson¹ has recently presented data pertaining to the acute rat oral toxicity of a group of 2-alkyl- and 2,6-dialkylaniline derivatives. Also presented was a very brief discussion of the structure-activity relationships which may be operative in this series.

be noted that the correlation between *P* and *E_s* for these compounds is quite high (*r* = 0.894); about 80% of the variation in *P* is accounted for by the variation in *E_s*. Although there may be steric effects influencing the activity of these compounds, the high correlation with *P* suggests that relative lipophilicity (*P*) is probably the primary toxicity determinative. If other than alkyl substituents were included in the series almost certainly electronic or steric effects would begin to play a role along with lipophilicity. The importance of *P* may reflect a lipophilicity-related *in vivo* distribution effect⁵ and/or a direct relationship between lipophilicity and *in vivo* degradation (metabolism, conjugation)⁶ for these materials.

Table I. Structural Parameters and Observed and Calculated Toxicity Values

No.	R	R ¹	Log <i>P</i>	<i>E_s</i> , R.R ¹	Rat peroral LD ₅₀ values, g/kg	
					Obsd	Calcd
1	H	H	0.90 ^a	2.48	0.44	0.46
2	H	CH ₃	1.30 ^a	1.24	0.90	0.68
3	CH ₃	CH ₃	1.70	0	0.84	1.0
4	H	C ₂ H ₅	1.80	1.07	1.26	1.06
5	C ₂ H ₅	C ₂ H ₅	2.70	-0.14	2.69	2.31
6	H	CH(CH ₃) ₂	2.10	0.75	1.18	1.43
7	CH(CH ₃) ₂	CH(CH ₃) ₂	3.30	-0.98	4.27	4.00
8	CH ₃	C ₂ H ₅	2.20	-0.07	1.18	1.52

^a From ref 2; the remainder was calculated by standard methods.

A high correlation (*r* = 0.926) has now been found between these toxicity data and the partition coefficients (*P*)² of the corresponding anilines. Regression of the acute oral data against log *P* provided a relationship which accounted for 85.9% of the observed variation in data. Inclusion of a parameter (*E_s*)^{3,4} related to the steric requirement of the ortho substituents(s) did not significantly change the accountability of eq 1. The correlation (*r* =

$$\log 1/C = 2.551 - 0.274 \log P \quad (1)$$

s = 0.0503; *MR*² = 0.8586; *F* = 36.45; *r* = 0.926 0.775, accountability 60.1%) between *E_s* and log 1/*C* was also significantly less than that observed with *P*. It should

The data employed in this study are recorded in Table I together with the calculated toxicities from eq 1. Log 1/*C* in eq 1 is calculated by eq 2.

$$\log 1/C = -\left[\log \frac{100 \times \text{LD}_{50}}{\text{mol wt}} \right] + 2 \quad (2)$$

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