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Communications to the Editor

Synthesis and Biological Effects of 13-Dehydro Derivatives of Natural Prostaglandin $F_{2\alpha}$ and E_2 and Their 15-Epi Enantiomers[¶]

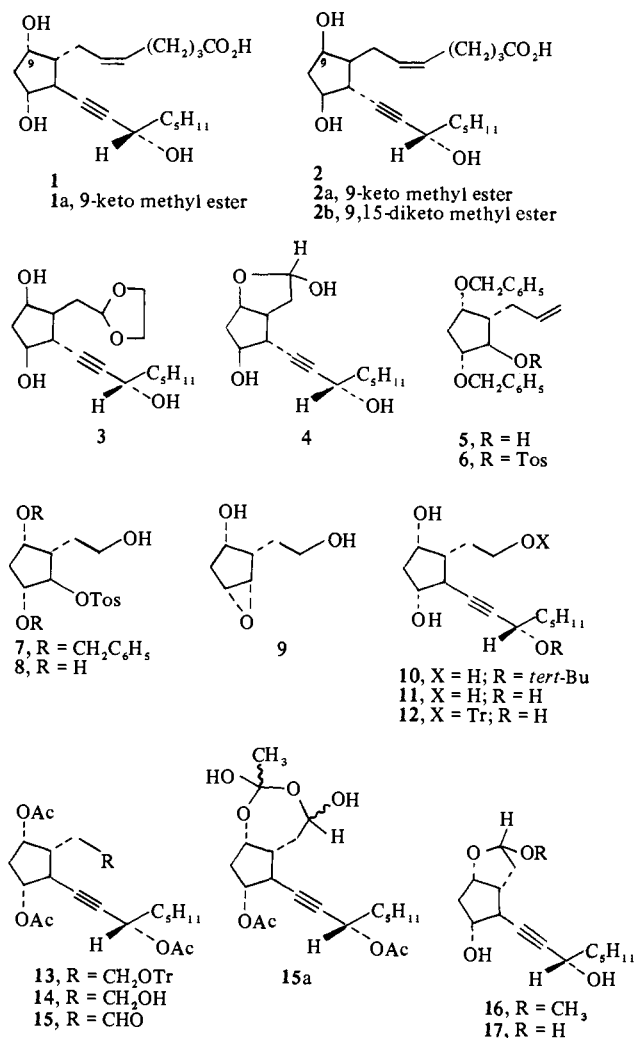
Sir:

In recent communications^{1,2} we have described synthetic routes leading to all the primary prostaglandins *via* acetylenic intermediates of type **3** and **11**. The convertibility of these same intermediates into acetylenic prostaglandins, *e.g.*, **1** and **2**, appeared an attractive synthetic goal, in view of the possibility that such substances containing a propargyl in place of the allylic alcohol moiety might turn out to be non-substrates, or even antagonists, for the highly specific prostaglandin 15-dehydrogenase^{3,4} and thus possess longevity of action not observed with the natural prostaglandins. Such blockage of dehydrogenation had previously been achieved by substituting a methyl group for the 15-hydrogen.⁵ We wish to report here the synthesis and biological properties in five test systems of *nat*-13-dehydroprostaglandin $F_{2\alpha}$ [†] (**1**) and *ent*-13-dehydro-15-epiprostaglandin $F_{2\alpha}$ [†] (**2**) and their corresponding E_2 methyl esters **1a** and **2a**. The data presented show that substitution of an acetylenic for the transolefinic function does indeed render these substances antagonists of the 15-dehydrogenase and, moreover, gives rise to remarkable changes in their activity profiles.

In planning the synthesis of **1** and **2** it was evident that one of the features of our synthetic route, namely, the resolution of the racemic cyclopentane moiety, *e.g.*, **9**, by introduction of the acetylenic 8-carbon side chain in optically active form^{1,2} as in **10** or **11** appeared to be precluded since chromatographic separation of the acetylenic diastereomers of type **3** and **11** could not be achieved. Fortunately, one of the diastereomers, **3**,[‡] could be obtained in crystalline form,[§] mp 102°, $[\alpha]_D -2.0^\circ$ (*c* 2.15),[#] and the synthesis of **2** completed by hydrolysis of the former with 0.1 *N* HCl in acetonitrile-water (2:1) at 25° for 24 hr to the hemiacetal **4**, $[\alpha]_D +1.4^\circ$ (*c* 2.60), followed by a Wittig reaction as previously described.¹ The resulting *ent*-13-dehydro-15-epiprostaglandin $F_{2\alpha}$ (**2**) obtained in 67% yield from **3**, after purification by high-pressure chromatography, had mp 22–23° and

$[\alpha]_{EtOH_D} -39.5^\circ$ (*c* 0.85).^{**} Conversion of the methyl ester of **2** into the corresponding PGE₂ derivative by selective 11,15-silylation with *N*-trimethylsilyldiethylamine in acetone at –40° followed by Collins oxidation⁶ furnished **2a**,^{**} $[\alpha]_{EtOH_D} -5.6^\circ$ (*c* 0.45), in 40% yield, together with the recovered methyl ester of **2** (18%) and the 9,15-diketo methyl ester **2b** (20%), λ_{max}^{alc} 222 nm (ϵ 9600).

Since the other diastereomer corresponding to **3** could not be obtained in pure crystalline form, it was necessary to carry out the synthesis of **1** with resolved (+)-**5**, and, of course, (3*S*)-3*t*-butyloxy-1-octynyldimethylalane,⁷ the resolution and determination of absolute configuration of



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[†]The prefixes *nat* and *ent* are employed here to indicate that all the chiral centers present in these prostaglandin analogs correspond to those in the natural prostaglandins (*nat*) or in their enantiomers (*ent*).

[‡]All structural formulas represent the absolute configurations shown.

[§]*Cf.* footnote 13 in ref 1. The purity of this substance was verified by LiAlH₄ reduction to the allylic alcohol, which proved to be a single diastereomer by tlc, a technique which readily distinguishes between such allylic 15-epimers.

[#]Rotations in chloroform at 28–30° unless indicated otherwise.

^{**}The ir and nmr spectra of this substance as well as the low-resolution mass spectrum of its trimethylsilyl ether methyl ester which showed the M⁺ peak were in accord with the assigned structure. Mass spectra were taken on a Finnigan 1015 quadrupole instrument equipped with gc inlet and interfaced with a Systems Industries computer system.

Table I. Biological Activities of 13-Dehydroprostaglandins

Compd	Gerbil colon contraction, ^{a,b} F _{2α} = 1	cAMP synthesis, mouse ovary ^{c,d}	Rat lipocyte binding ^{e,f}	Antifertility hamster, ^{g,h} F _{2α} = 1	Placental 15-dehydrogenase inhibition, ^k I ₅₀ , ^l μM
1	0.33 ± 0.15	1-2 × F _{2α}	0.24 × F _{2α} ± 0.07	2 ⁱ 5 ^j	47
1a		0.1-0.3 × E ₁	0.21 × E ₁ ± 0.07		7
2	0.0052 ± 0.0014	1-2 × F _{2α}	<0.04 × F _{2α}	0.25 ⁱ	50
2a		0.02-0.05 × E ₁	0.08 × E ₁ ± 0.03		20

^aReference 11. ^bPotencies and 95% CL calculated from dose-response curves for PGF_{2α} (six levels from 2 to 100 ng/ml) and test compounds. ^cReference 9. ^dPotency ranges derived by comparing per cent stimulation of adenylyl cyclase over control of test compounds at three dose levels, with maximum stimulation obtained with PGF_{2α} (100 μg/ml) and PGE₁ (1 μg/ml), respectively. Data for 1 and 2 from two independent experiments; those for 1a and 2a from one experiment. ^eReference 12. ^fPotencies and 95% CL obtained by comparing binding of test compounds at three dose levels with standard curves for PGE₁ and PGF_{2α}. ^gReference 10. ^hDerived by comparing "minimum effective doses" of test compounds with those for PGF_{2α}. The "minimum effective dose" is the minimum dose/animal/day that will result in no pregnancies in a group of five animals. For PGF_{2α} this dose is 12.5 μg, sc, and 1.0 mg, po. ⁱSubcutaneously. ^jPerorally. ^kReference 8. ^lConcentration producing 50% inhibition as measured by the ΔOD_{340 nm} after 15-min incubation at 25° of placental enzyme (0.01 ml), PGE₁ (20 μg), test substance (40, 60, 80 μg), and NAD (1 mg, added after 5 min) in 3 ml of phosphate buffer at pH 7.0.

which had already been achieved.¹ The synthetic sequence employed was identical up to (+)-11 with that previously described for *rac*-5,^{1,2} and the relevant structures are shown here for the purpose of presenting the specific rotation data: 6, [α]_D +16.6° (c 2.31); 7, +3.7° (c 2.14); 8, +27.0° (c 1.98); 9, +0.4° (c 1.0); 10, -33.4° (c 2.08); 11, +16.0° (c 2.85). Tritylation of 11 at 25° for 20 hr furnished in 80% yield the primary trityl ether 12, [α]_D -17.5° (c 2.70), which was acetylated quantitatively to 13, [α]_D -8.7° (c 2.50), and detritylated with 90% acetic acid at 25° for 24 hr to yield 14 (89%), [α]_D -43.5° (c 1.76). Collins oxidation of the latter for 10 min at 25° afforded the aldehyde 15,^{††} [α]_D -25.6° (c 1.16), which on treatment with 2% potassium carbonate in methanol at 25° for 5 hr gave rise to the methyl glycoside 16,^{‡‡} so characterized by nmr [broad singlets at δ 3.30 (OCH₃) and 5.10 (-CH(OR)₂)] and glc-ms of its bis(trimethylsilyl) ether, M⁺ 426. Hydrolysis of 16 with 0.1 N HCl in acetonitrile-water (2:1) at 25° for 48 hr gave the free hemiacetal 17, [α]_D -5.0° (c 0.42), in 63% yield from 14 after tlc. The synthesis was completed as described above for the enantiomeric 2 and 2a resulting in *nat*-13-dehydroprostaglandin F_{2α} (1),** [α]_D^{EtOH} +34.0° (c 0.66), as well as *nat*-13-dehydroprostaglandin E₂ methyl ester (1a).**,⁶

Bioassay data for 1, 1a, 2, and 2a are summarized in Table I. All four 13-dehydro derivatives proved to be nonsubstrates for the human placental 15-dehydrogenase⁵ as well as inhibitors of that highly specific prostaglandin-metabolizing enzyme. Furthermore, they all caused stimulation of cAMP synthesis in the mouse ovary.⁹ Most remarkable in this respect was the unexpectedly high activity of the enantiomeric substances 2 and 2a, in which all the asymmetric centers with the exception of C-15 are of unnatural chirality. Moreover, both chiral types, 1 and 2, caused termination of pregnancy in hamsters.¹⁰ Of prime interest is the highly significant dissociation in compound 2 of smooth muscle stimulating activity¹¹ and binding to a rat lipocyte "receptor" preparation specific for prostaglandins¹² from adenylyl cyclase stimulating and antifertility

activity. It is this latter fact which gives cause for optimism that the activity profiles of prostaglandins can be drastically altered by chemical modification.

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^{††}The triacetate aldehyde 15 exhibited aldehyde characteristics, methine proton at δ 9.78 (broad singlet), when the work-up excluded washing of the ether extract with bicarbonate. When a bicarbonate wash was performed the aldehyde methine proton was replaced by the hemiacetalic methine signal in the δ 5.3-5.7 region and the 9-proton was shifted upfield to form a multiplet centered at δ 3.75. We propose structure 15a for this intermediate. It is this bicarbonate washed material that was used in the subsequent step.

^{‡‡}This unusual reaction appears to involve the cyclic hemiacetal 15a, since the desacetyl hemiacetal 17 on treatment with potassium carbonate in methanol led to intractable products, probably by aldolization.